Integrating concepts of physiological and behavioral resistance to parasites

Health & Medicine



Introduction

Behavior establishes the first interface at which animal hosts and parasites interact (Moore, 2002). Given the parallels between physiological and behavioral forms of resistance to parasites, the concept of the "behavioral immune system" has emerged: the cognitive and behavioral mechanisms for avoiding infectious agents and their cues (Schaller and Duncan, 2007; Schaller and Park, 2011). In other words, through behavioral means, hosts can prevent establishment of parasites analogously to physiological resistance strategies. Does this analogy predict that behavioral resistance should follow the evolutionary and ecological patterns of innate and/or acquired forms of physiological resistance? Although behavioral resistance can also encompass strategies for controlling parasite growth, recovery from infection, and tolerance of infection (Moore, 2002; Hart, 2011; Hawley et al., 2011; Adelman and Hawley, 2017; Townsend et al., 2020), in this review, I explore the parallels between behavioral avoidance of infection and physiological resistance, highlighting potential limitations of acquiring behavioral resistance through individual experience.

Distinguishing Physiological and Behavioral Forms of Parasite Resistance

First, I establish definitions of physiological and behavioral resistance within the broader framework of host resistance to parasites. I use the term "parasite" for all infectious agents, including micro-organisms usually referred to as "pathogens." Infection of a host by a parasite occurs through a multistep process that can be broken down into two general steps: (1) contact between host and parasite (host exposure) and (2) establishment of parasite https://assignbuster.com/integrating-concepts-of-physiological-and-behavioral-resistance-to-parasites/

infection in or on the host (host susceptibility) (Combes, 2001; Hall et al., 2017). Exactly how contact is defined will depend upon the transmission mode of the parasite: for a vector-transmitted parasite, it could be the vector biting the host; for a sexually transmitted parasite, it could be deposition of the parasite into the reproductive tract during mating. The host has resistance to the parasite if it can interfere with at least one of these two broad steps along the parasite's path from the previous host.

Correspondingly, this two-step process generates two broad categories of resistance: (1) pre-contact resistance, sometimes called " avoidance," which prevents or reduces likelihood of the initial host-parasite contact, and (2) post-contact resistance, which, given contact between host and parasite, interferes with the establishment of parasite infection.

Behaviorals and physiologicals resistance roughly map onto, but do not exactly match, pre- and post-contact resistance. Indeed, behaviors are one form of pre-contact resistance, commonly conceptualized as an avoidance response of animals to a cue of parasite risk (Curtis, 2014). However, pre-contact resistance is not achieved only through behavioral responses: e. g., for pollinator-transmitted parasites of plants, earlier flowering can function as pre-contact resistance (Biere and Antonovics, 1996). Similarly, post-contact resistance does not have to be physiological: e. g., many ectotherms will raise their body temperatures by seeking warm environments to fight parasite infections with a " behavioral fever" (Kluger, 1979). Other behavioral forms of post-contact resistance include grooming (Mooring et al., 2004; Akinyi et al., 2013) and self-medication (Clayton and Wolfe, 1993). Although I do not discuss them thoroughly here, these forms of post-https://assignbuster.com/integrating-concepts-of-physiological-and-behavioral-resistance-to-parasites/

contact behavioral resistance might be expected to have many similarities to post-contact physiological resistance. For the sake of simplicity and drawing connections to the previous literature, I will limit the scope of "behavioral resistance" in this review to pre-contact behavioral resistance, and "physiological resistance" to post-contact physiological resistance.

To compare these two forms of resistance, I also distinguish between innate and acquired physiological resistance, a distinction mostly relevant to vertebrates. Innate resistance refers to the mechanisms that detect and defend against parasites that are present throughout the lifetime of an individual (Beutler, 2004). Acquired resistance refers to the ability to gain resistance in subsequent exposures to a parasite following recovery from an initial exposure, sometimes termed " immunological memory" (Bonilla and Oettgen, 2010). Here, I investigate whether delineating innate and acquired resistance is a useful paradigm to apply to behavior.

