

Sex manipulation technologies progress in livestock: a review

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Introduction

By 2050, the world's population is expected to increase by 9.6 billion, and the demand for livestock products, by 70 percent ([1](#)). Most of the population growth is however expected to occur in developing countries, where currently most croplands in production face challenges in coping with urbanization, biofuel production, and climate change ([2](#)). Therefore, to meet the forecasted global food demand, utilizing modern biotechnologies to promote sustainable production of livestock will be essential. Sex manipulation technologies are biotechnologies that can selectively predetermine the sex of animal offspring by intervening the normal reproductive process, and besides playing an important role in solving food shortage problem, it can address issues related to animal welfare such as castration. In this review, our objective was to provide suggestions for future research to improve sex manipulation technologies in livestock.

The Origin and Development of Sex Manipulation

In ancient times, boys were also preferred to girls since besides being more likely to appease the wrath of the gods, they were regarded to be a strong support for the elderly ([3](#)). The preference for producing male offspring therefore promoted sex manipulation research ([4](#)).

Theories of what determined the sex of an individual transitioned from myths to science in the 17th century. In 1651, William Harvey compared the reproductive organs of several animals and stated that “ all animals whatsoever, even viviparous creatures, man himself, are all engendered from an egg” ([5](#)). Later on, in the 1670's, AntonieVanLeeuwenhoek

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discovered a microscope which he used to observe sperm cells in human and dog seminal fluid. This discovery became a turning point in the research of sex manipulation ([6](#)).

In 1905 while studying the mealworm beetle *Tenebrio molitor* , Nettie Maria Stevens found that unlike in females, one chromosome was smaller than others in males. She concluded that the shorter chromosome was the “ Y” chromosome, and was responsible for sex determination alongside the larger chromosome, which she named chromosome “ X” ([7](#)).

By the end of the 1950's, the Y chromosome's role in determining the male sex was undisputed. Thus, over the next few decades, efforts were geared toward identifying a single testis-determining factor (*TDF*) in humans. In 1990, researchers found a 35-kb region from Y chromosome in four masculinized XX patients. Southern blotting revealed that the Y-fragment was the likely carrier of the TDF gene ([8](#)). Sequencing showed that an open reading frame (ORF) encoded a single-exon gene named the “ sex-determining” region of the “ Y chromosome” (*SRY* in humans) ([9](#)). Further, researchers found that *SRY* up-regulated the expression of *SOX9* in somatic (pre-Sertoli) cells of the genital ridge ([10](#)). Besides the relationship with autosomal sex reversal and Campomelic Dysplasia (CD) in human ([11](#)), *SOX9* can activate the expression of genes required for Sertoli cell differentiation [e. g., *FGF9* ([12](#))], causing the re-expression of genes associated with ovarian development [e. g., *WNT4* ([13](#))]. Importantly, *SOX9* also promotes its own expression, thereby bypassing the testicular differentiation that requires sustained *SRY* expression ([14](#)). Therefore, the

discovery of *SRY* on the Y chromosome was a major breakthrough in the mammalian sex determination theory and promoted the development of sex manipulation technologies by providing a theoretical basis. Thereafter, the research on sex manipulation technologies gained traction with various studies seeking to enhance the respective existing technologies. Herein, we discussed the advances in sex manipulation technologies in animals with a focus of providing an informative insight for future development.

Sex Selection Strategies During Spermatogenesis

The independent orientation of homologous chromosome pairs along the metaphase plate during metaphase I of meiosis and the subsequent separation of homologs and sister chromatids during anaphase I of meiosis allow an independent distribution of X or Y chromosomes to each secondary spermatocyte ([15](#)). During meiosis, some genes encode proteins with essential roles in structures or functions specific to spermatogenic cells, are expressed in developmentally regulated patterns, and are transcribed only in, or produce mRNAs unique to, spermatogenic cells ([16](#)). Therefore, the expression of these genes can regulate the ultimate meiotic stage-specific protein expression and change the ratio of X- and Y-bearing sperm during spermatogenesis.

In recent years, with the development of molecular biology and cell biology, there were more and more sex-determining genes being identified, including *Sry* ([17](#), [18](#)), *Amh* ([19](#)), *Sox* family ([10](#), [20](#), [21](#)), *Dmrt1* ([22](#)), *Fgf9* ([23](#)), *Gata4* ([24](#)), etc., which are associated with male sex determination, and *Wnt4* ([25](#) - [27](#)), *Dax1* ([28](#)), *Rspo1* ([29](#), [30](#)), *Foxl2* ([27](#), [31](#) - [33](#))

associated with female sex determination. Presently, gene edit and RNA interference targeting these sex-determining genes are the main methods often used to bias the ratio of X- and Y-bearing sperm during spermatogenesis.

The emergence of gene editors such as zinc finger nucleases (ZFN), transcription-activator-like endonucleases (TALENs), and the clustered regularly interspaced short palindromic repeats/Cas endonucleases (CRISPR/Cas) system has offered great potential in the selection of sex at the gene level ([34](#)).

