

Effect oil contaminated diet with oil palm leaf



**ASSIGN
BUSTER**

EFFECT OF PRETREATMENT OF CRUDE OIL CONTAMINATED DIET WITH OIL PALMLEAF ON LIPID PEROXIDATION AND XANTHINE OXIDASE ACTIVITY IN THE KIDNEY OF RATAchuba , F. I .

Department of Biochemistry, Delta State University, PMB 1, Abraka Nigeria.

 AbstractThe toxicity of petroleum hydrocarbon across the living systems is now a common knowledge among the scientific community. What is lacking is a mini-scale antidote that can be adopted by the inhabitants of crude oil producing areas of the world. This was the reason for this study. The study was comprised forty eight female Wister rats divided into six groups of eight rats each.

The rats were fed as described thus. Group 1: ((normal Control). Group 2: feed mixed with 5. 0g oil palm leaf. Group 3: feed mixed with 10. 0g oil palm leaf.

Group 4: Feed mixed with 4ml crude oil (Crude oil Control). Groups 5 and 6: Contaminated diet mixed with ground oil palm leaf (5. 0 g and 10. 0 g respectively). At the end of exposure periods (three and six months respectively), the rats were sacrificed and the kidney used to prepare supernatant used for the determinations oxidative stress indices (lipid peroxidation and xanthine oxidase activity).

The results show that pretreatment of crude oil contaminated diet with oil palm leaf tend to restore values of lipid peroxidation and xanthine oxidase activity close to control values. Thus, it is pertinent to state that there exist potentials in the use oil palm leaf in the treatment of crude oil toxicity. And indeed setting a fresh agenda for further serious scientific

investigations Keywords: Crude oil, Kidney, Lipid peroxidation, Oilpalm, . Xanthine oxidase, 1. 0 IntroductionThe impact of crude oil spillage on the ecosystem as a result of oil explorationactivities, equipment failures, corrosion, illegal bunkering, oil theft andillicit refining are well documented 1-3.

Crude oilhas been proved to alter levels of oxidative stress markers in animals 4, 5. These oxidative stress markers include lipid peroxidation and changes in theactivities of xanthine oxidase. Lipid peroxidation precedes oxidative damage inliving organisms and alterations in the level of antioxidants represent ameasure of oxidative stress. Besides, xanthine oxidase activity is an exampleof defense mechanism as well as a measure of oxidative stress 6. Exposure topetroleum hydrocarbons is a risk factor for the impairment of renal functionvia a mechanism dependent on oxidative stress 7. Variousparts of oil palm tree can be used medicinally. The juice squeezed out frompalm leaves can be applied to enhance wound healing while the sap can be usedas laxative 8. Biologically active compounds with known antibacterial andantioxidant properties can be very useful medicinally 9, 10.

It has beenconfirmed that flavonoid, tannin and phenolic are the main phytochemicalconstituents in oil palm leaves; hence its ability to act as an effectiveantioxidant 11. Oil palm fronds extract has 8% higher contents of non-toxicantioxidative phenolic compounds than various green tea extract 12. Oil palmfrond extract may be a potential new source of functional food ingredient, based on reports of its benefit on health 13. The aim of this study was todetermine the effects of palm leave treated crude oil contaminated diet on thelevel of lipid peroxidation and xanthine oxidase activity in the <https://assignbuster.com/effect-oil-contaminated-diet-with-oil-palm-leaf/>

kidney of rats. 2. 0 Materials and methods The crude oil used for this study was obtained from Nigeria National Petroleum Corporation (NNPC) Warri, Delta State, Nigeria.

The palm frond used was obtained from *Elaeis guineensis* tree in Obiaruku, Delta state, Nigeria. Forty eight (48) female albino wistar rats with weights ranging from 0. 088kg to 0. 182 kg obtained from the animal house of Department of Anatomy, Delta State University, Abraka were used for this study. The rats were housed in a standard wooden cage made up of wire gauze, net and solid woods and left to acclimatize for one week on grower's marsh and tap water at laboratory temperature of 28C and 12 hour day/night regime. After the acclimatization period, the rats were weighed and grouped. 2.

1 Preparation of leaf powder. The leaves were isolated from the stock and sun-dried. The dried leaf was then ground with domestic kitchen blender into a fine powder and stored in a clean and sealed plastic container 2.

2 Treatment of animals The forty eight (48) female albino wistar rats were assigned to six (6) groups according to their weights, with eight rats in each group. Rats in the control group which is Group 1 were fed with grower's marsh only. Rats in Group 2 were fed with grower's marsh and 5g of powdered palm frond.