Evolution of Innate Behavioral Resistance

Corresponding to innate forms of physiological resistance, behaviors for parasite resistance may be innate and shaped heavily by evolutionary forces. Evidence suggests that parasite avoidance behavior is under the control of genes and therefore likely to evolve. In a knockout experiment in laboratory mice (*Mus musculus*), the oxytocin gene was identified as a central component of the olfactory mechanism female mice used to avoid parasitized males (<u>Kavaliers et al., 2005a</u>). Domestic sheep (*Ovis aries* , Perendale lines) artificially selected for physiological resistance to parasites also avoided parasites in their grazing behavior (<u>Hutchings et al., 2007</u>). In

addition, parasite avoidance behavior has been experimentally evolved in *Caenorhabditis elegans* in response to a bacterial parasite, *Serratia marcescens* (Penley and Morran, 2018).

Despite evidence of genetic variation in parasite avoidance behaviors, little is known about the forces that drive their evolution (i. e., allele frequency change). A rich literature has considered theoretical aspects of ecological feedbacks on the evolution of physiological resistance (e. g., Antonovics and Thrall, 1994; Boots and Haraguchi, 1999; Boots et al., 2009). Capturing dynamics of the evolution of physiological resistance involves considering the nature and magnitude of the costs of resistance, and how evolutionary processes are affected by the numbers of hosts and pathogens, in addition to changes in allele frequencies (Antonovics and Thrall, 1994; Boots and Haraguchi, 1999). Identically to models of physiological resistance, theory predicts that if behavioral resistance to a directly transmitted parasite is assumed to carry a fixed cost, resistance is more likely to evolve when costs are low, and resistant and susceptible genotypes can exist in a stable polymorphism under certain conditions (Amoroso and Antonovics, 2020).

However, because behavioral resistance commonly acts before contact between the host and the parasite, the costs of resistance may depend on the transmission mode of the parasite, which defines a contact between the host and the parasite. For example, if social interactions are assumed to be beneficial but also carry a risk of parasite transmission, resistant hosts pay a cost of losing these social interactions, and behavioral resistance (avoidance of conspecifics) is much less likely to evolve (<u>Amoroso and Antonovics, 2020</u>

). These conclusions depend on the assumption that the parasite is contact-transmitted, because the costs and benefits of resistance are a function of the host's social behavior. In a social host, resistance could even function as groups avoiding other groups, with costs allocated or shared among group members. Under different transmission modes, the costs could take different forms: e. g., hypothetically, locomotion costs of avoiding habitats with high risk of environmentally transmitted parasites, or reproductive costs of missed mating opportunities when avoiding sexually transmitted parasites. To better understand how behavioral resistance to parasites could evolve, future work will need to conceptualize and measure the associated costs, which may be unique to different parasite transmission modes.

The genetics of coevolution between hosts and parasites have been intensively studied in terms of physiological resistance, generating predictions for coevolutionary dynamics of behavioral resistance. For example, the gene-for-gene model of resistance was originally developed to explain cases in which a plant host's resistance to infection and a parasite's ability to cause disease were each determined by a single genetic locus (

Flor, 1956; Frank, 1993). Similar to gene-for-gene dynamics, the host could detect and behaviorally avoid a specific elicitor produced by the parasite, and parasites could evade detection by losing that elicitor. A gene-for-gene model could possibly govern coevolution between *C. elegans*, in which the gene controlling lawn-leaving (avoidance) behavior has been identified (

Peng et al., 2018), and its bacterial parasite *Bacillus thuringiensis*, in which the gene has been identified that governs production of a toxin that *C. elegans* detects (Nakad et al., 2016). However, the coevolutionary

dynamics of these traits have not yet been examined. In addition, theoretical assessments of coevolutionary processes in mate choice have suggested that avoidance behaviors can influence virulence evolution in sexually transmitted parasites (<u>Ashby and Boots, 2015</u>; <u>Ashby, 2020</u>), and similar predictions could follow for the coevolutionary dynamics of other host avoidance behaviors and parasite transmission modes.

