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0013-7227/01/\$03. 00/0 Printed in U. S. A. Endocrinology 142(10): 4163—4169 Copyright © 2001 by The Endocrine Society Minireview: Ghrelin and the Regulation of Energy Balance—A Hypothalamic Perspective TAMAS L. HORVATH, SABRINA DIANO, PETER SOTONYI, MARK HEIMAN, " MATTHIAS TSCHOP AND Reproductive Neuroscience Unit (T. L. H., S. D.), Department of Obstetrics and Gynecology and Department of Neurobiology (T. L. H.), Yale Medical School, New Haven, Connecticut 06520; Department of Anatomy and Histology (P. S.), Faculty of Veterinary Science, Szent Istvan University, Budapest, Hungary 1400; and Lilly Research Laboratories (M. H., M. T.), Eli Lilly & Co., Indianapolis, Indiana 46285 The recently discovered hormone, ghrelin, has been recognized as an important regulator of GH secretion and energy homeostasis. Orexigenic and adipogenic ghrelin is produced by the stomach, intestine, placenta, pituitary, and possibly in the hypothalamus. The concentration of circulating ghrelin, principally derived from the stomach, is influenced by acute and chronic changes in nutritional state. To date, most studies focused on the role of ghrelin in GH secretion or its function in complementing leptin action to prevent energy deficits. The potential significance of ghrelin in the etiology of obesity and cachexia as well as in the regulation of growth processes is the subject of ongoing discussions. A large quantity of information based on clinical trials and experimental studies with ghrelin and previously available synthetic ghrelin receptor agonists (GH secretagogues) must now be integrated with a rapidly increasing amount of data on the central regulation of metabolism and appetite. In this overview, we summarize recent findings and strategies on the integration of ghrelin into neuroendocrine networks that regulate energy

homeostasis. (Endocrinology 142: 4163– 4169, 2001) OBESITY AND RELATED disorders are among the leading causes of illness and mortality in the developed world (1). To better understand the pathophysiological mechanisms that underlie metabolic disorders, increasing attention has been paid to central regulatory elements in energy homeostasis, including food intake and energy expenditure (2–5). The past two decades have provided overwhelming evidence of the critical role that hypothalamic peptidergic systems play in the central regulation of appetite and metabolism (6, 7). The discovery of ghrelin (8 –11) and its influence on appetite, fuel utilization, body weight, and body composition that is complementary to ghrelin's GH-releasing effect (12) adds yet another component to the complexity in the central regulation of energy balance. Discovery of Ghrelin Reverse pharmacology may be an appropriate term to describe the road to ghrelin's discovery. First, synthetic agonists with ghrelin-like activity [GH-releasing peptides (GHRPs) and GH secretagogues (GHSs)] were discovered by Bowers and co-workers in the late seventies (9, 13–15), followed by the cloning of ghrelin-GHS-receptor (GHS-R) in 1996 by Smith and co-workers (16 –18). Subsequently, the elegant studies by Kojima and co-workers led to the identification of an acylated 28 residue peptide as an endogenous bioactive ligand for the GHS-R (8 –11). It was called ghrelin, a term that contains "ghre-" as the etymological root for "growth" in many languages. "GH" and "relin," a suffix for releasing substances in generic names according to the USP Abbreviations: AGRP, Agouti-related protein; GHRP, GH-releasing peptide; GHS, GH-secretagogue; GHS-R, GHS receptor. Dictionary of USAN and International Drug Names, also represents an abbreviation for "growth-

hormone-release, " a characteristic effect of ghrelin (8, 11). The Ser3-acylation that seems to be responsible for bioactivity of ghrelin is a modification that has been observed for the first time in mammalian physiology. There are no data to support the tempting speculation that the purpose of this modification is to increase ghrelin's lipophilic properties to facilitate transport across the blood brain barrier. However, the octanoyl side chain is essential for binding and activation of the GHS-R subtype-1a in vitro (19). Ghrelin might also bind to different GHS-R subtypes or receptor families where the octanoyl side chain is not needed. Detection and purification of the gastric enzyme responsible for the acylation of ghrelin may shed light on this fascinating question and will possibly even reveal the existence of other putative hormones carrying this modification. Sources of Ghrelin Ghrelin is predominantly produced by the stomach (8, 20 –22), whereas substantially lower amounts are derived from bowel (21, 22), pituitary (23), kidney (24), placenta (25), and hypothalamus (8, 22). Although the majority of circulating ghrelin is produced in the stomach, other sources may increase ghrelin secretion in a compensatory manner. After gastrectomy, for example, plasma ghrelin level is surprisingly reduced only by 65% (26). One of the most urgent and debatable hypotheses is whether ghrelin is produced in physiologically relevant amounts in the hypothalamus. This unresolved issue is the focus of several research groups. Data published or presented at recent meetings, in accordance with our own on- 4163 4164 *Endocrinology*, October 2001, 142(10): 4163— 4169 Horvath et al. - Ghrelin and Regulation of Energy Balance going investigations, have shown ghrelin to be present in several regions of the hypothalamus through the use of

immunohistochemical detection methods (8, 22). Depending on the ghrelin epitope recognized by the antibody in use, ghrelin-positive cells have been identified in varying hypothalamic areas, whereas all of these antisera have been successfully used in RIAs. Detection of hypothalamic ghrelin mRNA by use of PCR has been accomplished (8); however, this issue needs to be further investigated by the detection and regional distribution of ghrelin mRNA by in situ hybridization. At this point, it is not clear if the detection of ghrelin in the hypothalamus using immunohistochemistry reflects ghrelin peptide that is produced by hypothalamic neurons or ghrelin peptide that is derived from the stomach. It seems logical that gastric ghrelin reflects an acute nutritional state. However, even minimal ghrelin expression in the hypothalamus (8, 12) or circulating placental ghrelin during pregnancy (25) may significantly influence food intake, nutrition partitioning, and fat utilization. Regardless of the source, in the end it is most likely the modulation of hypothalamic circuits by ghrelin that mediates changes in energy homeostasis. Ghrelin and GH Secretion Based on rodent experiments (27–29) and clinical studies (30–33), it is evident that ghrelin is indeed a potent GH-releasing agent. However, no significant correlation seems to exist between plasma ghrelin concentrations and circulating levels of GH or IGF-I (unpublished data, Tschöp et al.) even though both ghrelin and GH increase during fasting (34, 35). Very recent data indicate that most of the ghrelin-induced GH secretion is not only directly opposed by somatostatin action, but also involves mediation through GHRH (33, 36, 37). However, ghrelin also releases GH in vitro from primary rat pituitary cells (8, 12), and GHRP-2, a potent ghrelin receptor agonist, releases GH in vivo in patients with GHRH

