

Androgen-regulated cardiac metabolism in aging men

[Health & Medicine](#)



**ASSIGN
BUSTER**

Introduction

The multifactorial origin of cardiovascular diseases compels a comprehensive approach that incorporates lifestyle modification with an appropriate selection of medications for energy-regulation and its co-morbid conditions ([1](#) - [4](#)). In a physiological scenario, cardiometabolic adaptations involve a complex relationship among mechanisms responding to energy needs and substrate availability, in order to maintain homeostasis ([5](#) - [7](#)). During senescence, reduced ATP generation in the heart impairs normal contractile performance. There is a positive association between cardiac failure in age-related pathologies and insulin resistance, diabetes, sarcopenia and cardiovascular diseases ([8](#), [9](#)).

According to a 2019 update article from the American Heart Association, almost one in three adult men have some type of cardiovascular disease ([10](#)). Women are known to suffer cardiac disease 10-20 years later than men, which supports the hypothesis that physiological estrogen levels confer cardioprotective effects ([11](#) - [14](#)). In the past decades, the effect of sex-related steroid hormones on the cardiovascular system has been predominantly focused on estrogen actions, whereas research concerning the beneficial cardiac effects of androgens has been limited.

There is an extensive body of information indicating that administration of supraphysiologic doses of testosterone and cognate anabolic steroids induce adverse cardiovascular effects by triggering cardiac hypertrophy and heart failure ([15](#)). Although androgens have been considered previously to cause adverse cardiac outcomes, recent studies support favorable effects of these

hormones on cardiovascular homeostasis ([16](#) - [18](#)). Many clinical publications over the past few years have indicated that very low levels of plasma testosterone are associated with pathophysiological processes, such as dyslipidemias, metabolic syndrome and diabetes type 2, which are considered as the underlying mechanisms involved in age-related cardiovascular diseases in men ([19](#) - [23](#)). Low circulating testosterone levels, as found in late-onset hypogonadism and elderly men, have also been associated with different types of heart diseases ([24](#), [25](#)). Moreover, epidemiological reports show that decreased testosterone concentration is a predictor of mortality in senior men ([26](#)).

A recent report from the Mayo Clinic (2018) exhaustively reviewed and analyzed the main clinical publications over the past 10 years related to testosterone levels, testosterone administration and their impact on the cardiovascular system ([27](#)). Pharmacological replacement of testosterone prevents heart disease, improves exercise-induced myocardial ischemia, dilates the coronary arteries, and decreases insulin resistance ([28](#), [29](#)). The overall evidence indicates that physiological testosterone levels are beneficial for the male cardiovascular system, while low testosterone concentration is linked to unfavorable metabolic profile and increased cardiovascular risk ([27](#)).

Aging, at same time, is associated with a gradual decline of testosterone levels in men ([30](#)). Plasma levels of androgens fluctuate throughout life. During childhood and before puberty, testosterone concentrations are usually lower in males than females. After puberty, testosterone levels

increase in males, peaking at the age of 20–25. Thereafter, during aging, testosterone levels decrease ([31](#) - [33](#)). A cross-sectional study reported that in men over 40 years-old, total circulating testosterone levels decrease around 0.8% per year, while both free and albumin-bound testosterone levels decrease by 2%. In addition, plasma levels of sex hormone binding globulin (SHBG) increases by 1.6% per year, which may further decrease the bioavailable testosterone concentrations in elderly men ([30](#), [34](#)).

Circulating SHBG levels in humans are influenced by different factors, such as nutritional state, metabolism, hormonal factors and aging ([34](#) - [37](#)).

Testosterone is well-known for both its androgenic properties and its anabolic effects. This steroid hormone induces changes on organs and tissues promoting the adoption of the adult male phenotype ([38](#)). In the heart, testosterone associates key physiological input for metabolism and protein synthesis ([39](#)). Cardio-specific and concentration-dependent effects of testosterone are modulated by its circulating plasma levels, cellular metabolism, modulation of intracellular transduction pathways and androgen receptor expression ([15](#), [18](#)).

Age-related andropause is characterized by diminished plasma testosterone concentration in adult men. With the increasing aging of the world population, andropause is quickly becoming an epidemic condition associated with metabolic disorders and prominent cardiovascular risks ([32](#), [40](#), [41](#)). Decreased testosterone concentrations in older men are linked to changes in body composition, like increase of fat mass and reduction of lean body mass, dyslipidemia, insulin resistance, and reduced glucose

metabolism ([22](#)). The relationship between metabolic and cardiovascular risk in humans is evident in men suffering from hypogonadism, a condition in which the reduced functional activity of the gonads causes a decrease in testosterone levels ([42](#)). Hypogonadal men exhibit higher prevalence of cardiometabolic disorders compared to those with normal physiological levels of androgens ([43](#)). Retrospective studies of testosterone prescription databases have generated controversial and opposite results. Although testosterone replacement therapy to handle men hypogonadism it has been obtainable since 1939 ([44](#)), the apprehensions regarding the safety of testosterone treatment in men with cardiovascular diseases persist. However, to date, few systematic controlled studies have been performed to evaluate adverse events on cardiovascular system by testosterone administration ([45](#) - [47](#)). A recent randomized trial suggested that testosterone administration could increase cardiovascular risk in certain clinical populations, and it was suggested that pre-existing comorbidities as well as circulating lipid disturbances could influence the risk of cardiovascular events in older men. By contrast, several cross-sectional studies have demonstrated higher prevalence of cardiovascular diseases among men with low testosterone levels, and that replacement reduces cardiovascular risk ([48](#) , [49](#)). Likewise, subjects with low plasma testosterone concentrations are more prone to develop insulin resistance and diabetes, as well as central obesity and heart failure ([21](#) , [50](#) , [51](#)).

Similar responses have been observed in elderly men exhibiting diminished testosterone concentrations, which result in hormonal and metabolic alterations associated with increased risk for developing cardiomyopathies (<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[52](#)). Androgen supplementation is a focus of emerging interest for the treatment of age-related metabolic diseases and muscle wasting ([29](#), [53](#)). Accordingly, male sex steroids can regulate cardiometabolic functionality and energy production through transcriptional and post-transcriptional mechanisms, and therefore, can offer insights into energy spreading pathways and their mechanistic control during aging ([54](#)). The mechanisms by which testosterone contributes to beneficial metabolic actions on the development of metabolic syndrome and diabetes type 2 are revised and discussed by Kelly and Jones and these effects seem to involve multiple targets of lipid and carbohydrate metabolism ([55](#)).

Additionally to testosterone reduction in elderly men, low concentrations of testosterone are also found in late-onset hypogonadism, reduction of testicular volume and malfunction of the androgen production machinery, systemic accelerated testosterone metabolism and expression of defective androgen receptors ([24](#), [25](#), [42](#), [56](#)). In skeletal muscle, physiological testosterone levels regulate a host of metabolic enzymes and transcription factors that regulate the expression of nuclear-encoded mitochondrial oxidative phosphorylation proteins ([57](#)). In the elderly, the ATP production machinery is less efficient, and this condition represents an energy dilemma. Metabolic unbalance during aging in men must be resolved by adjusting the energy substrates and the expression of metabolic genes ([58](#) - [64](#)). Age-related cardiac metabolic adaptations must regulate energy demands with fuel supply under switching nutrient conditions. The impact of testosterone administration to increase skeletal muscle mass is recognized, but its therapeutic use in aging men is still controversial and the underlying

mechanisms remain to be defined. Recent reports indicate that testosterone therapy increases the expression of fibroblast growth factor 2 (FGF2) and decreases myogenic regulatory factor 4 (MRF4) and myostatin in skeletal muscle from men suffering hypogonadotropic hypogonadism, suggesting that the expression of these proteins contribute to muscle growth after testosterone therapy ([65](#)).

Cellular Mechanisms of Testosterone Action

As it is well known, the hypothalamic-pituitary-gonadal axis modulates testosterone production. The hypothalamus produces and secretes gonadotrophin-releasing hormone (GnRH), which stimulates the pituitary to induce the pulsatile secretion of luteinizing hormone (LH), which then prompts the Leydig cells of the testes to produce testosterone ([66](#), [67](#)), which, in turn, exerts a negative feedback on GnRH and gonadotropin secretion. As age progresses, both the amount of Leydig cells and their ability to produce testosterone are reduced, contributing to lower circulating levels of androgens in elderly men ([22](#), [30](#)). However, other authors have argued that there is not a reduction of Leydig cell mass with aging, and that the main defect occurs in intracellular cell signaling and cholesterol transport ([68](#)). During obesity and aging, a raise in the activity of aromatase enzyme converts testosterone into estrogens in men, further reducing circulating plasma levels and the ability of testosterone to exert its appropriate metabolic actions ([40](#), [69](#)).

The main mechanism of action of testosterone involves direct binding to the intracellular androgen receptor ([70](#) - [72](#)), which is a member of the

nuclear/steroid receptor superfamily. These receptors are proteins capable of binding their ligands in the cytoplasm or nucleus, and directly activating gene transcription ([73](#), [74](#)). The androgen receptor is a 110 kDa protein with three major functional regions for transactivation, a DNA binding domain and a hormone binding domain ([75](#)). After ligand binding, intracellular receptors are translocated to the nucleus, where they dimerize and bind to androgen response elements (ARE) to regulate target genes ([74](#)). Once bound to the hormone, other regulatory proteins or transcriptional coactivators can bind to the testosterone-androgen receptor complex to stabilize the promoter, thus achieving differential effects of this hormone either in a concentration-dependent or tissue-specific manner ([76](#)).

Previously, we and others have reported that testosterone also activates non-transcriptional signal transduction pathways, like extracellular signal-regulated kinase (ERK), phosphoinositide-3-kinase-protein kinase B/Akt (PI3K-PKB/Akt) and Ca²⁺-calmodulin-dependent protein kinase II (CaMKII) ([77](#) - [80](#)). In cardiomyocytes, testosterone induces hypertrophy through activation of the mechanistic target of rapamycin complex 1 (mTORC1) pathway ([79](#)) and glucose uptake by AMP-activated protein kinase (AMPK) activation ([80](#)). Overall, these evidences suggest that the effects of testosterone involve activation of anabolic and catabolic pathways. Thus, integration of transcriptional and non-transcriptional signals supplies cooperative mechanisms to support energy production under metabolic demand in cardiomyocytes.

As was mentioned above, SHBG is a protein that binds and transports testosterone within the bloodstream and regulates its bioavailability and access to extravascular target tissues ([35](#), [81](#)). Following the “ free hormone hypothesis,” there is a proportion of testosterone bound to SHBG with high affinity, the rest corresponds to free testosterone which is either loosely bound to albumin, or unbound to proteins ([82](#), [83](#)). Free testosterone can cross the plasma membrane and it associates directly with androgen receptors; therefore, it is regarded as the bioavailable fraction, which is responsible for the biological activity of this hormone ([84](#)). SHBG levels have been negatively correlated with insulin levels ([85](#)), and in a meta-analysis that included cross-sectional and prospective observational studies, Brand et al. found an inverse relationship between total testosterone and free testosterone with SHBG levels, and metabolic syndrome ([86](#), [87](#)) raising the question about the role that intracellular androgen binding protein levels play in endocrine cellular physiology.

Effects of Testosterone on the Cardiovascular System

Testosterone influences the cardiovascular system by acting directly on cardiac cells, the vascular tree, and by regulating cholesterol levels ([88](#) – [90](#)). In particular, exogenous administration of supra-physiological testosterone concentrations has been reported to produce cardiac hypertrophy, ventricular remodeling, cardiac failure, and sudden cardiac death ([39](#), [91](#), [92](#)). In humans and experimental animal models, testosterone has been related with higher risk of coronary artery disease through negative effects on plasma lipid and lipoprotein profiles, which may induce thrombosis and dilated cardiomyopathy ([15](#), [39](#), [88](#) – [90](#), [93](#)). It has been suggested that

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

testosterone replacement therapy can increase blood viscosity and develop myocardial infarction, underscoring that in each individual patient with various comorbidities, one or more thrombosis mechanism/s may be playing an effect. However, a systematic review meta-analysis in men did not clearly show a significant association between testosterone use and higher risk of venous thromboembolism ([94](#)).