Although hosts could respond to specific cues produced by the parasite, they might instead respond to general cues: a behavioral equivalent to the concept of broad spectrum resistance in plants (e.g., Ke et al., 2017), or cross-immunity of a host to multiple strains of a parasite (Haraguchi and Sasaki, 1997). Parasites are likely to be impossible for hosts to detect directly (Hart, 2011), and therefore selection may be based on detectable yet indirect cues: for example, avoiding feces (Sarabian and MacIntosh, 2015; Amoroso et al., 2017), detecting general signs of disease in conspecifics (Curtis et al., 2004; Paciência et al., 2019), or swatting away insect vectors (Hart and Hart, 1994). If evolved resistance behaviors are general, they could confer resistance to multiple parasites transmitted through the same route, similar to the broader protections offered by the innate immune system. In this way, coevolutionary dynamics between a host and one of its parasites could have ramifications for other parasites transmitted via the same route, possibly resulting in diffuse coevolution (Iwao and Rausher, 1997).

Acquired Behavioral Resistance

In addition to innate forms of resistance, behaviors for resisting parasites might be acquired in the course of an individual's lifetime, analogous to immunological memory. Evidence supports that animals can learn to avoid predators (reviewed in Griffin et al., 2000), raising the possibility that prior experience of infection with a parasite could induce a novel aversion to the conditions of exposure to that parasite. Classical conditioning research suggests that animals can reliably associate two things when they are separated by a very short time delay, on the order of seconds (Perin, 1943; Renner, 1964). Meanwhile, for most parasites, the incubation period—the time lag between a host being exposed to a parasite and the onset of clinical signs of infection—is on the order of days (Lessler et al., 2009; Azman et al., 2013; Lee et al., 2013; Rudolph et al., 2014). On the short end of this spectrum are parasites with incubation periods of 12 h (e. g., influenza B) (Lessler et al., 2009), and as short as 4 h for some foodborne illnesses (Eley, 1992). Thus, the onset of experienced signs of infection typically occurs too late relative to exposure for a host to associate the two events via classical forms of conditioning, making it exceedingly difficult for hosts to learn to avoid parasite exposure.

However, at least two exceptions exist to the expectation that parasite avoidance is unlikely to be acquired. First, association between exposure to parasites and infection signs would be possible when the host can perceive the parasite's attacks simultaneously with exposure, similar to conditions for learning predator avoidance, such as large, conspicuous ectoparasites like biting flies, whose bites are painful. A series of experiments have shown that

both laboratory mice and deer mice (Peromyscus maniculatus), upon naïve exposure to biting flies, will avoid the flies by burrowing into the bedding of their cages (Kavaliers et al., 2001, 2003, 2005b). On subsequent exposures, the mice will burrow even if the flies' biting mouthparts have been removed: evidence of a learned avoidance response (Kavaliers et al., 2001). Mice can also learn through observation to avoid the flies with mouthparts removed, without any prior experience of being bitten (Kavaliers et al., 2001, 2003, 2005b). If such avoidance can be learned, it could also have downstream effects on the vector-borne parasites that these conspicuous ectoparasites transmit. It is important to note that in addition to being detectable, a parasite must also induce experienced negative effects (e. g., pain, other clinical signs) in temporal proximity to the infestation (e. g., fly bite). For example, although rock pigeons (Columba livia) could detect lice in their feathers, prior experience did not improve rate or efficacy of preening (Villa et al., 2016). The authors suggest that the relatively low virulence and lack of immediate negative fitness effects of lice could explain the absence of a "priming" effect (Villa et al., 2016).