receptor mutations (38). This indicates the existence of GHRH-independent effects of ghrelin on GH secretion mediated by hypophyseal GHS-Rs, which were originally cloned from the pituitary (16). Alternatively, ghrelin may stimulate an unidentified hypothalamic agent (U-factor) that, in turn, stimulates GH release (39).

### Ghrelin and Energy Balance

The first published evidence for the involvement of ghrelin in the regulation of appetite was provided by Ghigo and co-workers (30). They described that 3 out of 4 healthy volunteers spontaneously reported hunger following ghrelin administration as a “side effect” in a clinical study analyzing GH release (30). This hunger-inducing effect of ghrelin has now been confirmed in two more studies, where, again, 3 out of 7 (33) and 9 out of 11 individuals report hunger as the only sensation after ghrelin injection (40). A large number of animal studies added strength to the argument that ghrelin is involved in the regulation of energy balance. For example, exogenous ghrelin induces adiposity in rodents by stimulating an acute increase in food intake, as well as a reduction in fat utilization (12, 41–46). Adipogenic as well as orexigenic effects of ghrelin are independent from its ability to stimulate GH secretion (12, 46) and are most likely mediated by a specific central network of neurons that is also modulated by leptin (2–7, 9, 12, 41–46). Regulation of ghrelin secretion, as well as its biological effects, appear to be opposite those of leptin. However, from a teleological point of view, ghrelin and leptin might really be complementary players of one regulatory system that has developed to inform the central nervous system about the current status of acute and chronic energy balance (12, 38–49). In addition, a specific role for ghrelin might be to ensure the provision of calories that GH requires for

growth and repair (41). In humans, circulating ghrelin levels are decreased in chronic (obesity) (48) and acute (caloric intake) (26, 34, 47) states of positive energy balance, whereas plasma levels of ghrelin are increased by fasting (12, 34) and in cachectic patients with anorexia nervosa (26). Of course, it has yet to be proven that the rather modest changes in circulating ghrelin, in the 100 fmol range, have physiological relevance for hypothalamic receptor sites. One plausible explanation is that if ghrelin is indeed a hormone signaling the need to conserve energy (12), ghrelin secretion is triggered to counter further deficit of energy storage and to prevent starvation or cachexia. A very recent study shows a pre-meal rise of human plasma ghrelin, suggesting a possible role of ghrelin as a hunger signal triggering meal initiation (34). In rodents, fasting and hypoglycemia increase ghrelin levels, whereas intake of food, especially carbohydrates (dextrose), decreases ghrelin secretion (12, 41, 50). We speculate that this obvious connection between glucose levels, ghrelin secretion and GH secretion is likely to be involved in the physiological mechanism of diagnostic procedures such as oral glucose tolerance testing (for acromegaly) and insulin tolerance testing (for GH deficiency). Differential effects of ghrelin might be mediated by separate ghrelin (GHS-R) subtypes as recently suggested by Thorner and co-workers (51). Based on a series of elaborate studies using GHS-R antagonists ([d-Lys3]GHRP-6 and BMS-265711, also an NPY-antagonist) and an NPY-Y1-R antagonist ([d-Trp32]NPY), they showed that the orexigenic effect of ghrelin can be dissociated from its GH releasing effects, suggesting distinct GHS-R-subtypes. Based on the observation of differential orexigenic effects of hexarelin and its analogs and GH secretagogue actions at the

pituitary gland (52, 53), the existence of additional subtypes of the GHS-R (16 —18) had previously been hypothesized. The putative adipogenic effects of ghrelin in humans remains to be shown because it is possible that ghrelin has different effects on energy balance in humans and rodents. In addition, ghrelin-induced adiposity could be only a transient effect and the therapeutic potential of ghrelin in cachectic humans might therefore turn out to be as disappointing as the efficacy of leptin for the therapy of human obesity (5, 54). Carefully conducted clinical studies focusing on body composition as well as long-term studies on ghrelin treatment in rodents are necessary to further address this question.

Ghrelin and Brain Centers of Energy Balance  
Our current understanding of the involvement of different hypothalamic systems in metabolic regulation arises from early degeneration studies in rats. Destruction of distinct Horvath et al. - Ghrelin and Regulation of Energy Balance *Endocrinology*, October 2001, 142(10): 4163— 4169 4165 hypothalamic regions, particularly the ventromedial nucleus but also the areas of the paraventricular and dorsomedial nuclei, induced hyperphagia (55— 60). In contrast, discrete lesions placed in the lateral hypothalamus (61, 62) reduced food intake. During the last two decades, a substantial amount of research demonstrated that NPY, administered into the cerebral ventricles (63) or other specific hypothalamic sites (64), induced food intake. However, in addition to NPY, several other hypothalamic peptides were found to affect appetite and feeding behavior (for details see Refs. 2—7). Appetite-stimulating neuropeptides include melanin concentrating hormone, hypocretins/orexins (produced in a distinct subset of neurons of the lateral hypothalamus perifornical region) (65— 68) and agouti-related protein