Group 3 rats were fed with grower's marsh and 10g of powdered palm leaves. Group 4 rats were fed with grower's marsh contaminated with crude oil (4ml per 100g of feed). Rats in Group 5 were fed grower's marsh

contaminated with crudeoil (4ml per 100g of feed) plus 5g of powdered palm fronds.

While rats in Group6 were fed with crude oil contaminated marsh (4ml per 100g of feed) plus 10g of powdered palm leaves. The rats in each group were allowed access to cleandrinking water while the experiment lasted. The feeds were prepared fresh dailyand stale feed remnants were discarded regularly.

The animals in each groupwere exposed to their respective diets for three and six months respectively. The National Institute of health guidefor the care and use of laboratory animals (NIH, 1985) wasadopted all through the experiment 2. 3 Collectionof samplesAfter the exposure period, all the rats weresacrificed and the kidneys were harvested. Five grams (5. 0 g) of the kidneyswere weighed in chilled conditions and homogenized with 5ml of normal saline ina mortar. The mixture was diluted with known amount ofbuffered saline before being centrifuged and the supernatant was transferred intoplastic tubes and stored at – 4C before used for analysis within forty eighthours.

2. 4 Determination of lipid peroxidation and xanthineoxidase activityThe activity ofxanthine oxidase in the kidney of rats was measured using the method of Bergmeyeret. al. 14, based on the oxidation of xanthine to uric acid, a molecule thatabsorbs light maximally at 290 nm. A unit of activity is that forming onemicromole of uric acid per minute at 25oC. Lipid peroxidationin the kidney of rats was measured by the thiobarbituric acid reacting substances TBARS, method of Gutteridge and Wilkins 15. 2.

5 Statistical Analysis Analysis of variance (ANOVA) and postHoc Fisher's test for multiple comparison was performed using statistical package for social science (SPSS), version 20 to determine statistical significant differences between means.

P values < 0.05 were taken as being significantly different. 3.0 Results and Discussion The effects of *Elaeis guineensis* leaf on kidney lipid peroxidation and xanthine oxidase activity against crude oil induced nephrotoxicity in rats after three and six months are shown in tables 1 and 2.

Lipid peroxidation in the kidney rats exposed to crude oil contaminated (group 4) was significantly ($P < 0.05$) higher in comparison with the control (group 1). Rats fed palm leaf pretreated diets (Group 2 and 3) showed significantly lower kidney levels of lipid peroxidation when compared with the control (group 4).

Moreover, rats fed crude oil contaminated diets that was pretreated with various amounts of oil palm leaves (Group 5 and 6) exhibited significantly lower kidney lipid peroxidation level when compared with the control (group 1) and rats fed crude oil contaminated diet alone (group 4). Lipid peroxidation, which is a potential marker of oxidative stress, induce disturbance of cell membrane, and functional loss of biomembranes, that results in inactivation of membrane bound receptors and enzymes [16, 17, 18]. The present study shows that the consumption of crude oil contaminated diet increased the level of lipid peroxidation in rats. This study shows that exposure to hydrocarbons present in crude oil can lead to oxidative damage of the kidney as evident by the rise in renal level of lipid peroxidation. This is

based on the premise that metabolism of hydrocarbons present in crude oil generate free radicals 19. This is in consonance with previous studies by 5, 6, 7, 20.

Oil palm (*Elaeis guineensis*) frond is rich in bioactive phytochemicals such as polyphenols and these polyphenolic compounds are considered to have antioxidant activity that is several fold higher than that of vitamins C and E 21, 22, 23. This may be the basis for the decreased level of lipid peroxidation in the kidney of rats exposed to crude oil that was treated with oil palm leaf. The kidney xanthine oxidase activities were significantly ($P < 0.05$) lower in the rats fed crude oil contaminated diets (group 4) in comparison with all the experimental groups (Tables 1 and 2). Rats fed with oil palm leaf treated crude oil contaminated diet (Groups 5 and 6) have significantly higher xanthine oxidase activities in the kidney when compared with rats fed with crude oil contaminated diet only (group 4). However, rats fed with only oil palm leaf treated diets (Groups 2 and 3) have significantly higher kidney xanthine oxidase activity when compared with rats fed with only crude oil contaminated diet (group 1).