A second exception is a phenomenon called "conditioned taste aversion," the association of taste and odor with gastrointestinal distress such as nausea or vomiting, which can take place over a delay of hours, longer than observed in traditional operant conditioning. In an experiment, an hour or more after ingesting flavored water, mice were injected with a toxin that induced gastric disturbances. On subsequent trials, the mice reliably avoided the flavored water, even after just one experience (Garcia et al., 1966). This phenomenon has since been confirmed in a wide variety of animals, over

delays commonly ranging from 1 to 7 h, and up to 24 h (Lin et al., 2017)— long enough to encompass some shorter incubation periods of parasites. Conditioned taste aversion has been extended to associations between specific flavors or odors and digestive discomfort induced specifically by parasite infection. For example, *Drosophila melanogaster* avoided smells present when they ingested a virulent bacterial parasite (Babin et al., 2014), and *Rattus norvegicus domestica* became averse to ingesting flavors following injection with third-stage nematode larvae (Keymer et al., 1983). Taste-conditioned avoidance of parasites has also been demonstrated in *C. elegans* with lawn-leaving assays (Zhang et al., 2005), and has been suggested to be heritable for multiple generations (Moore et al., 2019). Whether learned avoidance behaviors can be transmitted across generations in other host-parasite systems analogously to maternal transmission of antibodies (Grindstaff et al., 2003) deserves further research.

In humans, the improbability of individual learning as an effective strategy for parasite avoidance is underscored by the misattributions of the cause of infectious diseases prior to wide acceptance of germ theory. For example, the widespread belief in the miasma theory of disease—that diseases are acquired through inhaling "bad air"—as recently as the late 19th Century (Halliday, 2001) suggests that for most of our evolutionary history, humans could not reliably associate the particulars of our exposures to infectious agents with symptoms that ensued after a delay. However, human social learning and communication have since proven effective at coordinating widespread avoidance behaviors, including strategies such as hygiene and social distancing (Curtis, 2014; Townsend et al., 2020).

The literature reviewed here suggests that most host behavioral resistance strategies are likely to be innate, rather than acquired through past experience. Furthermore, given the constraints of classical learning processes and typical incubation periods of parasites, not only could acquiring behavioral avoidance of parasites be ineffective, but reliance on learning may lead to incorrect associations between harmless stimuli and experienced signs of infection. Some specialized learning processes, such as imprinting, have even been suggested to cause maladaptive attraction to conspecifics displaying signs of infection with directly transmitted parasites (Stephenson and Reynolds, 2016). Although aversion to parasites is only likely to be acquired in specific circumstances, prior experience could still influence the contexts in which individuals perform behavioral resistance strategies, e. g., if animals associate an innately aversive cue of parasite risk with a specific individual or habitat. Thus, even if behavioral resistance is unlikely to be acquired analogously to immunological memory, future research should consider what role individual experience could play in shaping behavioral resistance.

Discussion

The examples discussed here illustrate that extending the analogy of the "behavioral immune system" can be a useful framework to generate predictions about behavioral resistance to parasites, but should be applied cautiously and with attention to the biology of the processes involved (Table 1). A rich literature on physiological resistance to parasites has investigated the population genetics and numerical dynamics that lead to the evolution of different forms of resistance (Boots and Bowers, 2004; Boots et al., 2009).

Given that little is known about the specific processes that generate changes in frequency of alleles for behavioral resistance to parasites, expectations from theoretical and empirical research on physiological resistance are a useful starting point from which to advance. This review suggests that evolutionary dynamics of behavioral resistance may be predicted reasonably well by physiological resistance theory under certain assumptions. But more empirical research on the evolutionary dynamics of behavioral resistance is needed, especially on the costs, and the possibility that they may vary with parasite transmission mode. On the other hand, only in particular circumstances it is likely that behavioral resistance to a parasite would be acquired newly in the lifetime of an individual, a major departure from processes of acquired physiological resistance. If behavioral resistance is primarily an innate, evolved response to parasites, understanding the evolutionary—and coevolutionary—processes that have generated behavioral resistance, and whether general principles govern these processes should be central aims in future studies of host behavioral responses to parasites.

TABLE 1

Several key contrasts between physiological and behavioral resistance.