(AGRP, coproduced with NPY in the same arcuate nucleus neurons) (69 — 71). Appetite-suppressing neuropeptides include the POMC derivative,  $\alpha$ -MSH (6, 7, 72) that is produced in arcuate nucleus perikarya (73). An important milestone to link the central regulation of metabolism with peripheral levels of energy storage was the discovery of the adipose hormone, leptin. Genetic mouse or rat mutants, including db/db and ob/ob mice and fa/fa rats become strikingly obese. Molecular analysis has shown that the primary genetic defect in these animals relates to either abolished leptin production (ob/ob mice) or impaired leptin receptors (db/db mice; fa/fa rats; leptin-R) (5, 74 — 77). Similar examples of obesity in humans have been found and are associated with a mutation of leptin or the leptin-receptor (78 — 80). Leptin is released by adipose tissue and has been suggested to be the key-signal reflecting adipose stores. Leptin receptors are found in the hypothalamus, particularly in the arcuate nucleus where leptin is thought to exert its primary feedback signaling (81— 87). Recent experiments in rodents and primates have been attempting to tie together the diverse hypothalamic peptidergic systems with hormone receptors, including leptin receptors, to decipher the hypothalamic signaling modality underlying the regulation of daily energy homeostasis (81—92). A schematic illustration of some of these interactions and the way ghrelin signaling may be integrated into these circuits is shown on Fig. 1. Peripheral ghrelin is mainly produced in the gastrointestinal tract (8, 10, 22—24). It reaches ghrelin-receptors in the anterior pituitary and potentially in the mediobasal and mediolateral hypothalamus through the general circulation to stimulate GH release and to regulate energy homeostasis (12). It remains to be determined whether

circulating ghrelin can reach brain areas outside of the blood brain barrier only, such as the ventromedial arcuate nucleus (93), or it has the ability to target areas protected by the blood brain barrier. Areas protected by the blood brain barrier include most hypothalamic nuclei and the rest of the brain (93). Ghrelin-containing cells are also present in the mediobasal hypothalamus, where GHRH cells and the neuronal network that regulates energy balance are located (8, 22). Detailed phenotypes and macroscopic connectivity of different hypothalamic networks regulating metabolism have been described by numerous recent outstanding reviews (2–7). Among hypothalamic peptidergic circuits, particular significance is attributed to the arcuate nucleus opiate neurons that produce  $\alpha$ -MSH, a main anorexigen and energy expenditure enhancer (72), and to its interrelationship with another group of arcuate nucleus neurons that produce both NPY and an endogenous antagonist of  $\alpha$ -MSH, AGRP (69). The interaction between these two distinct populations of cells is currently considered as a *primum movens* in the regulation of energy homeostasis. However, there are other peptidergic circuits within the hypothalamus, including the lateral hypothalamic orexin/hypocretin- and melanin concentrating hormone-producing cells, that appear to respond to peripheral metabolic signals and alter food intake as well as energy expenditure (65–68). In light of the aforementioned excellent reviews (2–7), we will avoid an in-depth description of these peptidergic systems here but will attempt to emphasize a better appreciation of the neuronal doctrine for the integration of emerging experimental data on ghrelin. In the brain, receptors for ghrelin were detected in multiple hypothalamic nuclei as well as in the hippocampus, substantia nigra, ventral

tegmental area, and dorsal and median raphe nuclei (8, 94 —98). In a series of experiments, Dickson and co-workers, first using synthetic GHS-R agonist, and then ghrelin, provided evidence that this novel metabolic hormone, in fact, interacts with the aforementioned hypothalamic peptidergic systems in the central regulation of metabolism (99 —103). For example, they found that following central ghrelin administration, c-fos, an early proto-oncogen that reflects cellular activity, is induced in the medial arcuate nucleus where NPY/AGRP cells are located (103). It was also shown that Y1-receptor antagonists as well as melanocortin agonists and antisera to both NPY and AGRP may interfere with ghrelin's feeding-inducing effect (42, 43, 46). However, absence of NPY in genetically engineered NPY-ko mice does not diminish ghrelin-induced feeding or adiposity suggesting a key-role for AGRP in the mediation of ghrelin's effects on energy balance (12). The effect of ghrelin on metabolism seems to be the exact opposite to that of leptin (2—7, 9, 10, 12). In obesity, when plasma leptin levels are elevated, ghrelin plasma levels are decreased indicating physiological adaptations to the positive energy balance rather than an involvement in the etiology of obesity (48, 49). Of course, it is important to note that, while ghrelin is regulated acutely like a satiety factor, leptin levels are not regulated by meals, but rather by actual increase in adipose stores. Ghrelin's Hypothalamic Signaling Requires Synapses Figure 1 depicts a highly complex interaction between a variety of hypothalamic peptidergic systems, including the putative ghrelin network, in the central regulation of energy balance. It has to be noted, however, that this drawing is not all-inclusive and represents only the " tip of the iceberg. " There are many more hypothalamic and extra-hypothalamic

neurotransmitters and neuropeptides that act via the aforementioned circuits (for example coexistence of GABA with NPY; 91) or in separate pathways [for example, ciliary neurotrophic growth factor (CNTF), 104, 105], and are interconnected with the illustrated systems (for further review see Refs. 2, 6, 7). In addition, receptors for the different neuropeptides as well as for peripheral hormones that affect metabolism, including insulin, thyroid hormones, gonadal steroids and glucocorticoids, are also present in these re-