Xanthine oxidase is involved in phase one process in the inactivation of xenobiotics in animals 24. The increase in the activity of xanthine oxidase in rats exposed to oil palm leaf treated diet indicates response of the enzyme to enhance the metabolism of endogenous xanthine. This is in a bid to increase the production of uric acid, a potent antioxidant 7, 24, 25. The decrease in activity of xanthine oxidase in rats exposed to crude oil contaminated diet alone shows that the metabolism of crude oil hydrocarbons leads to a reduced ability to produce uric acid. Nevertheless, the increase in the activity

of oxidative enzymes had been reported as a measure of oxidative stress 26. However, addition of ground oil palm leaves resulted in decrease in toxic effects of crude oil. This is exhibited in the decrease in xanthine oxidase activity in rats fed with crude oil contaminated diets that were pretreated with oil palm leaves. This is due to the ability of oil palm leaves to act as an antioxidant, protecting endothelial cells of the kidney against reactive free radicals thereby decreasing the level of antioxidant enzymes 11, 13. Substances with antioxidant potentials possess health promoting properties, since they quench free radicals which are involved in many disease processes 13, 27, 28, 29. This study has indicated that the consumption of crude oil contaminated diet can result in increase in oxidative stress which causes corresponding increases in lipid peroxidation levels and xanthine oxidase activity.

However, the crude oil toxicities were reversed by the consumption of diets that were pretreated with oil palm leaves. This study, therefore, shows the protective role of oil palm leaf against crude oil induced nephrotoxicity. References 1. Otitoju O, Onwurah . INE (2007) Preliminary investigation into the possible endocrine disrupting activity of bonny light crude oil contaminated diet on wistar rats. *Biokemistri J.* 19(2): 23-282. Ovuru SS, Ekweozor IKE.

(2004) Haematological changes associated with crude oil ingestion in experimental rabbits. *Afr. J. Biotechnol.*

3(6): 346-3483. Ogudu AD, Esemuede IH. (2013).

Crude oil theft and its environmental consequences: The way forward. J. Nig. Environ. Society.

7(4): 1-184. Achuba FI; Osakwe SA (2003) Petroleum Induced Free Toxicity in African Catfish (*Clarias garieponus*). Fish Physiol. Biochem. 29: 97-1035.

Anozie OI, Onwurah IN (2001) Toxic Effects of Bonny Light Crude oil on Rats after Ingestion of contaminated diet. Nig. J. Biochem. Mol. Biol. 16: 1035-10856. Achuba FI (2014) Petroleum Products in Soil Mediated Oxidative Stress in Cowpea (*Vigna unguiculata*) and Maize (*Zea mays*) Seedlings.

Open J. Soil Sci. 4: 417-435. 7. Azeez OM, Akhigbe RE, Anigbogu CN (2013) Oxidative status in rat kidney exposed to petroleum hydrocarbons. J.

Nat. Sci. Biol. Med. 4(1): 149-1548. Sasidharan S, Logeswaran S, Latha LY (2012) Wound healing activity of *Elaeis guineensis* leaf extract ointment. Int.

J. Mol. Sci.

13: 336-3479. Chong KH, Zuraini Z, Sasidharan S, Devi PVK, Latha LY, Ramanathan S (2008). Antimicrobial activity of *Elaeis guineensis* leaf. Pharmacology online. 3: 379-38610. Rout SP, Choudary KA, Kar DM, Das L, Jain A (2009). Plants in traditional medicinal system-future source of new drugs. Int.

J. Pharm. Pharmaceutical Sci. 5(4): 137-14011. Phin KC, Syahriel A., Ng, SY (2013) Phytochemical constituents from leaves of *Elaeis guineensis* and their antioxidant and antimicrobial activities. Int. J.

Pharm. Pharmaceutical Sci. 5(4)137-14012. Runnie I., Nordin MM, Radzali M, Azizah H, Hapizah N (2003) Antioxidant and hypocholesteromic effects of *Elaeis guineensis* leaves extract on hypercholesteromic rabbits. ASEAN Food J.

12: 137-14713. Mohamed S (2014) Oil Palm Leaf: A New Functional Food Ingredient for Health and Disease Prevention. J.

Food Process Technol. 5(2): 300-30614. Bergmeyer HV, Gacoehm K, Grassl M (1974) In: Methods of Enzymatic Analysis, HV Bergmeyer (eds). New York: Academic Press. vol. 2 p. 428-429. 15.

Guttridge JMC, Wilkins C (1982) Copper-dependent hydroxyl radical damage to ascorbic acid. Formation of thiobarbituric acid reactive products. FEBS Lett. 137: 327-340. 16.

Halliwell B (1994) Free radicals and antioxidants: a personal view. Nutr. Rev. 5: 253-265. 17. Niki E (2008). Lipid peroxidation products as oxidative stress biomarkers. Biofactors.

34(2): 171-18018. Greenberg ME, Li XM, Giugiu BG, Gu X., Qin J, Salomon RG, Hazen S. (2008) The lipid whisker model of the structure of oxidized cell membranes. J. Biol Chem.

283: 2385-239619. Achuba FI (2010) Spent engine oil mediated oxidative stress in cowpea (*Vigna unguiculata*) seedlings. *EJEA FChE*.