I have considered behavioral and physiological resistance separately here, but behavioral resistance precedes physiological resistance in the course of a host's interaction with a single parasite; thus, these two forms of resistance are not independent. Physiological and behavioral resistance are

expected to interact with one another. For example, they made trade off, with individuals balancing investment in immune and behavioral defenses (

Zylberberg et al., 2013; Zylberberg, 2014). Together, behavioral and physiological resistance constitute a step-wise infection process, in which multiple sequential steps are required (Hall et al., 2017). Not only is a two-step process likely to lead to unique infection genetics, but genes for functionally independent resistance traits can become linked when they are jointly affected by coevolutionary processes (Fenton et al., 2009, 2012). Behavioral and physiological resistance evolution might be expected to interact in many ways, an exciting open direction for future investigation.

Author Contributions

The author confirms being the sole contributor of this work and has approved it for publication.

Funding

This study was supported by NIH (Grant No. R01GM122061), part of the joint NSF-NIH-USDA Ecology and Evolution of Infectious Diseases Program.

Conflict of Interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Acknowledgments

I would like to thank Janis Antonovics for providing feedback on the manuscript and for many helpful discussions on this topic.

References

Adelman, J. S., and Hawley, D. M. (2017). Tolerance of infection: A role for animal behavior, potential immune mechanisms, and consequences for parasite transmission. *Horm. Behav.* 88, 79–86. doi: 10. 1016/j. yhbeh. 2016. 10. 013

Akinyi, M. Y., Tung, J., Jeneby, M., Patel, N. B., Altmann, J., and Alberts, S. C. (2013). Role of grooming in reducing tick load in wild baboons (*Papio cynocephalus*). *Anim. Behav.* 85, 559–568. doi: 10. 1016/j. anbehav. 2012. 12. 012

Amoroso, C. R., and Antonovics, J. (2020). Evolution of behavioural resistance in host – pathogen systems. *Biol. Lett.* 16: 20200508.

Amoroso, C. R., Frink, A. G., and Nunn, C. L. (2017). Water choice as a counterstrategy to faecally transmitted disease: an experimental study in captive lemurs. *Behaviour* 154, 1239–1258. doi: 10. 1163/1568539X-00003466

Antonovics, J., and Thrall, P. H. (1994). The cost of resistance and the maintenance of genetic polymorphism in host-pathogen systems. *Proc. R. Soc. B Biol. Sci.* 257, 105–110.

Ashby, B. (2020). Antagonistic coevolution between hosts and sexually transmitted infections. *Evolution* 74, 43–56. doi: 10. 1111/evo. 13883

Ashby, B., and Boots, M. (2015). Coevolution of parasite virulence and host mating strategies. *Proc. Natl. Acad. Sci.* 112, 13290–13295. doi: 10. 1073/pnas. 1508397112

Azman, A. S., Rudolph, K. E., Cummings, D. A. T., Lessler, J., and Tor, E. (2013). The incubation period of cholera: A systematic review supplement. *J. Infect.* 66, 432–438. doi: 10. 1016/j. jinf. 2012. 11. 013. The

Babin, A., Kolly, S., Schneider, F., Dolivo, V., Zini, M., and Kawecki, T. J. (2014). Fruit flies learn to avoid odours associated with virulent infection. *Biol. Lett.* 10: 20140048.

Beutler, B. (2004). Innate immunity: An overview. *Mol. Immunol.* 40, 845-859. doi: 10. 1016/j. molimm. 2003. 10. 005

Biere, A., and Antonovics, J. (1996). Sex-specific costs of resistance to the fungal pathogen Ustilago violacea (Microbotryum violaceum) in Silene alba. *Evolution* 50, 1098–1110. doi: 10. 1111/j. 1558-5646. 1996. tb02350. x

Bonilla, F. A., and Oettgen, H. C. (2010). Adaptive immunity. *J. Allergy Clin. Immunol.* 125, S33–S40. doi: 10. 1016/j. jaci. 2009. 09. 017

Boots, M., Best, A., Miller, M. R., and White, A. (2009). The role of ecological feedbacks in the evolution of host defence: What does theory tell us? *Philos. Trans. R. Soc. B Biol. Sci.* 364, 27–36. doi: 10. 1098/rstb. 2008. 0160

Boots, M., and Bowers, R. G. (2004). The evolution of resistance through costly acquired immunity. *Proc. R. Soc. B Biol. Sci.* 271, 715–723. doi: 10. 1098/rspb. 2003. 2655

Boots, M., and Haraguchi, Y. (1999). The evolution of costly resistance in host-parasite systems. *Am. Nat.* 153, 359–370. doi: 10. 1086/303181

Clayton, D. H., and Wolfe, D. (1993). The adaptive significance of self-medication. *Trends Ecol. Evol.* 8, 60-63.