On the other hand, at normal physiological levels, androgen actions are necessary for a range of biological processes, including protein synthesis and cardiomyocyte metabolism. Androgens also induce other hemodynamic consequences, including vascular bed relaxation, thus reducing after-load and rapidly increasing cardiac contractility, which increases cardiac output ([95](#)). In humans, the effect of a 3-year testosterone administration did not increase atherosclerosis progression ([96](#)); however, another study showed that testosterone treatment of elderly men increased the volume of coronary artery plaques ([97](#)). The effects of androgen supplementation on plasma lipids depend on the dose, the route of administration and the subject population. In patients with congestive heart failure, testosterone would exert a beneficial role by improving functional capacity, cardiovascular parameters and quality of life ([98](#)). Interestingly, testosterone replacement therapy can reduce circulating levels of inflammatory mediators, including interleukin (IL)-1 β and tumor necrosis factor α (TNF- α), as well as total cholesterol in patients with simultaneous coronary artery disease and testosterone deficiency ([99](#), [100](#)). The possible health risks and benefits of long-term testosterone replacement on older men with andropause caused by reduced testosterone concentrations are unknown. An interesting

hypothesis has been postulated by Herring et al. suggesting that testosterone may simultaneously benefit and harm the cardiovascular system by different pathways ([101](#)). Caminiti et al. ([102](#)) reported that in elderly patients with congestive heart failure, testosterone replacement therapy improves functional capacity in, large-muscle strength, and glucose handling. The improvement of functional capacity and muscular strength are correlated with the higher plasma testosterone levels ([102](#)).

Hypertension is a risk factor for developing cardiovascular diseases. In adult men, hypertension is more frequent and occurs earlier than in women of similar age ([103](#) - [105](#)). In men, blood pressure rise has been associated with the effects and differences of sex-related steroid hormones. The different ranges in blood pressure in men, compared to women, remain until 60 years of age. Various epidemiological studies have reported that in men under 60 years old the systolic blood pressure is 6-7 mm Hg higher than in women, while diastolic pressure is higher by 3-5 mm Hg ([106](#)). On the other hand, in women over 60 years of age, blood pressure gradually increases, reaching a similar prevalence than in elderly men. The reduction of estrogens and the change in the estrogen/androgen ratio seems to be relevant for the increase in blood pressure in postmenopausal women ([107](#) , [108](#)). An inverse relationship between systolic pressure and plasma testosterone levels has been reported in men, and an increased prevalence of hypertension in men with decreased free circulating androgens ([104](#)). The positive results of testosterone replacement are well documented. In randomized, double-blind, case-control clinical studies, the administration of hormones was associated with reduction of vascular tone ([109](#)). A

beneficial role of testosterone was found in patients with congestive heart failure, by improving functional capacity, cardiovascular parameters, and quality of life ([98](#)). Several reports have suggested that testosterone vasodilatory action is mediated by the smooth muscle cell through ion channel modulation, modulating either potassium channel opening and/or calcium channel inactivation ([110](#)).

Androgen Activates Intracellular Players Related to Cardiac Metabolism

The heart demands a continuous supply of energy to maintain muscle excitation-contraction coupling, and other intracellular adaptations, including fine-tuning in the expression of genes, ion homeostasis, signaling pathways, energetic balance and survival signals ([58](#) , [63](#) , [111](#)). Under normal conditions, cardiomyocytes promptly and effectively decode metabolic signals to evoke intracellular settings that improve cardiac functions to maintain an adequate energy balance that preserves work output and efficiency of the heart ([112](#) , [113](#)). In the fetal period, glucose is the main energetic substrate for ATP generation in the heart, switching to fatty acid in adults to adjust to increased energy demands ([63](#)). Thus, in adult cardiomyocytes, under normal conditions, ATP is mostly produced by fatty acid β -oxidation. Glucose represents another substrate metabolized by glycolysis. Fatty acids are transported into the mitochondria by the enzyme carnitine palmitoyl transferase 1 (CPT-1). Glycolysis requires glucose uptake, which occurs in cardiac cells through glucose transporter 1 (GLUT1) and GLUT4 ([113](#)). Inside the cell, glucose can be phosphorylated by hexokinase and further metabolized to pyruvate. Both, β -oxidation and glycolysis produce acetyl-

CoA to generate NADH and FADH₂ via the citric acid cycle. These metabolites are later used by mitochondria to generate ATP through the electron transport chain. Aerobic respiration pathways by oxidative phosphorylation, produces up to 60% of their energy from fatty acid and triglyceride metabolism, 35% from carbohydrate metabolism, and 5% from amino acid metabolism. These metabolic pathways are regulated through substrate/product ratio, rate of enzyme action and gene expression of metabolic enzymes and transporters ([9](#), [113](#)).

Preference in energy substrate utilization may change in response to substrate availability or metabolic deregulation in cardiomyocytes ([9](#), [113](#), [114](#)). Under testosterone stimulation, the heart experiences a series of adaptive processes that enable acute metabolic changes for functional demands. If demand for increased effort is repeated or continuous, structural and metabolic changes occur ([115](#)). Dynamic adjustments of energy-generating machinery under either low- or high-testosterone inputs compel critical adaptive responses from cardiomyocytes to maintain work output and efficiency of the heart ([63](#), [116](#), [117](#)). Disturbed feedback between energy requirements and production impairs mitochondrial function and energetic efficiency of cardiomyocytes ([118](#) - [120](#)).

Testosterone Improves Mitochondrial Function in Cardiac Cells

Transcriptional control of mitochondrial energy-generating machinery involves coordinated expression of proteins from two distinct genomes. Due to the limited coding capacity of mitochondrial DNA, nuclear encoded genes are also required ([121](#)). Mitochondrial enzymes are regulated through allosteric, post-translational, and transcriptional modifications ([122](#)).
<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

Testosterone regulates the expression of mitochondrial genes encoded by the nuclear genome and also, through direct action on mitochondria ([123](#) - [125](#)). Thus, by regulating cytosolic and mitochondrial pathways, testosterone exerts metabolic functions, with a possible feedback system between energy-producing mechanisms and cardiometabolic actions of testosterone in cardiomyocytes. Previous studies have shown that testosterone enhances the expression of mitochondria-encoded subunits of the respiratory chain, modulating mitochondrial respiratory function promoting functional efficiency ([57](#), [126](#)). In addition, androgens have direct interactions with respiratory chain complexes ([123](#)). In skeletal muscle cells, overexpression of androgen receptors increases mitochondrial enzyme activities and oxygen consumption ([127](#)). Following orchiectomy, young male mice show a decrease in the expression of genes associated with energy metabolism and oxidative phosphorylation, a phenotype that was reversed by testosterone treatment ([61](#)). With advancing age, androgen levels decrease and cardiac cells exhibit less mitochondrial number and lower energy production efficiency ([57](#)).

Androgen receptor signaling controls the transcription of several metabolic genes by engaging nuclear coactivator and corepressor proteins ([128](#)). In cardiac cells, peroxisome proliferator-activated receptor γ co-activator 1 α (PGC-1 α) stimulates mitochondrial biogenesis ([129](#)). PGC-1 α is associated with cardiac energy metabolism through its upstream regulators and downstream targets ([130](#)) and it is highly expressed in the heart ([131](#)). The PGC-1 α N-terminal domain interacts with proteins containing histone acetyltransferase activity, which allows remodeling of chromatin structure

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

and transcriptional activation ([132](#)). Adjacent to the N-terminus, PGC-1 α contains a regulatory domain with a LXXLL motif that interacts with nuclear receptors ([133](#), [134](#)). The PGC-1 α C-terminus recruits proteins that facilitate its interaction with the transcription initiation machinery ([135](#)). Moreover, PGC-1 α also regulates cardiac metabolism coactivating several transcription factor partners, including the androgen receptor ([129](#)). Also, testosterone up-regulates transcription of the nuclear respiratory factor-1 (NRF1), which controls the expression of mitochondrial respiratory chain complex proteins ([136](#)). NRF1 promoter contains putative ARE motifs in the DNA capable of binding the androgen receptor ([125](#)). It has been proposed that testosterone has a key modulatory role over NRFs and PGC-1 α modulating mitochondrial biogenesis and metabolism ([137](#)). Moreover, androgens induce transcriptional and posttranslational regulation of Drp1, a key protein in the mitochondrial fission machinery ([57](#), [123](#), [138](#)). In contrast, low testosterone levels are associated with reduced expression of mitochondrial respiratory genes ([126](#)). In young male mice orchietomy reduces the expression of genes associated with energy metabolism, oxidative phosphorylation, and ubiquinone pathways ([139](#)). Androgen receptor overexpression in cardiomyocytes increases mitochondrial enzyme activities and oxygen consumption ([139](#)). Testosterone administration, together with low-intensity physical exercise, increases mitochondrial biogenesis, increasing mitochondrial quality, and enhancing spontaneous physical activity, respiration and muscle mass ([70](#)). Therefore, the expression of metabolic genes related to testosterone may represent an

important therapeutic modality to prevent or treat age- and gender-related cardiac diseases.

AMPK and Cardiac Metabolism

AMPK is a serine/threonine kinase considered a fundamental intracellular energy sensor that regulates cell metabolism ([6](#)). AMPK is activated in response to physiological or pathological stimuli that reduce cell energy levels, by sensing the AMP/ATP ratio ([140](#)). AMPK modulates the activity of acetyl-coenzyme carboxylase, which in turn affects the levels of malonyl-coenzyme A, which is a key cellular energy regulator. AMPK coordinates metabolic pathways by limiting ATP expenditure and promoting ATP production to adjust to energy demands. In general, AMPK stimulates catabolic processes ([141](#)). Thus, AMPK promotes: (1) fatty acid β -oxidation, increasing their input to mitochondria and by activating enzymes such as carnitine palmitoyltransferase-1; (2) Glycolysis, increasing glucose uptake by GLUT4 and activating enzymes such as phosphofructokinase-2. Furthermore, activated AMPK can deliver energy status information through transcription factors to regulate gene expression of key proteins related to energy producing routes ([142](#)). A recent report has indicated that intramuscular injections of testosterone increase the expression and phosphorylation of AMPK α in adipose tissue and skeletal muscle biopsies of hypogonadism patients; these findings suggest that testosterone therapy may improve insulin sensitivity in obesity-associated hypogonadotropic hypogonadism men ([143](#)).

It has been well accepted that AMPK is cardioprotective ([6](#)). AMPK

deficiency exacerbates cardiac necrosis and apoptosis following ischemic-
<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

reperfusion injury in transgenic mice expressing a dominant negative form of AMPK. Furthermore, the hearts of these mice show loss of contractile force and low ATP levels, suggesting that AMPK plays a crucial role in cardiac function ([144](#), [145](#)). Additionally, AMPK activation with AICAR blocks cardiac hypertrophy induced by several pro-hypertrophic stimuli, mainly by its inhibitory effect on the mTORC1 pathway ([146](#)). Metabolism during compensated cardiomyocyte growth implicates that anabolic processes are associated with controlled catabolic processes. In a prior work, we reported that stimulation of cardiomyocytes with testosterone during a short-time (<15 min) increases AMPK phosphorylation through CaMKII in a concentration- and time-dependent manner ([80](#)). Once AMPK is activated, GLUT4 translocation to the plasma membrane increases, thus increasing glucose uptake ([147](#)). Therefore, increased glucose uptake and utilization may be an adaptive response, because ATP production from glucose consumes less oxygen than that from fatty acids.