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FIG. 1. Schematic representation of the interaction between key hypothalamic peptidergic systems in the central regulation of daily energy homeostasis and their relationship to peripheral and putative hypothalamic ghrelin. Ghrelin, a hunger signal, is released from the stomach into the circulation and may be produced (?) in a subset of hypothalamic neurons (red). Leptin, a satiety signal, is released from white adipose tissue (WAT) into the circulatory system. Ghrelin (red arrows) and leptin (orange arrows) directly target the hypothalamus and brain stem areas. While brain stem areas on this drawing are illustrated as efferent targets of hypothalamic circuits, critical pathways exist from the brain stem to the hypothalamus, as well, that can mediate ascending ghrelin and leptin signaling. AGRP is produced in NPY cells (yellow) and acts to block the inhibitory action of the POMC derivative,  $\alpha$ -MSH (green), on feeding. Both AGRP/NPY and POMC cells are apparent targets of direct ghrelin action via GHS-R (\*). The NPY neurons that receive lateral hypothalamic input, including HCRT (brown) and melanin concentrating hormone (MCH) (blue) innervation, project to a number of regions of the brain, particularly those

implicated in feeding mechanisms, including the paraventricular nucleus (PVN), lateral hypothalamus, LH, ventromedial nucleus (VMH), perifornical region (PF), and dorsomedial nucleus (DMH). The same regions also receive direct lateral hypothalamic input as well as innervation from -MSH cells. These regions, in turn, project (large black arrow) widely throughout the brain to loci including the medial thalamic nuclei (MT), central gray (cg), dorsal motor nucleus of the vagus (DMV), cortex, nucleus of the solitary tract (NTS), locus coeruleus (LC), spinal cord, and amygdala. Ghrelin-targeted arcuate nucleus neurons may also affect neuroendocrine cells that are responsible for the regulation of pituitary hormone secretions, including gonadotrophs (LH/FSH), TSH, ACTH, and GH. It is yet to be determined what role central vs. peripheral ghrelin plays in the regulation of this circuitry and at what sites and subcellular levels ghrelin signaling is interacting with that of leptin. During food deprivation when leptin levels rapidly decline (106) and NPY/AGRP production is elevated, but POMC neurons are suppressed (106 –110), circulating ghrelin levels increase (12, 33, 41, 48) suggesting that leptin and ghrelin coregulate hypothalamic peptidergic systems in opposite ways. These observations further support the hypothesis that ghrelin, as a " hunger signal, " is the counterpart of leptin aiming to prevent further energy deficit. However, considering the extreme complexity of hypothalamic interactions of different peptidergic circuits and peripheral hormone receptors, it is necessary to determine the hierarchy and direction of signaling flow within these systems to understand ghrelin's central effect on metabolic regulation. For that, a multidisciplinary approach is mandatory. The hypothalamus is composed of a complicated set of regulatory neurons

that in most cases cannot be identified by traditional means of cell segregation, i. e. location, soma size, or dendritic arbor. Therefore, to identify specific types of neurons, cytochemistry must be used. In addition, as in all other brain areas, the primary mode of communication between hypothalamic peptidergic circuits is via synapses. The only reliable way for assessing synapses is by the use of conventional electron microscopy and electrophysiology because proximity of different cells assessed by light microscopy is not a convincing indicator of neuronal interaction. Thus, determination of the qualitative and quantitative synaptological relationship between GHS-Rs, ghrelin-producing neurons, and other key hypothalamic peptidergic systems and their receptors will be an important step for gaining insight into the hypothalamic signaling modality of ghrelin. Of course, the anatomical experiments alone will not be sufficient to determine the actual involvement of the pre- Horvath et al. - Ghrelin and Regulation of Energy Balance *Endocrinology*, October 2001, 142(10): 4163— 4169 4167 synaptic ghrelin system in the regulation of the postsynaptic circuit, but provides an invaluable map that is necessary for the correct interpretation of data gathered with other tools. In fact, anatomical studies need to be complemented by parallel electrophysiological analyses. An elegant example of such an approach is the recent work by Cowley and colleagues (111), in which leptin's effect was analyzed on genetically tagged arcuate nucleus - MSH cells and the qualitative synaptology of these cells was simultaneously assessed. That approach not only eliminated the pitfalls of the individual experimental techniques but immediately provided a more comprehensive view on a given hypothalamic neuronal system (111). The significance in

determining the spatial relationship between different afferents using anatomical and electrophysiological tools may further be appreciated when one considers that a synapse is more potently able to affect postsynaptic cells when located proximally either on the cell body or postsynaptic dendrite than when it is located more distally. In addition, both previous electrophysiological (111–113) and morphological observations (92, 111) indicate that an extensive interaction exists between presynaptic terminals to affect hypothalamic cells both in the arcuate nucleus where NPY/AGRP and -MSH cell bodies are located and in a model efferent target, the parvocellular paraventricular nucleus. The impact of ghrelin on arcuate and parvocellular paraventricular nucleus neurons will be readily dependent on their synaptic organization on the postsynaptic cells and their interaction with other systems presynaptically. One of the best examples to illustrate this synaptologic interaction is the relationship between the NPY/AGRP and -MSH systems. Electrophysiological and anatomical observations pointed to both the arcuate and paraventricular nuclei as primary sites for the interplay between AGRP and -MSH systems (111, 114). Because ghrelin's action appears to be mediated by the NPY/AGRP system, it is not unlikely that ghrelin will act in the arcuate nucleus as well as in the paraventricular nucleus to modulate the interaction between NPY/AGRP and -MSH. It may be that peripheral and central ghrelin contribute equally to the regulation of both of these hypothalamic areas, but it is also conceivable that stomach-derived ghrelin affects the arcuate nucleus where the blood-brain barrier is less effective, whereas hypothalamic ghrelin is more involved in the modulation of hypothalamic sites within the blood-brain barrier, such as the

paraventricular nucleus. An alternative and equally feasible pathway for ghrelin signaling from the stomach is via an ascending neural network through the vagus nerve and brain stem nuclei that ultimately reaches the hypothalamus (43). When electrophysiological and anatomical techniques are combined with conventional physiological and molecular biological approaches, as well as with the very recently developed revolutionary tracing technique of DeFalco et al. [(115) which allows tracing of inputs of chemically identified subpopulations of neurons], it is reasonable to expect that not only a thorough understanding of ghrelin's action will be achieved at a faster pace, but great advances will be made toward the general understanding of the hypothalamic machinery in metabolism regulation.