Combes, C. (2001). *Parasitism: The Ecology and Evolution of Intimate Interactions*. Chicago: University of Chicago Press.

Curtis, V. A. (2014). Infection-avoidance behaviour in humans and other animals. *Trends Immunol.* 35, 457–464. doi: 10. 1016/j. it. 2014. 08. 006

Curtis, V., Aunger, R., and Rabie, T. (2004). Evidence that disgust evolved to protect from risk of disease. *Proc. R. Soc. B Biol. Sci.* 271, S131–S133. doi: 10. 1098/rsbl. 2003. 0144

Eley, A. R. (1992). "Infective bacterial food poisoning," in *Microbial Food Poisoning*, ed. A. R. Eley (Boston, MA: Springer), 15–35.

Fenton, A., Antonovics, J., and Brockhurst, M. A. (2009). Inverse-gene-for-gene infection genetics and coevolutionary dynamics. *Am. Nat.* 174: 645087. doi: 10. 1086/645087

Fenton, A., Antonovics, J., and Brockhurst, M. A. (2012). Two-step infection processes can lead to coevolution between functionally independent infection and resistance pathways. *Evolution* 66, 2030–2041. doi: 10. 1111/j. 1558-5646. 2012. 01578. x

Flor, H. H. (1956). The complementary genic systems in flax and flax rust. *Adv. Genet.* 8, 29–54.

Frank, S. A. (1993). Coevolutionary genetics of plants and pathogens. *Evol. Ecol.* 7, 45–75. doi: 10. 1007/BF01237734

Garcia, J., Ervin, F. R., and Koelling, R. A. (1966). Learning with prolonged delay of reinforcement. *Psychon. Sci.* 5, 121–122. doi: 10. 3758/BF03328311

Griffin, A. S., Blumstein, D. T., and Evans, C. S. (2000). Training captive-bred or translocated animals to avoid predators. *Conserv. Biol.* 14, 1317–1326. doi: 10. 1046/j. 1523-1739. 2000. 99326. x

Grindstaff, J. L., Brodie, E. D., and Ketterson, E. D. (2003). Immune function across generations: Integrating mechanism and evolutionary process in maternal antibody transmission. *Proc. R. Soc. B Biol. Sci.* 270, 2309–2319. doi: 10. 1098/rspb. 2003. 2485

Hall, M. D., Bento, G., and Ebert, D. (2017). The Evolutionary Consequences of Stepwise Infection Processes. *Trends Ecol. Evol.* 32, 612–623. doi: 10. 1016/j. tree. 2017. 05. 009

Halliday, S. (2001). Death and miasma in Victorian London: An obstinate belief. *Br. Med. J.* 323, 1469–1471. doi: 10. 1136/bmj. 323. 7327. 1469

Haraguchi, Y., and Sasaki, A. (1997). Evolutionary pattern of intra-host pathogen antigenic drift: effect of corss-reactivity in immune response. *Philos. Trans. R. Soc. B Biol. Sci.* 352, 11–20.