Integrated Metabolic Actions of Testosterone and the AMPK/PGC-1 α Axis in Cardiomyocytes

Metabolic information obtained through cytosolic energy sensors must be decoded by specific downstream metabolic pathways to improve energy production capacity. Moreover, PGC-1 α interacts physically and functionally with well-known transcription factors involved in cardiomyocyte metabolism and growth ([135](#)). In fact, proximal PGC-1 α promoter has different putative DNA binding sites to bind transcription factors involved in re-expression of gene programs during cardiac metabolism and cardiomyocyte growth, such as GATA4 and Myocyte-enhancer factor 2 (MEF2). Mutations in these

transcription factors affect PGC-1 α promoter activity ([148](#), [149](#)). Some authors have reported that MEF2C and histone deacetylase 5 (HDAC5) have both, positive and negative modulation of PGC-1 α expression ([135](#)). Thus, PGC-1 α represents a metabolic regulator by modulating gene expression and cell growth, suggesting that the activation of the AMPK-PGC-1 α pathway is critical for the metabolic actions of androgens in the heart.

PGC-1 α is activated by exposure to cold, fasting, exercise and various stimuli that promote oxidative metabolism ([150](#), [151](#)). Signaling pathways associated with these stimuli include p38 MAP kinase, β -adrenergic/cAMP, nitric oxide, AMPK, and CaMKII. These diverse pathways modulate PGC-1 α activity by increasing PGC-1 α expression, nuclear transactivation and its downstream regulated genes ([141](#), [152](#), [153](#)). In the heart, PGC-1 α expression increases sharply at birth, coincident with a perinatal shift from glucose metabolism to fat oxidation ([154](#)). Different reports have indicated that low PGC-1 α expression correlates with pathological energy mechanisms and heart failure ([153](#), [155](#)). In young or ovariectomized animals models, sex steroids control mitochondrial energy production modulating the transcriptional and post-transcriptional machinery ([123](#)).

In the heart of neonatal mice, overexpression of PGC-1 α increases total mitochondrial mass ([130](#), [156](#)). In contrast, in adult mouse hearts, PGC-1 α overexpression results in modest mitochondrial biogenesis, followed by cardiomyopathy associated with mitochondrial abnormalities ([157](#)). PGC-1 α , together with PPAR α , coactivates the enhancement of genes involved in the fatty acid β -oxidation pathway ([116](#), [133](#), [158](#)). Conversely, PGC-1 α

induces GLUT4 expression in skeletal muscle, resulting in increased glucose uptake, which, in turn, significantly reduces plasma glucose levels ([159](#)). Furthermore, normal mitochondria biogenesis is activated in response to changes in the ATP/ADP ratio and subsequent AMPK activation, which increases PGC-1 α expression ([156](#), [160](#)). AMPK activation by AICAR increases β -oxidation of fatty acids by direct action on β -oxidation enzymes and by PGC-1 α and PPARs activation. Additionally, in response to chronic energy deprivation, mitochondrial biogenesis is dependent on AMPK ([6](#), [141](#)). In prostate cancer cells, testosterone promotes cell growth in an AMPK-dependent pathway, which allows metabolic changes by increasing PGC-1 α -dependent mitochondrial biogenesis ([141](#), [161](#)). In mice, treatment with testosterone increases PGC1 α expression levels ([136](#)), while low levels of testosterone are associated with reduced expression of PGC-1 α ([125](#), [162](#)). Furthermore, androgen receptor-deficient mice express low levels of PGC-1 α ([162](#)).

Effect of Sirtuins on Cardiac Metabolism

Protein acetylation/deacetylation play central roles in modulating cellular machinery related to metabolism ([163](#)). Mitochondria-mediated energy pathways contain acetylated proteins implicated in the tricarboxylic acid cycle, oxidative phosphorylation, fatty acid β -oxidation and glucose metabolism ([163](#)). In the heart, the protein sirtuin 3 (SIRT3) is a key regulator of mitochondrial function that adjusts energy availability, fuel sources and metabolic enzymes ([164](#)). Abnormal function of SIRT3 in pathophysiological processes is considered as the underlying mechanism of cardiovascular diseases ([165](#) – [168](#)). In cardiac cells SIRT3 is a stress-

responsive deacetylase that protects these cells from damage induced by genotoxic and oxidative stress-mediated agents. It has been shown that the increased expression of SIRT3 protects murine cardiomyocytes from genotoxic and oxidative stress-mediated cell death ([169](#), [170](#)). Current evidence associates impaired SIRT3 activity with higher risk of aging-associated illnesses like cardiovascular disease ([164](#), [171](#), [172](#)). Therefore, altered expression of SIRT3 may be the consequence of impaired upstream metabolic signaling that influences PGC-1 α activity, including AMPK and SIRT1 ([129](#), [154](#)). SIRT3 KO mice show cardiac mitochondrial function impairment and signs of premature aging ([173](#)). In addition, mice display contractile defects, such as a decrease of cardiac power, cardiac output, and developed pressure ([171](#), [174](#)). Porter et al. reported that decreased SIRT3 levels might raise the sensitivity of both heart cells and adult cardiac muscle to ischemia-reperfusion injury. This might contribute to a higher level of ischemia-reperfusion damage in the aged heart ([175](#)). Moreover, testosterone antagonizes doxorubicin-induced senescence of cardiomyocytes ([176](#)).

AMPK/PGC-1 α interaction is critical for the up-regulation of mitochondrial function and SIRT3 activity ([177](#), [178](#)). SIRT3 can also deacetylate and activate liver kinase B1 (LKB1) that, on its own, increases the activity of AMPK. NAD⁺ is considered an inhibitor of cardiac hypertrophic signaling pathways and it is regulated to prevent cardiac hypertrophy and heart failure ([6](#), [140](#)). Interestingly, disruption of the CD38 gene in male mice enhances cardiac function by elevating serum testosterone levels and producing a general increase in NAD⁺ tissue concentration ([179](#)). A key metabolic

regulator is AMPK, which controls mitochondrial homeostasis and metabolism by acting as an energy sensor ([150](#), [159](#), [180](#)). Moreover, the cytosolic deacetylase SIRT1 activates PGC-1 α in cardiomyocytes to increase transcriptional activity and mitochondrial biogenesis ([181](#)). In the nucleus, androgen receptor signaling stimulates PGC-1 α to increase the expression of various nuclear-encoded mitochondrial genes, including oxidative phosphorylation genes ([137](#)). SIRT3 is an important regulator of energy homeostasis and basal production of ATP. The heart expresses high levels of SIRT3, leading to a marked reduction of ATP in its absence ([182](#)). However, SIRT3 can boost ATP levels in mitochondria due to the acetylation process, which diminishes with age ([178](#)). Aging-induced tissue fibrosis is mediated by Glycogen Synthase Kinase 3 β (GSK3 β) ([183](#)). Therefore, deacetylation of GSK3 β by SIRT3 might reduce the tissue fibrosis associated to aging ([165](#)). Moreover, mitochondrial DNA content and activity, protein synthesis, oxidative capacity and ATP production are impaired by oxidative stress and free radicals. Regulated ROS production mediates redox signaling of transcription factors involved in mitochondrial biogenesis. However, an excess in the generation of mitochondrial ROS promotes oxidative stress that causes dysfunction and reduces mitochondrial biogenesis. Interestingly, SIRT3 reduces cardiac hypertrophy through increasing Foxo3a-dependent antioxidant defense mechanisms, suggesting that SIRT3 is an endogenous negative regulator of cardiac hypertrophy that protects the heart by suppressing cellular levels of ROS in mice ([165](#)). Thus, age-induced oxidative stress could be the underlying process that impairs mitochondrial

biogenesis and downregulation of genes required for mitochondrial function and biogenesis induced by testosterone in cardiac cells.

A decline in cardiometabolic adaptations possibly reflects several age-associated changes, including a decrease in circulating testosterone levels ([184](#)). Thus, prevention of androgen deficiency might improve cardiovascular outcomes and extend longevity. Because cardiomyocytes must meet energy demands with fuel supply under switching nutrient conditions, the responses to androgen signaling in the elderly would not be able to produce enough ATP for anabolic effects, resulting in reduced energetic efficiency in cardiomyocytes. As was mentioned above, despite that testosterone controls gene-expression programs related to energy metabolism—a crucial requisite for the induction of energy-producing mechanisms in mitochondria—there is limited information about the signaling pathways interlinking metabolism and growth mediated by changes in circulating plasma testosterone levels and their effect on cardiometabolic homeostasis.

Testosterone Metabolites in Aging

Testosterone can be transformed by the enzyme 5 α reductase to 5 α -dihydrotestosterone (DHT) mainly in skin, liver, hair follicles and prostate, where it acts locally ([74](#)). DHT is considered one of the main endogenous androgens ([185](#)). DHT binds to androgen receptors and induces the transcription of gene targets like testosterone. However, the dissociation constant of DHT-androgen receptor complex is 2–5 times lower than testosterone adduct, while DHT has a 10-fold higher potency on the signaling, which means that the effects of DHT and testosterone are different, but complementary ([75](#)).

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

Some reports suggest that DHT induces cardiac hypertrophy in cultured rat cardiomyocytes ([186](#), [187](#)) and in a rat model ([188](#)). On the other hand, treatment with finasteride, which inhibits the transformation of testosterone to DHT, reduces both cardiac hypertrophy and remodeling ([187](#), [189](#)).

Evidence has indicated that the conversion of testosterone to DHT is required for mediating some of the effects of androgen on the cardiovascular system. In patients with mutations in type 2 5 α reductase enzyme or finasteride treatment, the DHT levels are lower than healthy men but the androgenic phenotype is preserved. Nonetheless, these patients still show significant levels of circulating DHT. These results suggest that the conversion of testosterone to DHT is not essential for mediating its effects on muscle mass and strength ([190](#)). However, other studies have indicated that DHT may be an important risk predictor for cardiovascular disease in aging men. Healthy androgen levels are associated to survival and the total mortality of senior men displaying midrange concentrations of T and DHT is lower than men with low androgen levels, whereas those with higher DHT levels have shown lower ischemic heart disease mortality ([191](#)).

In men, estrogen levels increase during aging ([192](#)). Testosterone is converted to estradiol by the aromatase enzyme ([193](#), [194](#)), which is mainly expressed in adipose tissue ([195](#)). However, other factors also increase circulating estrogen levels, including impaired liver function, zinc deficiency, obesity, excessive use of alcohol and, environmental estrogens. Furthermore, estrogen levels are increased in men by various medications, such as statins and some blood pressure medications, antidepressants, and nonsteroidal anti-inflammatory drugs. In the case of obesity, aromatase

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

activity increases estrogen levels and reduces testosterone levels ([196](#)). In turn, the generated estradiol exerts a negative feedback effect on LH secretion, further reducing plasma testosterone concentrations ([192](#)). In healthy men, pharmacological inhibition of aromatase reduces insulin sensitivity. Furthermore, patients with CYP19 aromatase mutations display reduced muscle and fat mass, and suffer insulin resistance ([69](#), [196](#)). Experimental gene selection data suggest that aromatization of testosterone to estradiol may be important in mediating the effects of androgens on body composition. The effects of testosterone on lean mass, muscle size, and strength were not reduced when its conversion to estradiol was inhibited by the treatment with aromatase inhibitors. However, the effects of testosterone on fat mass and sexual desire seemed to be mediated by estradiol ([197](#)). These results suggest that the different effects of sex hormones are complex and dependent on the relative levels of testosterone, DHT and estradiol, factors associated with health in elderly men. More studies are required to evaluate the mechanism by which androgens might influence the cardiovascular system in older men, in order to determine the risks and benefits of clinical intervention.