9(5): 910-917 20. Alisi CS., Ojiako AO., Osuagwu CG, Onyeze GOC (2011) Response pattern of antioxidants in carbon tetrachloride-induced hepatotoxicity is tightly logistic in rabbits. *Eur J Med Plants*. 1: 118-12921. Cowan MM (1999) Plant products as antimicrobial agents. *Clin. Microbiol. Rev.* 12(4): 564-58222. LeeYL, Jian SY, Lian PY, Mau JL. (2008) Antioxidant properties of extract from a white mutant of the mushroom *Hypsizygus marmoreus*. *J. Food Compos. Anal.* 21: 116-12423. Jaffri JM, Mohamed S, Ahmad IN, Mustapha NM, Manap YA, Rohimi N (2011) Effects of catechin-rich oil palm leaf extract on normal and hypertensive rats kidney and liver. *Food. Chem.* 128: 433-44124.

Ezedom T, Asagba SO (2016) Effect of a controlled food-chain mediated exposure to cadmium and arsenic on oxidative enzymes in the tissues of rat *Toxicology Reports* (3) : 708-71525. Achuba FI (2008). African land snail *Achatina marginatus*, as bioindicator of environmental pollution. *North-Western Journal of Zoology* 4 (1): 1-526.

Förstermann U, Xia N, Li H {2017} Roles of Vascular Oxidative Stress and Nitric Oxide in the Pathogenesis of Atherosclerosis. *Circulation Research*. 120: 713-73527. Hybertson BM, Gao, B, Bose, SK., McCord JM (2011) Oxidative Stress in health and disease: The therapeutic potential of Nrf2 activation. *Mol Asp Med* 32(4): 234-24628.

<https://assignbuster.com/effect-oil-contaminated-diet-with-oil-palm-leaf/>

Galli F, Piroddi M., Annetti C, Aisa C, Floridi E., Floridi A (2005) Oxidative stress and reactive oxygen species. *Contrib Nephrol* 149: 240-26029.

Barnham KJ, Masters CL, Bush AI (2004). Neurodegenerative diseases and oxidative stress.

Nat Rev Drug Discovery. 3: 205-214 Table 1. The effect of *Elaeis guineensis* leaf on the level of lipid peroxidation and xanthine oxidase activity in the kidney of rats after three months of exposure to crude oil contaminated diet. Groups Lipid peroxidation (nmol/g tissue) Xanthine oxidase activity (units/g tissue) Group 1 0.35 ± 0.05 a 60.04 ± 4.

28 a Group 2 0.14 ± 0.02 b 60.83 ± 1.76 a Group 3 0.10 ± 0.03 b 69.

28 ± 3.34 b Group 4 0.76 ± 0.10 c 42.43 ± 1.78 c Group 5 0.

52 ± 0.01 d 51.09 ± 2.70 d Group 6 0.

34 ± 0.01 a 57.05 ± 5.89 a Each value represents mean ± standard deviation. n = 4 in each group. Values not sharing a common superscript letter in the same column differ significantly at (P < 0.05).

Group 1: ((Normal Control). Group 2: feed mixed with 5.0g oil palm leaf.

Group 3: feed mixed with 10.0g oil palm leaf. Group 4: Feed mixed with 4ml crude oil (Crude oil Control).

Group 5: Contaminated diet mixed with 5.0 g of oil palm leaf. Group 6: contaminated diet mixed with 10.0 g of oil palm leaf. Table 2.

The effect of *Elaeis guineensis* leaf on the level of lipid peroxidation and xanthine oxidase activity in the kidney of rats after six months of

exposure to crude oil contaminated diet. Groups Lipid peroxidation (nmol/g tissue) Xanthine oxidase activity (units/g tissue) Group 1 0.42 ± 0.08 a 62.04 ± 3.80 a Group 2 0.22 ± 0.01 b 61.41 ± 2.64 a Group 3 0.11 ± 0.04 b 68.24 ± 2.22 b Group 4 0.89 ± 0.11 c 38.43 ± 2.66 c Group 5 0.66 ± 0.12 d 54.11 ± 3.50 d Group 6 0.53 ± 0.06 a 55.44 ± 6.70 a

Each value represents mean ± standard deviation. n = 4 in each group.

Values not sharing a common superscript letter in the same column differ significantly at (P < 0.05). Group 1: (Normal Control). Group 2: feed mixed with 5.0g oil palm leaf. Group 3: feed mixed with 10.0g oil palm leaf. Group 4: Feed mixed with 4ml crude oil (Crude oil Control). Group 5: Contaminated diet mixed with 5.0 g of oil palm leaf. Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.

Group 6: contaminated diet mixed with 10.0 g of oil palm leaf.