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responses to leptin. *Nature Neuroscience* 6: 445— 450 5. Friedman JM, Halaas JL 1998 Leptin and the regulation of body weight in mammals. *Nature* 395: 763—770 6. Ahima RS, Osei SY 2001 Molecular regulation of eating behavior: new insights and prospects for therapeutic strategies. *Trends Mol Med* 7: 205—213 7. Spiegelman BM, Flier JS 2001 Obesity and the regulation of energy balance. *Cell* 104: 531—543 8. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K 1999 Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 402: 656 — 660 9. Bowers CY 2001 Unnatural growth hormone-releasing peptide begets natural ghrelin. *J Clin Endocrinol Metab* 86: 1464 —1469 10. Kojima M, Hosoda H, Matsuo H, Kangawa K 2001 Ghrelin: discovery of the natural endogenous ligand for the growth hormone secretagogue receptor. *Trends Endocrinol Metab* 12: 118 — 122 11. Hosoda H, Kojima M, Matsuo H, Kangawa K 2000 Purification and characterization of rat des-Gln14-Ghrelin, a second endogenous ligand for the growth hormone secretagogue receptor. *J Biol Chem* 275: 21995—2000 12. Tschop M, Smiley D, Heiman ML 2000 Ghrelin induces adiposity in rodents. *Nature* 407: 908 —913 13. Momany FA, Bowers CY, Reynolds GA, Chang D, Hong A, Newlander K 1981 Design, synthesis, and biological activity of peptides which release growth hormone in vitro. *Endocrinology* 108: 31—39 14. Bowers CY, Momany F, Reynolds GA, Chang D, Hong A, Chang K 1980 Structure-activity relationships of a synthetic pentapeptide that specifically releases growth hormone in vitro. *Endocrinology* 106: 663—667 15. Bowers CY 1998 Growth hormone-releasing peptide (GHRP). *Cell Mol Life Sci* 54: 1316 —1329 16. Howard AD, Feighner SD, Cully DF, et al. 1996 A receptor in pituitary and hypothalamus that functions in growth hormone

release. *Science* 273: 974 —977 17. Smith RG, Pong SS, Hickey G, et al. 1996 Modulation of pulsatile GH release through a novel receptor in hypothalamus and pituitary gland. *Rec Prog Horm Res* 51: 261—286 18. Smith RG, Van der Ploeg LH, Howard AD, et al. 1997 Peptidomimetic regulation of growth hormone secretion. *Endocr Rev* 18: 621— 645 19. Bednarek MA, Feighner SD, Pong SS, et al. 2000 Structure-function studies on the new growth hormone-releasing peptide, ghrelin: minimal sequence of ghrelin necessary for activation of growth hormone secretagogue receptor 1a. *J Med Chem* 43: 4370 — 4376 20. Dornonville de la Cour C, Bjorkqvist M, Sandvik AK, et al. 2001 A-like cells in the rat stomach contain ghrelin and do not operate under gastrin control. *Regul Pept* 99: 141—150 21. Date Y, Kojima M, Hosoda H, et al. 2000 Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. *Endocrinology* 141: 4255— 4261 22. Kagotani Y, Sakata I, Yamazaki M, Nakamura K, Hayashi Y, Kangawa K 2001 Localization of ghrelin-immunopositive cells in the rat hypothalamus and intestinal tract. *Proceedings of the 83rd Annual Meeting of The Endocrine Society, Denver, CO, 2001, p. 337* 23. Korbonits M, Kojima M, Kangawa K, Grossman AB 2001 Presence of ghrelin in normal and adenomatous human pituitary. *Endocrine* 14: 101—104 24. Mori K, Yoshimoto A, Takaya K, et al. 2000 Kidney produces a novel acylated peptide, ghrelin. *FEBS Lett* 486: 213—216 25. Gualillo O, Caminos J, Blanco M, et al. 2001 Ghrelin, a novel placental-derived hormone. *Endocrinology* 142: 788 —794 26. Ariyasu H, Takaya K, Tagami T, et al. 2001 Plasma ghrelin levels are in- 4168 *Endocrinology*, October 2001, 142(10): 4163— 4169 Horvath et al. - Ghrelin and Regulation of Energy Balance 27.

28. 29. 30. 31. 32. 33. 34. 35. 36. 37. 38. 39. 40. 41. 42. 43. 44. 45. 46. 47.  
48. 49. 50. 51. 52. 53. influenced by acute and chronic feeding states in humans. Proceedings of the 83rd Annual Meeting of The Endocrine Society, Denver, CO, 2001, p 336 Date Y, Murakami N, Kojima M, et al. 2000 Central effects of a novel acylated peptide, ghrelin, on growth hormone release in rats. *Biochem Biophys Res Commun* 275: 477— 480 Seoane LM, Tovar S, Baldelli R, et al. 2000 Ghrelin elicits a marked stimulatory effect on GH secretion in freely-moving rats. *Eur J Endocrinol* 143: R7—R9 Tolle V, Zizzari P, Tomasetto C, Rio MC, Epelbaum J, Bluet-Pajot MT 2001 In vivo and in vitro effects of ghrelin/motilin-related peptide on growth hormone secretion in the rat. *Neuroendocrinology* 73: 54 — 61 Arvat E, Di Vito L, Broglio F, et al. 2000 Preliminary evidence that Ghrelin, the natural GH secretagogue (GHS)-receptor ligand, strongly stimulates GH secretion in humans. *J Endocrinol Invest* 23: 493— 495 Peino R, Baldelli R, Rodriguez-Garcia J, et al. 2000 Ghrelin-induced growth hormone secretion in humans. *Eur J Endocrinol* 143: R11—R14 Takaya K, Ariyasu H, Kanamoto N, et al. 2000 Ghrelin strongly stimulates growth hormone release in humans. *J Clin Endocrinol Metab* 85: 4908 — 4911 Arvat E, Maccario M, Di Vito L, et al. 2001 Endocrine activities of ghrelin, a natural growth hormone secretagogue (GHS), in humans: comparison and interactions with hexarelin, a nonnatural peptidyl GHS, and GH-releasing hormone. *J Clin Endocrinol Metab* 86: 1169 —1174 Cummings E, Purnell JQ, Frayo SR, Schmidova K, Wisse BE, Weigle DS 2001 A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 50: 1714 —1719 Maccario M, Aimaretti G, Corneli G, et al. 2000 Short-term fasting abolishes the sex-related difference in GH and leptin