Given the important roles of androgens in normal physiology of men, abnormal low levels must be considered as one of the main causes implicated in several disorders and pathological conditions in aging men. In the context of human disease relevance, androgen deficiency treated with testosterone prescriptions at physiological concentrations has been associated with lower cardiometabolic risk and treatment outcomes. In 2015 the international expert consensus panel suggested that we need more

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

research regarding the cardioprotective benefits of testosterone replacement, implying that there is enough evidence about the safety of testosterone therapy in hypogonadal and aging men and that the future research should be to study the suitable therapeutic options for age-related cardiovascular diseases ([198](#)).

Conclusion and Future Research

Age-related cardiometabolic actions of testosterone are tightly regulated by its circulating plasma concentrations. This is an essential aspect regarding male physiology, since testosterone levels decline in older men, concomitantly increasing metabolic- and gender-related cardiovascular diseases. Further research on cardiometabolic testosterone effects are required to determine their effective cardiac properties. By applying controlled, randomized studies, working to attain physiological testosterone concentrations, we will obtain new data to understand the role of testosterone as a metabolic modulator that can improve ATP production, and, in parallel, increase cardiac performance. These further studies on the divergent energy-controlling mechanisms that mediate testosterone effects and testosterone-related metabolic gene expression, may represent an important therapeutic modality for preventing or treating gender-related cardiac diseases.

Author Contributions

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

Funding

This work was supported by Fondo Nacional de Ciencia y Tecnología (FONDECYT) Grant 1151118 (to ME) and 1190406 (to PL).

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Cruz-Jentoft AJ, Baeyens JP, Bauer JM, Boirie Y, Cederholm T, Landi F, et al. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age Ageing* . (2010) 39: 412–23. doi: 10. 1093/ageing/afq034

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

2. Appelman Y, van Rijn BB, Ten Haaf ME, Boersma E, Peters SA. Sex differences in cardiovascular risk factors and disease prevention. *Atherosclerosis* . (2015) 241: 211–8. doi: 10. 1016/j. atherosclerosis. 2015. 01. 027

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

3. Bianchi VE. Impact of nutrition on cardiovascularfunction. *Curr Probl Cardiol* . (2018) 45: 100391. doi: 10. 1016/j. cpcardiol. 2018. 08. 003

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

4. Csige I, Ujvarosy D, Szabo Z, Lorincz I, Paragh G, Harangi M, et al. The impact of obesity on the cardiovascular system. *J Diabetes Res* . (2018) 2018: 3407306. doi: 10. 1155/2018/3407306

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

5. Allard MF, Parsons HL, Saeedi R, Wambolt RB, Brownsey R. AMPK and metabolic adaptation by the heart to pressure overload. *Am J Physiol Heart Circ Physiol* . (2007) 292: H140–8. doi: 10. 1152/ajpheart. 00424. 2006

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

6. Arad M, Seidman CE, Seidman JG. AMP-activated protein kinase in the heart: role during health and disease. *Circ Res* . (2007) 100: 474–88. doi: 10. 1161/01. RES. 0000258446. 23525. 37

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

7. Bernardo BC, Weeks KL, Pretorius L, McMullen JR. Molecular distinction between physiological and pathological cardiac hypertrophy: experimental findings and therapeutic strategies. *Pharmacol Ther* . (2010) 128: 191–227. doi: 10. 1016/j. pharmthera. 2010. 04. 005

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

8. Chimenti C, Kajstura J, Torella D, Urbanek K, Heleniak H, Colussi C, et al. Senescence and death of primitive cells and myocytes lead to premature cardiac aging and heart failure. *Circ Res* . (2003) 93: 604–13. doi: 10. 1161/01. RES. 0000093985. 76901. AF

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

9. Stanley WC, Recchia FA, Lopaschuk GD. Myocardial substrate metabolism in the normal and failing heart. *Physiol Rev* . (2005) 85: 1093–129. doi: 10.1152/physrev.00006.2004

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

10. Benjamin EJ, Muntner P, Alonso A, Bittencourt MS, Callaway CW, Carson AP, et al. Heart disease and stroke statistics-2019 update: A Report From the American Heart Association. *Circulation* . (2019) 139: e56–28. doi: 10.1161/CIR.0000000000000659

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

11. Malhotra A, Buttrick P, Scheuer J. Effects of sex hormones on development of physiological and pathological cardiac hypertrophy in male and female rats. *Am J Physiol*. (1990) 259(3 Pt 2): H866–71. doi: 10.1152/ajpheart.1990.259.3.H866

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

12. Weinberg EO, Thienelt CD, Katz SE, Bartunek J, Tajima M, Rohrbach S, et al. Gender differences in molecular remodeling in pressure overload hypertrophy. *J Am Coll Cardiol* . (1999) 34: 264–73. doi: 10.1016/S0735-1097(99)00165-5

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

13. Wallen WJ, Cserti C, Belanger MP, Wittnich C. Gender-differences in myocardial adaptation to afterload in normotensive and hypertensive rats. *Hypertension* . (2000) 36: 774–9. doi: 10. 1161/01. HYP. 36. 5. 774

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

14. Odening KE, Deiss S, Dilling-Boer D, Didenko M, Eriksson U, Nedios S, et al. Mechanisms of sex differences in atrial fibrillation: role of hormones and differences in electrophysiology, structure, function, and remodelling. *Europace* . (2019) 21: 366–76. doi: 10. 1093/europace/euy215

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

15. Liu PY, Death AK, Handelsman DJ. Androgens and cardiovascular disease. *Endocr Rev* . (2003) 24: 313–40. doi: 10. 1210/er. 2003-0005

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

16. Ikeda Y, Aihara K, Sato T, Akaike M, Yoshizumi M, Suzaki Y, et al. Androgen receptor gene knockout male mice exhibit impaired cardiac growth and exacerbation of angiotensin II-induced cardiac fibrosis. *J Biol Chem* . (2005) 280: 29661–6. doi: 10. 1074/jbc. M411694200

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

17. Ikeda Y, Aihara K, Yoshida S, Sato T, Yagi S, Iwase T, et al. Androgen-androgen receptor system protects against angiotensin II-induced vascular remodeling. *Endocrinology* . (2009) 150: 2857–64. doi: 10. 1210/en. 2008-1254

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

18. Ikeda Y, Aihara K, Yoshida S, Akaike M, Matsumoto T. Effects of androgens on cardiovascular remodeling. *J Endocrinol* . (2012) 214: 1–10. doi: 10.1530/JOE-12-0126

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

19. Svartberg J, von Muhlen D, Schirmer H, Barrett-Connor E, Sundfjord J, Jorde R. Association of endogenous testosterone with blood pressure and left ventricular mass in men. The Tromso Study. *Eur J Endocrinol* . (2004) 150: 65–71. doi: 10.1530/eje.0.1500065

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

20. Jones RD, Nettleship JE, Kapoor D, Jones HT, Channer KS. Testosterone and atherosclerosis in aging men: purported association and clinical implications. *Am J Cardiovasc Drugs* . (2005) 5: 141–54. doi: 10.2165/00129784-200505030-00001

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

21. Cohen PG. Diabetes mellitus is associated with subnormal levels of free testosterone in men. *BJU Int* . (2006) 97: 652–3. doi: 10.1111/j.1464-410X.2006.06111_2.x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

22. Laughlin GA, Barrett-Connor E, Bergstrom J. Low serum testosterone and mortality in older men. *J Clin Endocrinol Metab* . (2008) 93: 68–75. doi: 10.1210/jc.2007-1792

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

23. Traish AM, Saad F, Feeley RJ, Guay A. The dark side of testosterone deficiency: III. Cardiovascular disease. *J Androl* . (2009) 30: 477–94. doi: 10.2164/jandrol.108.007245

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

24. Guay AT. The emerging link between hypogonadism and metabolic syndrome. *J Androl* . (2009) 30: 370–6. doi: 10.2164/jandrol.108.006015

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

25. Dandona P, Dhindsa S. Update: hypogonadotropic hypogonadism in type 2 diabetes and obesity. *J Clin Endocrinol Metab* . (2011) 96: 2643–51. doi: 10.1210/jc.2010-2724

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

26. Maggio M, Lauretani F, Ceda GP, Bandinelli S, Ling SM, Metter EJ, et al. Relationship between low levels of anabolic hormones and 6-year mortality in older men: the aging in the Chianti Area (InCHIANTI) study. *Arch Intern Med* . (2007) 167: 2249–54. doi: 10.1001/archinte.167.20.2249

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

27. Elagizi A, Kohler TS, Lavie CJ. Testosterone and cardiovascular health. *Mayo Clin Proc* . (2018) 93: 83–100. doi: 10. 1016/j. mayocp. 2017. 11. 006

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

28. Jeong SM, Ham BK, Park MG, Oh MM, Yoon DK, Kim JJ, et al. Effect of testosterone replacement treatment in testosterone deficiency syndrome patients with metabolic syndrome. *Korean J Urol* . (2011) 52: 566–71. doi: 10. 4111/kju. 2011. 52. 8. 566

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

29. Kloner RA. Testosterone replacement therapy: new data on efficacy and cardiovascular safety. *J Cardiovasc Pharmacol Ther* . (2016) 22: 54–5. doi: 10. 1177/1074248416646938

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

30. Feldman HA, Longcope C, Derby CA, Johannes CB, Araujo AB, Coviello AD, et al. Age trends in the level of serum testosterone and other hormones in middle-aged men: longitudinal results from the Massachusetts male aging study. *J Clin Endocrinol Metab* . (2002) 87: 589–98. doi: 10. 1210/jcem. 87. 2. 8201

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

31. Shores MM, Mocerri VM, Gruenewald DA, Brodtkin KI, Matsumoto AM, Kivlahan DR. Low testosterone is associated with decreased function and increased mortality risk: a preliminary study of men in a geriatric

rehabilitation unit. *J Am Geriatr Soc* . (2004) 52: 2077–81. doi: 10. 1111/j. 1532-5415. 2004. 52562. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

32. Handelsman DJ, Liu PY. Andropause: invention, prevention, rejuvenation. *Trends Endocrinol Metab* . (2005) 16: 39–45. doi: 10. 1016/j. tem. 2005. 01. 002

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

33. Schaap LA, Pluijm SM, Deeg DJ, Penninx BW, Nicklas BJ, Lips P, et al. Low testosterone levels and decline in physical performance and muscle strength in older men: findings from two prospective cohort studies. *Clin Endocrinol* . (2008) 68: 42–50. doi: 10. 1111/j. 1365-2265. 2007. 02997. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

34. Stellato RK, Feldman HA, Hamdy O, Horton ES, McKinlay JB. Testosterone, sex hormone-binding globulin, and the development of type 2 diabetes in middle-aged men: prospective results from the Massachusetts male aging study. *Diabetes Care* . (2000) 23: 490–4. doi: 10. 2337/diacare. 23. 4. 490

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

35. Carlstrom K, Eriksson A, Stege R, Rannevik G. Relationship between serum testosterone and sex hormone-binding globulin in adult men with intact or absent gonadal function. *Int J Androl* . (1990) 13: 67–73. doi: 10. 1111/j. 1365-2605. 1990. tb00961. x

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

36. Krasnoff JB, Basaria S, Pencina MJ, Jasuja GK, Vasani RS, Ulfelder J, et al. Free testosterone levels are associated with mobility limitation and physical performance in community-dwelling men: the Framingham Offspring Study. *J Clin Endocrinol Metab* . (2010) 95: 2790–9. doi: 10.1210/jc.2009-2680

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

37. Eichholzer M, Barbir A, Basaria S, Dobs AS, Feinleib M, Guallar E, et al. Serum sex steroid hormones and frailty in older American men of the Third National Health and Nutrition Examination Survey (NHANES III). *Aging Male* . (2012) 15: 208–15. doi: 10.3109/13685538.2012.705366