Hart, B., and Hart, L. (1994). Fly switching by Asian elephants: tool use to control parasites. *Anim. Behav.* 48, 35–45. doi: 10. 1006/anbe. 1994. 1209

Hart, B. L. (2011). Behavioural defences in animals against pathogens and parasites: parallels with the pillars of medicine in humans. *Philos. Trans. R. Soc. B Biol. Sci.* 366, 3406–3417. doi: 10. 1098/rstb. 2011. 0092

Hawley, D. M., Etienne, R. S., Ezenwa, V. O., and Jolles, A. E. (2011). Does animal behavior underlie covariation between hosts' exposure to infectious agents and susceptibility to infection? Implications for disease dynamics. *Integr. Comp. Biol.* 51, 528–539. doi: 10. 1093/icb/icr062

Hutchings, M. R., Knowler, K. J., McAnulty, R., and McEwan, J. C. (2007). Genetically resistant sheep avoid parasites to a greater extent than do susceptible sheep. *Proc. R. Soc. B Biol. Sci.* 274, 1839–1844. doi: 10. 1098/rspb. 2007. 0398

Iwao, K., and Rausher, M. D. (1997). Evolution of plant resistance to multiple herbivores: Quantifying diffuse coevolution. *Am. Nat.* 149, 316–335. doi: 10. 1086/285992

Kavaliers, M., Choleris, E., Ågmo, A., Muglia, L. J., Ogawa, S., and Pfaff, D. W. (2005a). Involvement of the oxytocin gene in the recognition and avoidance of parasitized males by female mice. *Anim. Behav.* 70, 693–702. doi: 10. 1016/j. anbehav. 2004. 12. 016

Kavaliers, M., Choleris, E., and Colwell, D. D. (2001). Learning from others to cope with biting flies: social learning of fear-induced conditioned analgesia and active avoidacne. *Behav. Neurosci.* 115, 661–674.

Kavaliers, M., Colwell, D. D., and Choleris, E. (2003). Learning to fear and cope with a natural stressor: Individually and socially acquired corticosterone and avoidance responses to biting flies. *Horm. Behav.* 43, 99–107. doi: 10. 1016/S0018-506X(02)00021-1

Kavaliers, M., Colwell, D. D., and Choleris, E. (2005b). Kinship, familiarity and social status modulate social learning about "micropredators" (biting flies) in deer mice. *Behav. Ecol. Sociobiol.* 58, 60–71. doi: 10. 1007/s00265-004-0896-0

Ke, Y., Deng, H., and Wang, S. (2017). Advances in understanding broadspectrum resistance to pathogens in rice. *Plant J.* 90, 738–748. doi: 10. 1111/tpj. 13438

Keymer, A., Crompton, D. W. T., and Sahakian, B. J. (1983). Parasite-induced learned taste aversion involving Nippostrongylus in rats. *Parasitology* 86, 455–460. doi: 10. 1017/S0031182000050642

Kluger, M. J. (1979). Fever in ectotherms: Evolutionary implications. *Integr. Comp. Biol.* 19, 295–304. doi: 10. 1093/icb/19. 1. 295

Lee, R. M., Lessler, J., Lee, R. A., Rudolph, K. E., Reich, N. G., Perl, T. M., et al. (2013). Incubation periods of viral gastroenteritis: A systematic review. *BMC Infect. Dis.* 13: 446. doi: 10. 1186/1471-2334-13-446

Lessler, J., Reich, N. G., Brookmeyer, R., Perl, T. M., Nelson, K. E., and Cummings, D. A. T. (2009). Incubation period of acute respiratory viral infections: a systematic review. *Lancet Infect. Dis.* 9, 291–300.

Lin, J. Y., Arthurs, J., and Reilly, S. (2017). Conditioned taste aversions: From poisons to pain to drugs of abuse. *Psychon. Bull. Rev.* 24, 335–351. doi: 10. 3758/s13423-016-1092-8

Moore, J. (2002). *Parasites and the behavior of animals.* New York: Oxford University Press.

Moore, R. S., Kaletsky, R., and Murphy, C. T. (2019). Piwi/PRG-1 Argonaute and TGF-β Mediate Transgenerational Learned Pathogenic Avoidance. *Cell* 177, 1827. e-1841. e. doi: 10. 1016/j. cell. 2019. 05. 024

Mooring, M. S., Blumstein, D. T., and Stoner, C. J. (2004). The evolution of parasite-defence grooming in ungulates. *Biol. J. Linn. Soc.* 81, 17–37. doi: 10. 1111/j. 1095-8312. 2004. 00273. x

Nakad, R., Snoek, L. B., Yang, W., Ellendt, S., Schneider, F., Mohr, T. G., et al. (2016). Contrasting invertebrate immune defense behaviors caused by a single gene, the Caenorhabditis elegans neuropeptide receptor gene npr-1.