secretion in humans. *Am J Physiol Endocrinol Metab* 279: E411—E416 Gurd W, Parent G, Eniojukan R, Bowers CY, Tannenbaum GS 2001 Interrelationship between the novel peptide ghrelin and somatostatin/GHRH in regulation of pulsatile GH secretion. Proceedings of the 83rd Annual Meeting of The Endocrine Society, Denver, CO, 2001, p 72 Tannenbaum GS, Bowers CY 2001 Interactions of growth hormone secretagogues and growth hormone-releasing hormone/somatostatin. *Endocrine* 14: 21—27 Gondo RG, Aguiar-Oliveira MH, Hayashida CY, et al. 2001 Growth hormone-releasing peptide-2 stimulates GH secretion in GH-deficient patients with mutated GH-releasing hormone receptor. *J Clin Endocrinol Metab* 86: 3279 —3283 Bowers CY 1998 Synergistic release of growth hormone by GHRP and GHRH: Scope and implication. In: Bercu BB, Walker BF, eds. *Growth hormone secretagogues in clinical practice*. Ed. 1. New York: Marcel Dekker Inc.; 1—27 Broglio F, Arvat E, Benso A, et al., Ghrelin, a natural GH secretagogue produced by the stomach, induces hyperglycemia and reduces insulin secretion in humans. *J Clin Endocrinol Metab*, in press Heiman ML, Tschop M 2001 Ghrelin acts to provide the calories that growth hormone requires for growth and repair. Proceedings of the 83rd Annual Meeting of The Endocrine Society, Denver, CO, 2001, p 33 Shintani M, Ogawa Y, Ebihara K, et al. 2001 Ghrelin, an endogenous growth hormone secretagogue, is a novel orexigenic peptide that antagonizes leptin action through the activation of hypothalamic neuropeptide Y/Y1 receptor pathway. *Diabetes* 50: 227—232 Asakawa A, Inui A, Kaga T, et al. 2001 Ghrelin is an appetite-stimulatory signal from stomach with structural resemblance to motilin. *Gastroenterology* 120: 337—345 Kamegai J, Tamura H, Shimizu T, Ishii S, Sugihara H, Wakabayashi I 2000

Central effect of ghrelin, an endogenous growth hormone secretagogue, on hypothalamic peptide gene expression. *Endocrinology* 141: 4797– 4800

Wren AM, Small CJ, Ward HL, et al. 2000 The novel hypothalamic peptide ghrelin stimulates food intake and growth hormone secretion. *Endocrinology* 141: 4325– 4328

Nakazato M, Murakami N, Date Y, et al. 2001 A role for ghrelin in the central regulation of feeding. *Nature* 409: 194 –198

Tschop M, Wawarta R, Riepl RL, et al. 2001 Post-prandial decrease of circulating human ghrelin levels. *J Endocrinol Invest* 24: RC19-RC21

Tschop M, Weyer C, Tataranni PA, Devanarayan V, Ravussin E, Heiman ML 2001 Circulating ghrelin levels are decreased in human obesity. *Diabetes* 50: 707–709

Ravussin E, Tschop M, Heiman ML, Bouchard C 2001 Plasma ghrelin concentration and energy balance: overfeeding and negative energy balance studies in twins. *J Clin Endocrinol Metab* 86: 4547– 4551

Toshinai K, Mondal MS, Nakazato M, et al. 2001 Upregulation of Ghrelin expression in the stomach upon fasting, insulin-induced hypoglycemia, and leptin administration. *Biochem Biophys Res Commun* 281: 1220 –1225

Toth DW, Hellmann PH, Gordon DA, et al. 2001 Evidence that the orexigenic effect of ghrelin is mediated by a distinct GHS-R subtype. *Proceedings of the 83rd Annual Meeting of The Endocrine Society, Denver, CO, 2001, p 169*

Chen C 2000 Growth hormone secretagogue actions on the pituitary gland: multiple receptors for multiple ligands? *Clin Exp Pharmacol Physiol* 27: 323–329

Torsello A, Locatelli V, Melis MR, et al. 2000 Differential orexigenic effects of hexarelin and its analogs in the rat hypothalamus: indication for multiple

54. 55. 56. 57. 58. 59. 60. 61. 62. 63. 64. 65. 66. 67. 68. 69. 70. 71. 72. 73. 74. 75. 76. 77. 78. 79. 80. 81. 82. growth hormone secretagogue receptor