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

38. Gobinet J, Poujol N, Sultan C. Molecular action of androgens. *Mol Cell Endocrinol* . (2002) 198: 15–24. doi: 10.1016/S0303-7207(02)00364-7

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

39. Sullivan ML, Martinez CM, Gennis P, Gallagher EJ. The cardiac toxicity of anabolic steroids. *Prog Cardiovasc Dis* . (1998) 41: 1–15. doi: 10.1016/S0033-0620(98)80019-4

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

40. Yialamas MA, Hayes FJ. Androgens and the ageing male and female. *Best Pract Res Clin Endocrinol Metab* . (2003) 17: 223–36. doi: 10.1016/S1521-690X(03)00018-6

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

41. Theodoraki A, Bouloux PM. Testosterone therapy in men. *Menopause Int* . (2009) 15: 87–92. doi: 10. 1258/mi. 2009. 009025

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

42. Dandona P, Rosenberg MT. A practical guide to male hypogonadism in the primary care setting. *Int J Clin Pract* . (2010) 64: 682–96. doi: 10. 1111/j. 1742-1241. 2010. 02355. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

43. Miner MM, Khera M, Bhattacharya RK, Blick G, Kushner H. Baseline data from the TRIUS registry: symptoms and comorbidities of testosterone deficiency. *Postgrad Med* . (2011) 123: 17–27. doi: 10. 3810/pgm. 2011. 05. 2280

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

44. Morales A. The long and tortuous history of the discovery of testosterone and its clinical application. *J Sex Med* . (2013) 10: 1178–83. doi: 10. 1111/jsm. 12081

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

45. Basaria S, Coviello AD, Travison TG, Storer TW, Farwell WR, Jette AM, et al. Adverse events associated with testosterone administration. *N Engl J Med* . (2010) 363: 109–22. doi: 10. 1056/NEJMoa1000485

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

46. Borst SE, Shuster JJ, Zou B, Ye F, Jia H, Wokhlu A, et al. Cardiovascular risks and elevation of serum DHT vary by route of testosterone administration: a systematic review and meta-analysis. *BMC Med* . (2014) 12: 211. doi: 10. 1186/s12916-014-0211-5

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

47. Onasanya O, Iyer G, Lucas E, Lin D, Singh S, Alexander GC. Association between exogenous testosterone and cardiovascular events: an overview of systematic reviews. *Lancet Diabetes Endocrinol* . (2016) 4: 943–56. doi: 10. 1016/S2213-8587(16)30215-7

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

48. Haddad RM, Kennedy CC, Caples SM, Tracz MJ, Bolona ER, Sideras K, et al. Testosterone and cardiovascular risk in men: a systematic review and meta-analysis of randomized placebo-controlled trials. *Mayo Clin Proc* . (2007) 82: 29–39. doi: 10. 1016/S0025-6196(11)60964-6

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

49. Corona G, Monami M, Rastrelli G, Aversa A, Tishova Y, Saad F, et al. Testosterone and metabolic syndrome: a meta-analysis study. *J Sex Med* . (2011) 8: 272–83. doi: 10. 1111/j. 1743-6109. 2010. 01991. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

50. Haffner SM. Sex hormones, obesity, fat distribution, type 2 diabetes and insulin resistance: epidemiological and clinical correlation. *Int J Obes Relat Metab Disord.* (2000) 24 (Suppl. 2): S56–8. doi: 10. 1038/sj. ijo. 0801279

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

51. Corona G, Monami M, Rastrelli G, Aversa A, Sforza A, Lenzi A, et al. Type 2 diabetes mellitus and testosterone: a meta-analysis study. *Int J Androl.* (2011) 34(6 Pt 1): 528–40. doi: 10. 1111/j. 1365-2605. 2010. 01117. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

52. Hackett G, Kirby M, Sinclair AJ. Testosterone deficiency, cardiac health, and older men. *Int J Endocrinol .* (2014) 2014: 143763. doi: 10. 1155/2014/143763

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

53. Giagulli VA, Triggiani V, Corona G, Carbone D, Licchelli B, Tafaro E, et al. Evidence-based medicine update on testosterone replacement therapy (TRT) in male hypogonadism: focus on new formulations. *Curr Pharm Des .* (2011) 17: 1500–11. doi: 10. 2174/138161211796197160

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

54. Traish AM, Haider A, Haider KS, Doros G, Saad F. Long-term testosterone therapy improves cardiometabolic function and reduces risk of cardiovascular disease in men with hypogonadism: a real-life observational registry study setting comparing treated and untreated (control) groups. *J*

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

Cardiovasc Pharmacol Ther . (2017) 22: 414–33. doi: 10.

1177/1074248417691136

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

55. Kelly DM, Jones TH. Testosterone: a metabolic hormone in health and disease. *J Endocrinol* . (2013) 217: R25–45. doi: 10. 1530/JOE-12-0455

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

56. Kapoor D, Aldred H, Clark S, Channer KS, Jones TH. Clinical and biochemical assessment of hypogonadism in men with type 2 diabetes: correlations with bioavailable testosterone and visceral adiposity. *Diabetes Care* . (2007) 30: 911–7. doi: 10. 2337/dc06-1426

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

57. Petersson SJ, Christensen LL, Kristensen JM, Kruse R, Andersen M, Hojlund K. Effect of testosterone on markers of mitochondrial oxidative phosphorylation and lipid metabolism in muscle of aging men with subnormal bioavailable testosterone. *Eur J Endocrinol* . (2014) 171: 77–88. doi: 10. 1530/EJE-14-0006

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

58. LaPier TL, Rodnick KJ. Changes in cardiac energy metabolism during early development of female SHR. *Am J Hypertens* . (2000) 13: 1074–81. doi: 10. 1016/S0895-7061(00)00297-1

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

59. Morley JE. Testosterone replacement in older men and women. *J Genet Specif Med* . (2001) 4: 49–53.

[PubMed Abstract](#) | [Google Scholar](#)

60. Marzetti E, Leeuwenburgh C. Skeletal muscle apoptosis, sarcopenia and frailty at old age. *Exp Gerontol* . (2006) 41: 1234–8. doi: 10. 1016/j. exger. 2006. 08. 011

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

61. Haren MT, Siddiqui AM, Armbrecht HJ, Kevorkian RT, Kim MJ, Haas MJ, et al. Testosterone modulates gene expression pathways regulating nutrient accumulation, glucose metabolism and protein turnover in mouse skeletal muscle. *Int J Androl* . (2011) 34: 55–68. doi: 10. 1111/j. 1365-2605. 2010. 01061. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

62. Degens H, Korhonen MT. Factors contributing to the variability in muscle ageing. *Maturitas* . (2012) 73: 197–201. doi: 10. 1016/j. maturitas. 2012. 07. 015

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

63. Kolwicz SC Jr, Purohit S, Tian R. Cardiac metabolism and its interactions with contraction, growth, and survival of cardiomyocytes. *Circ Res* . (2013) 113: 603–16. doi: 10. 1161/CIRCRESAHA. 113. 302095

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

64. Lyons MR, Peterson LR, McGill JB, Herrero P, Coggan AR, Saeed IM, et al. Impact of sex on the heart's metabolic and functional responses to diabetic therapies. *Am J Physiol Heart Circ Physiol* . (2013) 305: H1584–91. doi: 10.1152/ajpheart.00420.2013

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

65. Ghanim H, Dhindsa S, Batra M, Green K, Abuaysheh S, Kuhadiya ND, et al. Effect of testosterone on FGF2, MRF4, and myostatin in hypogonadotropic hypogonadism: relevance to muscle growth. *J Clin Endocrinol Metab* . (2019) 104: 2094–102. doi: 10.1210/jc.2018-01832

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

66. Kuhn-Velten N, Bos D, Schermer R, Staib W. Age-dependence of the rat Leydig cell and Sertoli cell function. Development of the peripheral testosterone level and its relation to mitochondrial and microsomal cytochromes P-450 and to androgen-binding protein. *Acta Endocrinol*. (1987) 115: 275–81. doi: 10.1530/acta.0.1150275

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

67. Walker WH, Cheng J. FSH and testosterone signaling in Sertoli cells. *Reproduction* . (2005) 130: 15–28. doi: 10.1530/rep.1.00358

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

68. Wang Y, Chen F, Ye L, Zirkin B, Chen H. Steroidogenesis in Leydig cells: effects of aging and environmental factors. *Reproduction* . (2017) 154: R111-22. doi: 10. 1530/REP-17-0064

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

69. Baghaei F, Rosmond R, Westberg L, Hellstrand M, Eriksson E, Holm G, et al. The CYP19 gene and associations with androgens and abdominal obesity in premenopausal women. *Obes Res* . (2003) 11: 578-85. doi: 10. 1038/oby. 2003. 81

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

70. Mooradian AD, Morley JE, Korenman SG. Biological actions of androgens. *Endocr Rev* . (1987) 8: 1-28. doi: 10. 1210/edrv-8-1-1

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

71. Marsh JD, Lehmann MH, Ritchie RH, Gwathmey JK, Green GE, Schiebinger RJ. Androgen receptors mediate hypertrophy in cardiac myocytes. *Circulation* . (1998) 98: 256-61. doi: 10. 1161/01. CIR. 98. 3. 256

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

72. Janne OA, Moilanen AM, Poukka H, Rouleau N, Karvonen U, Kotaja N, et al. Androgen-receptor-interacting nuclear proteins. *Biochem Soc Trans* . (2000) 28: 401-5. doi: 10. 1042/bst0280401

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

73. Simental JA, Sar M, Wilson EM. Domain functions of the androgen receptor. *J Steroid Biochem Mol Biol* . (1992) 43: 37–41. doi: 10.1016/0960-0760(92)90185-L

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

74. Roy AK, Tyagi RK, Song CS, Lavrovsky Y, Ahn SC, Oh TS, et al. Androgen receptor: structural domains and functional dynamics after ligand-receptor interaction. *Ann N Y Acad Sci* . (2001) 949: 44–57. doi: 10.1111/j.1749-6632.2001.tb04001.x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

75. Heinlein CA, Chang C. Androgen receptor (AR) coregulators: an overview. *Endocr Rev* . (2002) 23: 175–200. doi: 10.1210/edrv.23.2.0460

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

76. Simental JA, Sar M, Lane MV, French FS, Wilson EM. Transcriptional activation and nuclear targeting signals of the human androgen receptor. *J Biol Chem* . (1991) 266: 510–8.