BMC Genomics 17: 1–20. doi: 10. 1186/s12864-016-2603-8

Paciência, F. M. D., Rushmore, J., Chuma, I. S., Lipende, I. F., Caillaud, D., Knauf, S., et al. (2019). Mating avoidance in female olive baboons (Papio anubis) infected by Treponema pallidum. *Sci. Adv.* 5, 1-8. doi: 10. 1126/sciadv. aaw9724

Peng, D., Luo, X., Zhang, N., Guo, S., Zheng, J., Chen, L., et al. (2018). Small RNA-mediated Cry toxin silencing allows Bacillus thuringiensis to evade Caenorhabditis elegans avoidance behavioral defenses. *Nucleic Acids Res.* 46, 159–173. doi: 10. 1093/nar/gkx959

Penley, M. J., and Morran, L. T. (2018). Host mating system and coevolutionary dynamics shape the evolution of parasite avoidance in Caenorhabditis elegans host populations. *Parasitology* 145, 724–730.

Perin, C. T. (1943). A quantitative investigation of the delay-of-reinforcement gradient. *J. Exp. Psychol.* 32, 37–51.

Renner, K. E. (1964). Delay of reinforcement: a historical review. *Psychol. Bull.* 61, 341–361.

Rudolph, K. E., Lessler, J., Moloney, R. M., Kmush, B., and Cummings, D. A. T. (2014). Review article: Incubation periods of mosquito-borne viral infections: a systematic review. *Am. J. Trop. Med. Hyg.* 90, 882–891. doi: 10. 4269/ajtmh. 13-0403

Sarabian, C., and MacIntosh, A. J. J. (2015). Hygienic tendencies correlate with low geohelminth infection in free-ranging macaques. *Biol. Lett.* 11: 20150757.

Schaller, M., and Duncan, L. A. (2007). "The behavioral immune system: Its evolution and social psychological implications," in *Evolution and the social mind: Evolutionary psychology and social cognition*, eds J. P. Forgas, M. G. Haselton, and W. von Hippel (New York, NY: Psychology Press), 293–307.

Schaller, M., and Park, J. H. (2011). The Behavioral Immune System (and Why It Matters). *Curr. Dir. Psychol. Sci.* 20, 99–103. doi: 10.

Stephenson, J. F., and Reynolds, M. (2016). Imprinting can cause a maladaptive preference for infectious conspecifics. *Biol. Lett.* 12, 9–12. doi: 10. 1098/rsbl. 2016. 0020

Townsend, A. K., Hawley, D. M., Stephenson, J. F., and Williams, K. E. G. (2020). Emerging infectious disease and the challenges of social distancing in human and non-human animals. *Proc. Biol. Sci.* 287: 20201039. doi: 10. 1098/rspb. 2020. 1039

Villa, S. M., Campbell, H. E., Bush, S. E., and Clayton, D. H. (2016). Does antiparasite behavior improve with experience? An experimental test of the priming hypothesis. *Behav. Ecol.* 27, 1167–1171. doi: 10. 1093/beheco/arw032

Zhang, Y., Lu, H., and Bargmann, C. I. (2005). Pathogenic bacteria induce aversive olfactory learning in Caenorhabditis elegans. *Nature* 438, 179–184. doi: 10. 1038/nature04216

Zylberberg, M. (2014). Galapagos ground finches balance investment in behavioural and immunological pathogen defences. *IBIS* 156, 615–626. doi: 10. 1111/ibi. 12165

Zylberberg, M., Klasing, K. C., and Hahn, T. P. (2013). House finches (Carpodacus mexicanus) balance investment in behavioural and

immunological defences against pathogens. Biol. Lett. 9: 856. doi: 10.

1098/rsbl. 2012. 0856