subtypes. *Neuroendocrinology* 72: 327—332 Heymsfield SB, Greenberg AS, Fujioka K, et al. 1999 Recombinant leptin for weight loss in obese and lean adults: a randomized, controlled, dose-escalation trial. *JAMA* 282: 1568 —75 Brobeck JR 1946 Mechanism of the development of obesity in animals with hypothalamic lesions. *Physiol Rev* 26: 541—559 Powley TL, Opsahl CH, Cox JE, Weingarten HP 1980 The role of the hypothalamus in energy homeostasis. In: Morgane PJ, Panskepp J, eds. *Handbook of the hypothalamus. Part A: Behavioral studies of the hypothalamus*. New York: Marcel Dekker, Inc.; pp 211—298 Bernadis LL, Berlinger LL 1987 The dorsomedial hypothalamic nucleus revisited. *Brain Res* 434: 321—381 Aravich PF, Scalfani A 1983 Paraventricular hypothalamic lesions and medial hypothalamic cuts produce similar hyperphagia syndromes. *Behav Neurosci* 97: 970 —983 Weingarten HP, Chang P, McDonald TJ 1985 Comparisons of the metabolic and behavioral disturbances following paraventricular and ventromedial hypothalamic lesions. *Brain Res Bull* 14: 1551—1559 Tokunaga K, Fukushima M, Kemnitz JW, Bray GA 1986 Comparison of ventromedial and paraventricular lesions in rats that become obese. *Am J Physiol* 251: R1221—R1227 Powley TL, Keesey RE 1970 Relationship of body weight to the lateral hypothalamic feeding syndrome. *J Comp Physiol Psychol* 70: 25—36 van den Pol AN 1982 Lateral hypothalamic damage and body weight regulation: role of gender, diet, and lesion placement. *Am J Physiol* 243: R265— R274 Clark JT, Kalra PS, Crowley WR, Kalra SP 1984 Neuropeptide Y and human pancreatic polypeptide stimulate feeding behavior in rats. *Endocrinology* 115: 427— 429 Stanley BG, Chin AS, Leibovitz SF 1985 Feeding and drinking elicited by central injection of neuropeptide Y: evidence for a hypothalamic

site(s) of action. *Brain Res Bull* 14: 521—524 de Lecea L, Kilduff TS, Peyron C, et al. 1998 The hypocretins: two hypothalamic peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* 95: 322—327 Sakurai T, Amemiya A, Ishii M, et al. 1998 Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 92: 573—585 Bittencourt JC, Presse F, Arias C, et al. 1992 The melanin-concentrating hormone system of the rat brain: an immuno- and hybridization histochemical characterization. *J Comp Neurol* 319: 218 —245 Qu D, Ludwig DS, Gammeltoft S, et al. 1996 A role for melanin-concentrating hormone in the central regulation of feeding behavior. *Nature* 380: 243—247 Lu D, Willard D, Patel IR, et al. 1994 Agouti protein is an antagonist of the melanocyte-stimulating-hormone receptor. *Nature* 371: 799 — 802 Ollmann MM, Wilson BD, Yang YK, et al. 1997 Antagonism of central melanocortin receptors in vitro and in vivo by agouti-related protein. *Science* 278: 135—138 Fan W, Boston BA, Kesterson RA, Hraby VJ, Cone RD 1997 Role of melanocortinergic neurons in feeding and the agouti obesity syndrome. *Nature* 385: 165—168 Hahn TM, Breininger JF, Baskin DG, Schwartz MW 1998 Coexpression of *Agrp* and *NPY* in fasting-activated hypothalamic neurons. *Nature Neurosci* 1: 271—272 Mezey E, Kiss JZ, Mueller GP, Eskay R, Odonohue TL, Palkovits M 1985 Distribution of pro-opiomelanocortin derived peptides, adrenocorticotrope hormone, -melanocyte-stimulating hormone and -endorphin (*ACTH*, *-MSH*, *-END*) in the rat hypothalamus. *Brain Res* 328: 341—347 Halaas JL, Gajiwala KS, Maffei SL, et al. 1995 Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269: 543—546 Pellemounter MA, Cullen MJ, Baker MB, et al. 1995 Effects of the obese gene

product on body weight regulation in ob/ob mice. *Science* 269: 540 —543

Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P 1995 Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science* 269: 546 —549

Leibel RL, Chung WK, Chua Jr SC 1997 The molecular genetics of rodent single gene obesities. *J Biol Chem* 272: 31937—31940

Strobel A, Issad T, Camoin L, Ozata M, Strosberg AD 1998 A leptin missense mutation associated with hypogonadism and morbid obesity. *Nat Genet* 18: 213—215

Montague CT, Farooqi IS, Whitehead JP, et al. 1997 Congenital leptin deficiency is associated with severe early-onset obesity in humans. *Nature* 387: 903—908

Clement K, Vaisse C, Lahlou N, et al. 1998 A mutation in the human leptin receptor gene causes obesity and pituitary dysfunction. *Nature* 392: 398 — 401

Mercer JG, Hoggard N, Williams LM, et al. 1996 Localization of leptin receptor mRNA and the long splice variant (O-Rb) in mouse hypothalamus and adjacent brain regions by in situ hybridization. *FEBS Lett* 387: 113—116

Diano S, Kalra SP, Horvath TL 1998 Leptin receptor immunoreactivity is associated with the Golgi apparatus of hypothalamic cells. *J Neuroendocrinol* 9: 647—650

Elmquist JK, Elias CF, Saper CB 1999 From lesions to leptin: hypothalamic control of food intake and body weight. *Neuron* 2: 221—232

Elmquist JK, Bjorbaek C, Ahima RS, Flier JS, Saper CB 1998 Distributions of leptin receptor mRNA isoforms in the rat brain. *J Comp Neurol* 395: 535—547

Hakansson ML, Brown H, Ghilardi N, Skoda RC, Meister B 1998 Leptin receptor immunoreactivity in chemically defined target neurons of the