[PubMed Abstract](#) | [Google Scholar](#)

77. Heinlein CA, Chang C. The roles of androgen receptors and androgen-binding proteins in nongenomic androgen actions. *Mol Endocrinol* . (2002) 16: 2181–7. doi: 10.1210/me.2002-0070

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

78. Vicencio JM, Ibarra C, Estrada M, Chiong M, Soto D, Parra V, et al.

Testosterone induces an intracellular calcium increase by a nongenomic mechanism in cultured rat cardiac myocytes. *Endocrinology* . (2006) 147: 1386–95. doi: 10.1210/en.2005-1139

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

79. Altamirano F, Oyarce C, Silva P, Toyos M, Wilson C, Lavandero S, et al.

Testosterone induces cardiomyocyte hypertrophy through mammalian target of rapamycin complex 1 pathway. *J Endocrinol* . (2009) 202: 299–307. doi: 10.1677/JOE-09-0044

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

80. Wilson C, Contreras-Ferrat A, Venegas N, Osorio-Fuentealba C, Pavez M, Montoya K, et al. Testosterone increases GLUT4-dependent glucose uptake in cardiomyocytes. *J Cell Physiol* . (2013) 228: 2399–407. doi: 10.1002/jcp.24413

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

81. Gyawali P, Martin SA, Heilbronn LK, Vincent AD, Taylor AW, Adams RJT, et al. The role of sex hormone-binding globulin (SHBG), testosterone, and other sex steroids, on the development of type 2 diabetes in a cohort of community-dwelling middle-aged to elderly men. *Acta Diabetol* . (2018) 55: 861–72. doi: 10.1007/s00592-018-1163-6

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

82. Hagen FS, Arguelles C, Sui LM, Zhang W, Seidel PR, Conroy SC, et al. Mammalian expression of the human sex steroid-binding protein of plasma (SBP or SHBG) and testis (ABP). Characterization of the recombinant protein. *FEBS Lett* . (1992) 299: 23–7. doi: 10. 1016/0014-5793(92)80091-T

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

83. Laurent MR, Helsen C, Antonio L, Schollaert D, Joniau S, Vos MJ, et al. Effects of sex hormone-binding globulin (SHBG) on androgen bioactivity *in vitro* . *Mol Cell Endocrinol* . (2016) 437: 280–91. doi: 10. 1016/j. mce. 2016. 08. 041

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

84. Hammes A, Andreassen TK, Spoelgen R, Raila J, Hubner N, Schulz H, et al. Role of endocytosis in cellular uptake of sex steroids. *Cell* . (2005) 122: 751–62. doi: 10. 1016/j. cell. 2005. 06. 032

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

85. Mohammed M, Al-Habori M, Abdullateef A, Saif-Ali R. Impact of metabolic syndrome factors on testosterone and SHBG in type 2 diabetes mellitus and metabolic syndrome. *J Diabetes Res* . (2018) 2018: 4926789. doi: 10. 1155/2018/4926789

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

86. Brand JS, van der Schouw YT. Testosterone, SHBG and cardiovascular health in postmenopausal women. *Int J Impot Res* . (2010) 22: 91-104. doi: 10. 1038/ijir. 2009. 64

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

87. Brand JS, Wareham NJ, Dowsett M, Folkerd E, van der Schouw YT, Luben RN, et al. Associations of endogenous testosterone and SHBG with glycosylated haemoglobin in middle-aged and older men. *Clin Endocrinol* . (2011) 74: 572-8. doi: 10. 1111/j. 1365-2265. 2010. 03951. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

88. English KM, Mandour O, Steeds RP, Diver MJ, Jones TH, Channer KS. Men with coronary artery disease have lower levels of androgens than men with normal coronary angiograms. *Eur Heart J* . (2000) 21: 890-4. doi: 10. 1053/euhj. 1999. 1873

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

89. Hayward CS, Webb CM, Collins P. Effect of sex hormones on cardiac mass. *Lancet* . (2001) 357: 1354-6. doi: 10. 1016/S0140-6736(00)04523-2

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

90. Jones RD, Pugh PJ, Hall J, Channer KS, Jones TH. Altered circulating hormone levels, endothelial function and vascular reactivity in the testicular feminised mouse. *Eur J Endocrinol* . (2003) 148: 111-20. doi: 10. 1530/eje. 0. 1480111

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

91. Corona G, Rastrelli G, Vignozzi L, Mannucci E, Maggi M. Testosterone, cardiovascular disease and the metabolic syndrome. *Best Pract Res Clin Endocrinol Metab* . (2011) 25: 337-53. doi: 10. 1016/j. beem. 2010. 07. 002

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

92. Oskui PM, French WJ, Herring MJ, Mayeda GS, Burstein S, Kloner RA. Testosterone and the cardiovascular system: a comprehensive review of the clinical literature. *J Am Heart Assoc* . (2013) 2: e000272. doi: 10. 1161/JAHA. 113. 000272

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

93. Sullivan ML, Martinez CM, Gallagher EJ. Atrial fibrillation and anabolic steroids. *J Emerg Med* . (1999) 17: 851-7. doi: 10. 1016/S0736-4679(99)00095-5

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

94. Houghton DE, Alsawas M, Barrioneuvo P, Tello M, Farah W, Beuschel B, et al. Testosterone therapy and venous thromboembolism: A systematic review and meta-analysis. *Thromb Res* . (2018) 172: 94-103. doi: 10. 1016/j. thromres. 2018. 10. 023

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

95. Liu XK, Katchman A, Whitfield BH, Wan G, Janowski EM, Woosley RL, et al. *In vivo* androgen treatment shortens the QT interval and increases the
<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

densities of inward and delayed rectifier potassium currents in orchietomized male rabbits. *Cardiovasc Res* . (2003) 57: 28–36. doi: 10.1016/S0008-6363(02)00673-9

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

96. Basaria S, Harman SM, Travison TG, Hodis H, Tsitouras P, Budoff M, et al. Effects of testosterone administration for 3 years on subclinical atherosclerosis progression in older men with low or low-normal testosterone levels: a randomized clinical trial. *JAMA* . (2015) 314: 570–81. doi: 10.1001/jama. 2015. 8881

[CrossRef Full Text](#) | [Google Scholar](#)

97. Budoff MJ, Ellenberg SS, Lewis CE, Mohler ER III, Wenger NK, Bhasin S, et al. Testosterone treatment and coronary artery plaque volume in older men with low testosterone. *JAMA*. (2017) 317: 708–16. doi: 10.1001/jama. 2016. 21043

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

98. Mirdamadi A, Garakyaraghi M, Pourmoghaddas A, Bahmani A, Mahmoudi H, Gharipour M. Beneficial effects of testosterone therapy on functional capacity, cardiovascular parameters, and quality of life in patients with congestive heart failure. *Biomed Res Int* . (2014) 2014: 392432. doi: 10.1155/2014/392432

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

99. Malkin CJ, Pugh PJ, Jones RD, Jones TH, Channer KS. Testosterone as a protective factor against atherosclerosis-immunomodulation and influence upon plaque development and stability. *J Endocrinol* . (2003) 178: 373-80. doi: 10.1677/joe.0.1780373

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

100. Malkin CJ, Pugh PJ, Jones RD, Kapoor D, Channer KS, Jones TH. The effect of testosterone replacement on endogenous inflammatory cytokines and lipid profiles in hypogonadal men. *J Clin Endocrinol Metab* . (2004) 89: 3313-8. doi: 10.1210/jc.2003-031069

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

101. Herring MJ, Oskui PM, Hale SL, Kloner RA. Testosterone and the cardiovascular system: a comprehensive review of the basic science literature. *J Am Heart Assoc* . (2013) 2: e000271. doi: 10.1161/JAHA.113.000271

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

102. Caminiti G, Volterrani M, Iellamo F, Marazzi G, Massaro R, Miceli M, et al. Effect of long-acting testosterone treatment on functional exercise capacity, skeletal muscle performance, insulin resistance, and baroreflex sensitivity in elderly patients with chronic heart failure a double-blind, placebo-controlled, randomized study. *J Am Coll Cardiol* . (2009) 54: 919-27. doi: 10.1016/j.jacc.2009.04.078

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

103. Dubey RK, Oparil S, Imthurn B, Jackson EK. Sex hormones and hypertension. *Cardiovasc Res* . (2002) 53: 688–708. doi: 10. 1016/S0008-6363(01)00527-2

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

104. Fogari R, Preti P, Zoppi A, Fogari E, Rinaldi A, Corradi L, et al. Serum testosterone levels and arterial blood pressure in the elderly. *Hypertens Res* . (2005) 28: 625–30. doi: 10. 1291/hypres. 28. 625

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

105. Kienitz T, Quinkler M. Testosterone and blood pressure regulation. *Kidney Blood Press Res* . (2008) 31: 71–9. doi: 10. 1159/000119417

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

106. Stamler J, Stamler R, Riedlinger WF, Algera G, Roberts RH. Hypertension screening of 1 million Americans. Community Hypertension Evaluation Clinic (CHEC) program, 1973 through 1975. *JAMA*. (1976) 235: 2299–306. doi: 10. 1001/jama. 1976. 03260470017018

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

107. Akkad AA, Halligan AW, Abrams K, al-Azzawi F. Differing responses in blood pressure over 24 hours in normotensive women receiving oral or transdermal estrogen replacement therapy. *Obstet Gynecol*. (1997) 89: 97–103. doi: 10. 1016/S0029-7844(97)84258-5

[CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

108. Pripp U, Hall G, Csemiczky G, Eksborg S, Landgren BM, Schenck-Gustafsson K. A randomized trial on effects of hormone therapy on ambulatory blood pressure and lipoprotein levels in women with coronary artery disease. *J Hypertens* . (1999) 17: 1379–86. doi: 10. 1097/00004872-199917100-00004

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

109. Malkin CJ, Jones RD, Jones TH, Channer KS. Effect of testosterone on ex vivo vascular reactivity in man. *Clin Sci* . (2006) 111: 265–74. doi: 10. 1042/CS20050354

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

110. Kelly DM, Jones TH. Testosterone: a vascular hormone in health and disease. *J Endocrinol* . (2013) 217: R47–71. doi: 10. 1530/JOE-12-0582

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

111. Stuck BJ, Lenski M, Bohm M, Laufs U. Metabolic switch and hypertrophy of cardiomyocytes following treatment with angiotensin II are prevented by AMP-activated protein kinase. *J Biol Chem* . (2008) 283: 32562–9. doi: 10. 1074/jbc. M801904200

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

112. Chan AY, Soltys CL, Young ME, Proud CG, Dyck JR. Activation of AMP-activated protein kinase inhibits protein synthesis associated with

hypertrophy in the cardiac myocyte. *J Biol Chem* . (2004) 279: 32771–9. doi: 10.1074/jbc.M403528200

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

113. Kolwicz SC Jr, Tian R. Glucose metabolism and cardiac hypertrophy. *Cardiovasc Res* . (2011) 90: 194–201. doi: 10.1093/cvr/cvr071

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

114. Nascimben L, Ingwall JS, Lorell BH, Pinz I, Schultz V, Tornheim K, et al. Mechanisms for increased glycolysis in the hypertrophied rat heart. *Hypertension* . (2004) 44: 662–7. doi: 10.1161/01.HYP.0000144292.69599.0c

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

115. Cavasin MA, Tao ZY, Yu AL, Yang XP. Testosterone enhances early cardiac remodeling after myocardial infarction, causing rupture and degrading cardiac function. *Am J Physiol Heart Circ Physiol* . (2006) 290: H2043–50. doi: 10.1152/ajpheart.01121.2005

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

116. Tian R. Transcriptional regulation of energy substrate metabolism in normal and hypertrophied heart. *Curr Hypertens Rep* . (2003) 5: 454–8. doi: 10.1007/s11906-003-0052-7

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

117. Scolletta S, Biagioli B. Energetic myocardial metabolism and oxidative stress: let's make them our friends in the fight against heart failure. *Biomed Pharmacother* . (2010) 64: 203–7. doi: 10. 1016/j. biopha. 2009. 10. 002

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

118. Chandel A, Dhindsa S, Topiwala S, Chaudhuri A, Dandona P. Testosterone concentration in young patients with diabetes. *Diabetes Care* . (2008) 31: 2013–7. doi: 10. 2337/dc08-0851

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

119. Abel ED, Doenst T. Mitochondrial adaptations to physiological vs. pathological cardiac hypertrophy. *Cardiovasc Res* . (2011) 90: 234–42. doi: 10. 1093/cvr/cvr015

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

120. Cheng LF, Wang F, Lopatin AN. Metabolic stress in isolated mouse ventricular myocytes leads to remodeling of t tubules. *Am J Physiol Heart Circ Physiol* . (2011) 301: H1984–95. doi: 10. 1152/ajpheart. 00304. 2011

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

121. Wallace DC, Fan W. Energetics, epigenetics, mitochondrial genetics. *Mitochondrion* . (2010) 10: 12–31. doi: 10. 1016/j. mito. 2009. 09. 006

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

122. Villena JA, Vinas O, Mampel T, Iglesias R, Giralt M, Villarroya F.

Regulation of mitochondrial biogenesis in brown adipose tissue: nuclear respiratory factor-2/GA-binding protein is responsible for the transcriptional regulation of the gene for the mitochondrial ATP synthase beta subunit.