hypothalamus. *J Neurosci* 18: 559 —572 Yarnell DO, Knight DS, Hamilton K, Tulp O, Tso P 1998 Localization of leptin receptor immunoreactivity in the lean and obese zucker rat brain. *Brain Res* 785: 80 —90 Schwartz MW, Seeley RJ, Campfield LA, Burn P, Baskin DG 1996 Identification of leptin action in rat hypothalamus. *J Clin Invest* 98: 1101—1106 Broberger C, de Lecea L, Sutcliffe JG, Hokfelt T 1998 Hypocretin/orexin and melanin-concentrating hormone-expressing cells from distinct populations in the rodent lateral hypothalamus: relationship to the neuropeptide Y and agouti gene-related protein systems. *J Comp Neurol* 402: 460 — 474 Elias CF, Saper CB, Maratos-Flier E, et al. 1998 Chemically defined projections linking the mediobasal hypothalamus and the lateral hypothalamus area. *J Comp Neurol* 402: 442— 459 Horvath TL, Kalra SP, Naftolin F, Leranth C 1995 Morphological evidence for a galanin-opiate interaction in the rat mediobasal hypothalamus. *J Neuroendocrinol* 7: 579 —588 Horvath TL, Bechmann I, Kalra SP, Naftolin F, Leranth C 1997 Heterogeneity in the neuropeptide Y-containing neurons of the rat arcuate nucleus: GABAergic and non-GABAergic subpopulations. *Brain Res* 756: 283—286 Horvath TL, Diano S, van den Pol AN 1999 Synaptic interaction between hypocretin (orexin) and neuropeptide Y cells in the rodent and primate hypothalamus: a novel circuit implicated in metabolic and endocrine regulations. *J Neurosci* 19: 1072—1087 Merchenthaler I 1991 Neurons with access to the general circulation in the central nervous system of the rat: a retrograde tracing study with fluoro-gold. *Neuroscience* 44: 655— 662 Guan XM, Yu H, Palyha OC, et al. 1997 Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. *Brain Res Mol Brain Res* 48: 23—29

Tannenbaum GS, Lapointe M, Beaudet A, Howard AD 1998 Expression of growth hormone secretagogue-receptors by growth hormone-releasing hormone neurons in the mediobasal hypothalamus. *Endocrinology* 139: 4420 — 4423

Shuto Y, Shibasaki T, Wada K, et al. 2001 Generation of polyclonal antiserum against the growth hormone secretagogue receptor (GHS-R): evidence that the GHS-R exists in the hypothalamus, pituitary and stomach of rats. *Life Sci* 68: 991—996

Willesen MG, Kristensen P, Romer J 1999 Co-localization of growth hormone secretagogue receptor and NPY mRNA in the arcuate nucleus of the rat. *Neuroendocrinology* 70: 306 —316

Smith RG, Leonard R, Bailey AR, et al. 2001 Growth hormone secretagogue receptor family members and ligands. *Endocrine* 14: 9 —14

Luckman SM, Rosenzweig I, Dickson SL 1999 Activation of arcuate nucleus 100. 101. 102. 103. 104. 105. 106. 107. 108. 109. 110. 111. 112. 113. 114. 115. neurons by systemic administration of leptin and growth hormone-releasing peptide-6 in normal and fasted rats. *Neuroendocrinology* 70: 93—100

Dickson SL, Leng G, Robinson IC 1993 Systemic administration of growth hormone-releasing peptide activates hypothalamic arcuate neurons. *Neuroscience* 52: 303—306

Dickson SL, Luckman SM 1997 Induction of c-fos messenger ribonucleic acid in neuropeptide Y and growth hormone (GH)-releasing factor neurons in the rat arcuate nucleus following systemic injection of the GH secretagogue, GH-releasing peptide-6. *Endocrinology* 138: 771—777

Bailey AR, Von Englehardt N, Leng G, Smith RG, Dickson SL 2000 Growth hormone secretagogue activation of the arcuate nucleus and brainstem occurs via a non-noradrenergic pathway. *J Neuroendocrinol* 12: 191—197

Hewson AK, Dickson SL 2000 Systemic administration of ghrelin induces Fos and Egr-1 proteins in

the hypothalamic arcuate nucleus of fasted and fed rats. *J Neuroendocrinol* 12: 1047—1049 Lambert PD, Anderson KD, Sleeman MW, et al. 2001 Ciliary neurotrophic factor activates leptin-like pathways and reduces body fat, without cachexia or rebound weight gain, even in leptin-resistant obesity. *Proc Natl Acad Sci USA* 98: 4652— 4657 Kalra SP 2001 Circumventing leptin resistance for weight control. *Proc Natl Acad Sci USA* 98: 4279 — 4281 Saladin R, De Vos P, Guerre-Millo M, et al. 1995 Transient increase in obese gene expression after food intake or insulin administration. *Nature* 377: 527 —529 Sahu A, Kalra PS, Kalra SP 1988 Food deprivation and ingestion induced reciprocal changes in neuropeptide Y concentrations in the paraventricular nucleus. *Peptides* 9: 83— 86 Sahu A, White JD, Kalra PS, Kalra SP 1992 Hypothalamic neuropeptide Y gene expression in rats on scheduled feeding regimen. *Mol Brain Res* 15: 15—18 Sahu A 1998 Evidence suggesting that galanin (GAL), melanin-concentrating hormone (MCH), neurotensin (NT), proopiomelanocortin (POMC) and neuropeptide Y (NPY) are targets of leptin signaling in the hypothalamus. *Endocrinology* 139: 795—798 Schwartz MW, Seeley RJ, Woods SC, et al. 1997 Leptin increases hypothalamic pro-opiomelanocortin mRNA expression in the rostral arcuate nucleus. *Diabetes* 46: 2119 —2123 Cowley MA, Smart JL, Rubinstein M, et al. 2001 Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 411: 480 — 484 Cowley MA, Pronchuk N, Fan W, Dinulescu DM, Colmers WF, Cone RD 1999 Integration of NPY, AGRP, and melanocortin signals in the hypothalamic paraventricular nucleus: evidence of a cellular basis for the adipostat. *Neuron* 24: 155—163 van den Pol AN, Gao XB, Obrietan K, Kilduff T, Belousov A 1998 Pre- and postsynaptic actions

and modulation of neuroendocrine neurons by a new hypothalamic peptide, hypocretin/orexin. *J Neurosci* 18: 7962—7971 Broberger C, Johansen J, Johansson C, Schalling M, Hokfelt T 1998 The neuropeptide Y/agouti gene-related protein (AGRP) brain circuitry in normal, anorectic, and monosodium glutamate-treated mice. *Proc Natl Acad Sci USA* 95: 15043—10048 DeFalco J, Tomishima M, Liu H, et al. 2001 Virus-assisted mapping of neural inputs to a feeding center in the hypothalamus. *Science* 291: 2608 —2613