Biochem J. (1998) 331: 121–7. doi: 10. 1042/bj3310121

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

123. Doeg KA, Polomski LL, Doeg LH. Androgen control of mitochondrial and nuclear DNA synthesis in male sex accessory tissue of castrate rats.

Endocrinology . (1972) 90: 1633–8. doi: 10. 1210/endo-90-6-1633

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

124. Juang HH, Hsieh ML, Tsui KH. Testosterone modulates mitochondrial aconitase in the full-length human androgen receptor-transfected PC-3 prostatic carcinoma cells. *J Mol Endocrinol* . (2004) 33: 121–32. doi: 10.

1677/jme. 0. 0330121

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

125. Wang RS, Chang HY, Kao SH, Kao CH, Wu YC, Yeh S, et al. Abnormal mitochondrial function and impaired granulosa cell differentiation in androgen receptor knockout mice. *Int J Mol Sci* . (2015) 16: 9831–49. doi: 10.

3390/ijms16059831

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

126. Li YX, Jiang B, Li Y, Xia F, Yu J, Yang LZ, et al. Mitochondrial apoptotic pathways: a mechanism for low androgen-induced vascular endothelial injury in male rats. *Horm Metab Res* . (2011) 43: 374–9. doi: 10. 1055/s-0031-1271745

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

127. Fernando SM, Rao P, Niel L, Chatterjee D, Stagljar M, Monks DA. Myocyte androgen receptors increase metabolic rate and improve body composition by reducing fat mass. *Endocrinology* . (2010) 151: 3125–32. doi: 10. 1210/en. 2010-0018

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

128. van de Wijngaart DJ, Dubbink HJ, van Royen ME, Trapman J, Jenster G. Androgen receptor coregulators: recruitment via the coactivator binding groove. *Mol Cell Endocrinol*. (2012) 352: 57–69. doi: 10. 1016/j. mce. 2011. 08. 007

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

129. Duncan JG, Finck BN. The PPARalpha-PGC-1alpha axis controls cardiac energy metabolism in healthy and diseased myocardium. *PPAR Res* . (2008) 2008: 253817. doi: 10. 1155/2008/253817

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

130. Lehman JJ, Barger PM, Kovacs A, Saffitz JE, Medeiros DM, Kelly DP. Peroxisome proliferator-activated receptor gamma coactivator-1 promotes

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

cardiac mitochondrial biogenesis. *J Clin Invest* . (2000) 106: 847–56. doi: 10.1172/JCI10268

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

131. Arany Z, He H, Lin J, Hoyer K, Handschin C, Toka O, et al. Transcriptional coactivator PGC-1 alpha controls the energy state and contractile function of cardiac muscle. *Cell Metab* . (2005) 1: 259–71. doi: 10.1016/j.cmet.2005.03.002

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

132. Wallberg AE, Yamamura S, Malik S, Spiegelman BM, Roeder RG. Coordination of p300-mediated chromatin remodeling and TRAP/mediator function through coactivator PGC-1alpha. *Mol Cell* . (2003) 12: 1137–49. doi: 10.1016/S1097-2765(03)00391-5

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

133. Vega RB, Huss JM, Kelly DP. The coactivator PGC-1 cooperates with peroxisome proliferator-activated receptor alpha in transcriptional control of nuclear genes encoding mitochondrial fatty acid oxidation enzymes. *Mol Cell Biol* . (2000) 20: 1868–76. doi: 10.1128/MCB.20.5.1868-1876.2000

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

134. Huss JM, Kopp RP, Kelly DP. Peroxisome proliferator-activated receptor coactivator-1alpha (PGC-1alpha) coactivates the cardiac-enriched nuclear receptors estrogen-related receptor-alpha and -gamma. Identification of

novel leucine-rich interaction motif within PGC-1alpha. *J Biol Chem.* (2002) 277: 40265–74. doi: 10. 1074/jbc. M206324200

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

135. Czubryt MP, McAnally J, Fishman GI, Olson EN. Regulation of peroxisome proliferator-activated receptor gamma coactivator 1 alpha (PGC-1 alpha) and mitochondrial function by MEF2 and HDAC5. *Proc Natl Acad Sci USA* . (2003) 100: 1711–6. doi: 10. 1073/pnas. 0337639100

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

136. Usui T, Kajita K, Kajita T, Mori I, Hanamoto T, Ikeda T, et al. Elevated mitochondrial biogenesis in skeletal muscle is associated with testosterone-induced body weight loss in male mice. *FEBS Lett* . (2014) 588: 1935–41. doi: 10. 1016/j. febslet. 2014. 03. 051

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

137. Shiota M, Yokomizo A, Tada Y, Inokuchi J, Tatsugami K, Kuroiwa K, et al. Peroxisome proliferator-activated receptor gamma coactivator-1alpha interacts with the androgen receptor (AR) and promotes prostate cancer cell growth by activating the AR. *Mol Endocrinol* . (2010) 24: 114–27. doi: 10. 1210/me. 2009-0302

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

138. Juang HH, Costello LC, Franklin RB. Androgen modulation of multiple transcription start sites of the mitochondrial aspartate aminotransferase

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

gene in rat prostate. *J Biol Chem* . (1995) 270: 12629–34. doi: 10. 1074/jbc. 270. 21. 12629

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

139. Koenig H, Goldstone A, Lu CY. Androgens regulate mitochondrial cytochrome c oxidase and lysosomal hydrolases in mouse skeletal muscle. *Biochem J* . (1980) 192: 349–53. doi: 10. 1042/bj1920349

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

140. Canto C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, et al. AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature* . (2009) 458: 1056–60. doi: 10. 1038/nature07813

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

141. Canto C, Auwerx J. PGC-1alpha, SIRT1 and AMPK, an energy sensing network that controls energy expenditure. *Curr Opin Lipidol* . (2009) 20: 98–105. doi: 10. 1097/MOL. 0b013e328328d0a4

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

142. McGee SL, Hargreaves M. AMPK and transcriptional regulation. *Front Biosci* . (2008) 13: 3022–33. doi: 10. 2741/2907

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

143. Ghanim H, Dhindsa S, Batra M, Green K, Abuaysheh S, Kuhadiya ND, et al. Testosterone increases the expression and phosphorylation of AMP kinase

alpha in men with hypogonadism and Type 2 diabetes. *J Clin Endocrinol Metab* . (2020) 105: dgz288. doi: 10. 1210/clinem/dgz288

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

144. Pang T, Rajapurohitam V, Cook MA, Karmazyn M. Differential AMPK phosphorylation sites associated with phenylephrine vs. antihypertrophic effects of adenosine agonists in neonatal rat ventricular myocytes. *Am J Physiol Heart Circ Physiol* . (2010) 298: H1382–90. doi: 10. 1152/ajpheart. 00424. 2009

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

145. Meng R, Pei Z, Zhang A, Zhou Y, Cai X, Chen B, et al. AMPK activation enhances PPARalpha activity to inhibit cardiac hypertrophy via ERK1/2 MAPK signaling pathway. *Arch Biochem Biophys* . (2011) 511: 1–7. doi: 10. 1016/j. abb. 2011. 04. 010

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

146. Li Y, Chen C, Yao F, Su Q, Liu D, Xue R, et al. AMPK inhibits cardiac hypertrophy by promoting autophagy via mTORC1. *Arch Biochem Biophys* . (2014) 558: 79–86. doi: 10. 1016/j. abb. 2014. 06. 023

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

147. Klip A. The many ways to regulate glucose transporter 4. *Appl Physiol Nutr Metab* . (2009) 34: 481–7. doi: 10. 1139/H09-047

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

148. Cortes R, Rivera M, Rosello-Lleti E, Martinez-Dolz L, Almenar L, Azorin I, et al. Differences in MEF2 and NFAT transcriptional pathways according to human heart failure aetiology. *PLoS ONE* . (2012) 7: e30915. doi: 10.1371/journal.pone.0030915

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

149. Desjardins CA, Naya FJ. The function of the MEF2 family of transcription factors in cardiac development, cardiogenomics, and direct reprogramming. *J Cardiovasc Dev Dis*. (2016) 3: 26. doi: 10.3390/jcdd3030026

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

150. Leick L, Hellsten Y, Fentz J, Lyngby SS, Wojtaszewski JF, Hidalgo J, et al. PGC-1alpha mediates exercise-induced skeletal muscle VEGF expression in mice. *Am J Physiol Endocrinol Metab* . (2009) 297: E92–103. doi: 10.1152/ajpendo.00076.2009

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

151. Hondares E, Rosell M, Diaz-Delfin J, Olmos Y, Monsalve M, Iglesias R, et al. Peroxisome proliferator-activated receptor alpha (PPARalpha) induces PPARgamma coactivator 1alpha (PGC-1alpha) gene expression and contributes to thermogenic activation of brown fat: involvement of PRDM16. *J Biol Chem* . (2011) 286: 43112–22. doi: 10.1074/jbc.M111.252775

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

152. Irrcher I, Ljubcic V, Kirwan AF, Hood DA. AMP-activated protein kinase-regulated activation of the PGC-1alpha promoter in skeletal muscle cells. *PLoS ONE* . (2008) 3: e3614. doi: 10. 1371/journal. pone. 0003614

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

153. Rowe GC, Jiang A, Arany Z. PGC-1 coactivators in cardiac development and disease. *Circ Res* . (2010) 107: 825–38. doi: 10. 1161/CIRCRESAHA. 110. 223818

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

154. Lai L, Leone TC, Zechner C, Schaeffer PJ, Kelly SM, Flanagan DP, et al. Transcriptional coactivators PGC-1alpha and PGC-1beta control overlapping programs required for perinatal maturation of the heart. *Genes Dev* . (2008) 22: 1948–61. doi: 10. 1101/gad. 1661708

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

155. Liang H, Ward WF. PGC-1alpha: a key regulator of energy metabolism. *Adv Physiol Educ* . (2006) 30: 145–51. doi: 10. 1152/advan. 00052. 2006

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

156. Finck BN, Kelly DP. PGC-1 coactivators: inducible regulators of energy metabolism in health and disease. *J Clin Invest* . (2006) 116: 615–22. doi: 10. 1172/JCI27794

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

157. Amat R, Planavila A, Chen SL, Iglesias R, Giralt M, Villarroya F. SIRT1 controls the transcription of the peroxisome proliferator-activated receptor-gamma Co-activator-1alpha (PGC-1alpha) gene in skeletal muscle through the PGC-1alpha autoregulatory loop and interaction with MyoD. *J Biol Chem* . (2009) 284: 21872–80. doi: 10. 1074/jbc. M109. 022749

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

158. Barger PM, Kelly DP. PPAR signaling in the control of cardiac energy metabolism. *Trends Cardiovasc Med* . (2000) 10: 238–45. doi: 10. 1016/S1050-1738(00)00077-3

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

159. Leick L, Fentz J, Bienso RS, Knudsen JG, Jeppesen J, Kiens B, et al. PGC-1{alpha} is required for AICAR-induced expression of GLUT4 and mitochondrial proteins in mouse skeletal muscle. *Am J Physiol Endocrinol Metab* . (2010) 299: E456–65. doi: 10. 1152/ajpendo. 00648. 2009

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

160. Zong H, Ren JM, Young LH, Pypaert M, Mu J, Birnbaum MJ, et al. AMP kinase is required for mitochondrial biogenesis in skeletal muscle in response to chronic energy deprivation. *Proc Natl Acad Sci USA* . (2002) 99: 15983–7. doi: 10. 1073/pnas. 252625599

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

161. Turdi S, Fan X, Li J, Zhao J, Huff AF, Du M, et al. AMP-activated protein kinase deficiency exacerbates aging-induced myocardial contractile dysfunction. *Aging Cell* . (2010) 9: 592–606. doi: 10. 1111/j. 1474-9726. 2010. 00586. x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

162. Tennakoon JB, Shi Y, Han JJ, Tsouko E, White MA, Burns AR, et al. Androgens regulate prostate cancer cell growth via an AMPK-PGC-1alpha-mediated metabolic switch. *Oncogene* . (2014) 33: 5251–61. doi: 10. 1038/onc. 2013. 463

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

163. Backs J, Olson EN. Control of cardiac growth by histone acetylation/deacetylation. *Circ Res* . (2006) 98: 15–24. doi: 10. 1161/01. RES. 0000197782. 21444. 8f

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

164. Sack MN. The role of SIRT3 in mitochondrial homeostasis and cardiac adaptation to hypertrophy and aging. *J Mol Cell Cardiol* . (2012) 52: 520–5. doi: 10. 1016/j. yjmcc. 2011. 11. 004

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

165. Sundaresan NR, Gupta M, Kim G, Rajamohan SB, Isbatan A, Gupta MP. Sirt3 blocks the cardiac hypertrophic response by augmenting Foxo3a-

dependent antioxidant defense mechanisms in mice. *J Clin Invest* . (2009) 119: 2758–71. doi: 10. 1172/JCI39162

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

166. Pillai VB, Sundaresan NR, Jeevanandam V, Gupta MP. Mitochondrial SIRT3 and heart disease. *Cardiovasc Res* . (2010) 88: 250–6. doi: 10. 1093/cvr/cvq250

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

167. Hu DX, Liu XB, Song WC, Wang JA. Roles of SIRT3 in heart failure: from bench to bedside. *J Zhejiang Univ Sci B* . (2016) 17: 821–30. doi: 10. 1631/jzus. B1600253

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

168. Lu Y, Wang YD, Wang XY, Chen H, Cai ZJ, Xiang MX. SIRT3 in cardiovascular diseases: Emerging roles and therapeutic implications. *Int J Cardiol* . (2016) 220: 700–5. doi: 10. 1016/j. ijcard. 2016. 06. 236

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

169. Hirschey MD, Shimazu T, Huang JY, Schwer B, Verdin E. SIRT3 regulates mitochondrial protein acetylation and intermediary metabolism. *Cold Spring Harb Symp Quant Biol* . (2011) 76: 267–77. doi: 10. 1101/sqb. 2011. 76. 010850

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

170. Bause AS, Haigis MC. SIRT3 regulation of mitochondrial oxidative stress. *Exp Gerontol* . (2013) 48: 634–9. doi: 10. 1016/j. exger. 2012. 08. 007

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

171. Koentges C, Pfeil K, Schnick T, Wiese S, Dahlbock R, Cimolai MC, et al. SIRT3 deficiency impairs mitochondrial and contractile function in the heart. *Basic Res Cardiol* . (2015) 110: 36. doi: 10. 1007/s00395-015-0493-6

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

172. Yu W, Gao B, Li N, Wang J, Qiu C, Zhang G, et al. Sirt3 deficiency exacerbates diabetic cardiac dysfunction: Role of Foxo3A-Parkin-mediated mitophagy. *Biochim Biophys Acta* . (2017) 1863: 1973–83. doi: 10. 1016/j. bbadis. 2016. 10. 021

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

173. Hafner AV, Dai J, Gomes AP, Xiao CY, Palmeira CM, Rosenzweig A, et al. Regulation of the mPTP by SIRT3-mediated deacetylation of CypD at lysine 166 suppresses age-related cardiac hypertrophy. *Aging* . (2010) 2: 914–23. doi: 10. 18632/aging. 100252

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

174. Koentges C, Pfeil K, Meyer-Steenbuck M, Lothar A, Hoffmann MM, Odening KE, et al. Preserved recovery of cardiac function following ischemia-reperfusion in mice lacking SIRT3. *Can J Physiol Pharmacol* . (2016) 94: 72–80. doi: 10. 1139/cjpp-2015-0152

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

175. Porter GA, Urciuoli WR, Brookes PS, Nadtochiy SM. SIRT3 deficiency exacerbates ischemia-reperfusion injury: implication for aged hearts. *Am J Physiol Heart Circ Physiol* . (2014) 306: H1602–9. doi: 10. 1152/ajpheart. 00027. 2014

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

176. Altieri P, Barisione C, Lazzarini E, Garuti A, Bezante GP, Canepa M, et al. Testosterone antagonizes doxorubicin-induced senescence of cardiomyocytes. *J Am Heart Assoc*. (2016) 5: e002383. doi: 10. 1161/JAHA. 115. 002383

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

177. Giralt A, Hondares E, Villena JA, Ribas F, Diaz-Delfin J, Giralt M, et al. Peroxisome proliferator-activated receptor-gamma coactivator-1alpha controls transcription of the Sirt3 gene, an essential component of the thermogenic brown adipocyte phenotype. *J Biol Chem* . (2011) 286: 16958–66. doi: 10. 1074/jbc. M110. 202390

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

178. Giralt A, Villarroya F. SIRT3, a pivotal actor in mitochondrial functions: metabolism, cell death and aging. *Biochem J* . (2012) 444: 1–10. doi: 10. 1042/BJ20120030

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

179. Gan L, Jiang W, Xiao YF, Deng L, Gu LD, Guo ZY, et al. Disruption of CD38 gene enhances cardiac functions by elevating serum testosterone in the male null mice. *Life Sci* . (2011) 89: 491–7. doi: 10. 1016/j. lfs. 2011. 07. 020

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

180. Ramjiawan A, Bagchi RA, Blant A, Albak L, Cavasin MA, Horn TR, et al. Roles of histone deacetylation and AMP kinase in regulation of cardiomyocyte PGC-1alpha gene expression in hypoxia. *Am J Physiol Cell Physiol* . (2013) 304: C1064–72. doi: 10. 1152/ajpcell. 00262. 2012

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

181. Planavila A, Iglesias R, Giralt M, Villarroya F. Sirt1 acts in association with PPARalpha to protect the heart from hypertrophy, metabolic dysregulation, and inflammation. *Cardiovasc Res* . (2011) 90: 276–84. doi: 10. 1093/cvr/cvq376

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

182. Ahn TG, Yang G, Lee HM, Kim MD, Choi HY, Park KS, et al. Molecular mechanisms underlying the anti-obesity potential of prunetin, an O-methylated isoflavone. *Biochem Pharmacol* . (2013) 85: 1525–33. doi: 10. 1016/j. bcp. 2013. 02. 020

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

183. Frame S, Cohen P, Biondi RM. A common phosphate binding site explains the unique substrate specificity of GSK3 and its inactivation by phosphorylation. *Mol Cell* . (2001) 7: 1321–7. doi: 10.1016/S1097-2765(01)00253-2

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

184. Hyde Z, Norman PE, Flicker L, Hankey GJ, Almeida OP, McCaul KA, et al. Low free testosterone predicts mortality from cardiovascular disease but not other causes: the Health in Men Study. *J Clin Endocrinol Metab* . (2012) 97: 179–89. doi: 10.1210/jc.2011-1617

[CrossRef Full Text](#) | [Google Scholar](#)

185. Swerdloff RS, Dudley RE, Page ST, Wang C, Salameh WA. Dihydrotestosterone: biochemistry, physiology, and clinical implications of elevated blood levels. *Endocr Rev* . (2017) 38: 220–54. doi: 10.1210/er.2016-1067

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

186. Flo FJ, Kanu O, Teleb M, Chen Y, Siddiqui T. Anabolic androgenic steroid-induced acute myocardial infarction with multiorgan failure. *Proc (Bayl Univ Med Cent)* . (2018) 31: 334–6. doi: 10.1080/08998280.2018.1460130

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

187. Froese N, Wang H, Zwadlo C, Wang Y, Grund A, Gigina A, et al. Anti-androgenic therapy with finasteride improves cardiac function, attenuates

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

remodeling and reverts pathologic gene-expression after myocardial infarction in mice. *J Mol Cell Cardiol* . (2018) 122: 114–24. doi: 10. 1016/j. yjmcc. 2018. 08. 011

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

188. Hou M, Gu HC, Wang HH, Liu XM, Zhou CL, Yang Q, et al. Prenatal exposure to testosterone induces cardiac hypertrophy in adult female rats through enhanced Pkcdelta expression in cardiac myocytes. *J Mol Cell Cardiol* . (2019) 128: 1–10. doi: 10. 1016/j. yjmcc. 2019. 01. 008

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

189. Zwadlo C, Schmidtman E, Szaroszyk M, Kattih B, Froese N, Hinz H, et al. Antiandrogenic therapy with finasteride attenuates cardiac hypertrophy and left ventricular dysfunction. *Circulation* . (2015) 131: 1071–81. doi: 10. 1161/CIRCULATIONAHA. 114. 012066

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

190. Borst SE, Lee JH, Conover CF. Inhibition of 5alpha-reductase blocks prostate effects of testosterone without blocking anabolic effects. *Am J Physiol Endocrinol Metab* . (2005) 288: E222–7. doi: 10. 1152/ajpendo. 00305. 2004

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

191. Yeap BB, Alfonso H, Chubb SA, Handelsman DJ, Hankey GJ, Almeida OP, et al. In older men an optimal plasma testosterone is associated with

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

reduced all-cause mortality and higher dihydrotestosterone with reduced ischemic heart disease mortality, while estradiol levels do not predict mortality. *J Clin Endocrinol Metab* . (2014) 99: E9–18. doi: 10.1210/jc.2013-3272

[CrossRef Full Text](#) | [Google Scholar](#)

192. Greenblatt RB, Oettinger M, Bohler CS. Estrogen-androgen levels in aging men and women: therapeutic considerations. *J Am Geriatr Soc* . (1976) 24: 173–8. doi: 10.1111/j.1532-5415.1976.tb04294.x

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

193. Simpson ER, Mahendroo MS, Means GD, Kilgore MW, Hinshelwood MM, Graham-Lorence S, et al. Aromatase cytochrome P450, the enzyme responsible for estrogen biosynthesis. *Endocr Rev* . (1994) 15: 342–55. doi: 10.1210/edrv-15-3-342

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

194. Blakemore J, Naftolin F. Aromatase: contributions to physiology and disease in women and men. *Physiology* . (2016) 31: 258–69. doi: 10.1152/physiol.00054.2015

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

195. Longcope C, Baker R, Johnston CC Jr. Androgen and estrogen metabolism: relationship to obesity. *Metabolism* . (1986) 35: 235–7. doi: 10.1016/0026-0495(86)90206-4

<https://assignbuster.com/androgen-regulated-cardiac-metabolism-in-aging-men/>

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

196. Xu X, Wang L, Luo D, Zhang M, Chen S, Wang Y, et al. Effect of testosterone synthesis and conversion on serum testosterone levels in obese men. *Horm Metab Res* . (2018) 50: 661–70. doi: 10. 1055/a-0658-7712

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

197. Finkelstein JW, Susman EJ, Chinchilli VM, D'Arcangelo MR, Kunselman SJ, Schwab J, et al. Effects of estrogen or testosterone on self-reported sexual responses and behaviors in hypogonadal adolescents. *J Clin Endocrinol Metab* . (1998) 83: 2281–5. doi: 10. 1210/jc. 83. 7. 2281

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)

198. Morgentaler A, Zitzmann M, Traish AM, Fox AW, Jones TH, Maggi M, et al. Fundamental concepts regarding testosterone deficiency and treatment: international expert consensus resolutions. *Mayo Clin Proc* . (2016) 91: 881–96. doi: 10. 1016/j. mayocp. 2016. 04. 007

[PubMed Abstract](#) | [CrossRef Full Text](#) | [Google Scholar](#)