Successful ZFN-mediated disruption of a sex-determining gene in rainbow trout ([35](#)) and the generation of *Sry* knockout mouse via oocyte injection using TALEN ([36](#)) have since been reported. However, standard molecular biology laboratories cannot afford the demand of ZFN molecules because most ZFN molecules are commercial. Although TALENs are more effective than ZFNs, the design of TALEN is complicated and time-consuming due to the high amounts of plasmids involved. Moreover, the specificity of TALEN to target the expected DNA sequences remains unconfirmed. The CRISPR/Cas system is superior to ZFNs and TALENs due to its ease of use and cost-effectiveness, provided a molecular biology laboratory has access to CRISPR kits. Besides, the CRISPR/Cas system can identify and induce mutation on target loci with high specificity ([37](#)). Although these gene editors cannot be used in livestock production presently, the CRISPR/Cas system is the most suitable technique for sex selection among them.

The CRISPR/Cas system is usually introduced into zygotes via microinjection ([38](#), [39](#)) or combined with somatic cell nuclear transfer or surgical transfer to generate offspring ([40](#)). Recently, researchers have biased the sex ratio of populations of different organisms such as plants, insects, crustacean, and fish, at the gene level ([41](#)). The CRISPR/Cas system has been used to control agricultural pest through female-elimination approaches that give rise to more males in an insect population. Such a rise can either result in a population decrease or eliminate the population occasioned by the lack of females ([42](#)).

A recent study in mice described a new strategy to bias the sex ratio of the offspring. Three CRISPRguide RNAs targeting three autosomal genes which are essential for early development in mouse were encoded on murine Y paternal line, while the CRISPR-Cas9 enzyme was encoded into the maternal line. After fertilization, both the Y-encoded guide RNAs from the paternal sperm and the Cas9 protein from the maternal egg targets resulted in self-destructing males ([41](#)). The high homology of the Sp1-binding site between humans and rabbits makes the latter a suitable animal model for studying the clinical male-to-female sex reversal syndrome in humans. The first rabbit hermaphroditism model was recently generated using the CRISPR/Cas9 system. Ten potential off-target sites were analyzed by Sanger sequencing and the T7E1 assay to determine whether off-target effects occurred in the *SRY* mosaic rabbit. To determine whether chimeric mutations in the *SRY* gene induced hermaphroditism, the internal genitalia of *SRY*-mutant chimeric rabbits were also analyzed. The generation of the model provided a platform to understand the pathogenesis of hermaphroditism and identify

novel therapies for human clinical hermaphroditism with male-to-female sex reversal syndrome ([43](#)).

Gene editing technology requires highly skilled personnel, and the lack of a clear role of the sex-determining gene in livestock sex determination hinders its application in this field ([34](#)). However, the optimization of the existing gene editors, the development of new gene editors, and continued research on the sex-determining gene will aid in breaking the barrier. However, there has been a long running debate over whether bio-technologies should be applied into food production. Although, many of GM (genetically modified) organisms are already deemed safe, the safety of GM food has been questioned by the general public ever since their introduction in the late 90's ([44](#), [45](#)), which hindered the development of bio-technologies in food production. Therefore, government may play an important role in driving the bio-technologies in food production forward by setting stricter food quality standards, improving legal regulation on GM food, and choosing a mandatory GM food labeling system to protect the consumer's right to know. Changing the sex ratio of offspring at the gene level cannot only promote the development of livestock production but also provide a therapeutic biological model for human diseases.

RNA interference (RNAi) is a biological process that uses small interfering RNA (siRNA) to induce sequence-specific gene silencing through mRNA degradation ([46](#)). Since its first use in mammalian cells in 2001, RNAi technology has successfully been implemented in mammalian gene silencing systems ([47](#)). Genes such as *Sry*, *zfy*, and *zfx* that are known to affect sex

determination during spermatogenesis could be silenced through RNAi to influence the sex ratio of animal offspring.

Research has demonstrated that injecting RNAi vectors into mouse testis can skew the sex-ratio of offspring ([48](#)). In a separate study, knocking out *Sry* mRNA by direct injection of siRNA into pregnant mice through the tail vein generated mouse embryos with feminized gonads ([49](#)).

The mechanisms of these sex-determining genes in spermatogenesis and sex determination are uncertain, and very little is known on whether these genes could be knocked-down using RNAi technology or the CRISPR/Cas9 system. This is because the expression level of mRNA is influenced by several factors including non-specific creation, the activation of innate immune response, and off-target effects. In addition, transfection protocols yield low efficiency for primary cells, while suspension requires optimization for different cell types. Besides, poor bioavailability, rapid hydrolysis, and the inability to cross biological barriers such as the blood-brain barrier are some of the major concerns that hinder RNAi adoption. However, it has been established that exosomes can transfer siRNAs with high target specificity and without any immunologic reactions as a natural vehicle, thereby providing a new method for delivering of siRNAs into the cells ([50](#)).

Primarily, RNAi is a simple method that has had a significant effect on the expression of the sex-determining gene. Future studies should therefore explore the mechanisms of sex-determining genes in spermatogenesis and sex determination, improve transfection efficiency, and reduce

electropuncture damage. Besides, using exosome as a vehicle for targeted delivery has good prospects for the future.

Sperm-Separating Technologies in Sex Manipulation

Several methods have been previously used for separating X- and Y-bearing sperm. The initial sperm sexing procedures designed for sex determination involved DNA probes ([51](#), [52](#)), albumin gradients ([53](#), [54](#)), Percoll gradients ([55](#), [56](#)), the Sephadex gel-filtration method ([57](#) - [59](#)), and “swim-up” procedures ([60](#), [61](#)). However, the efficiency of these methods in distorting the sex ratio of livestock offspring has been unconvincing ([62](#)). Additionally, all these methods can damage sperm motility. Therefore, flow cytometry and immunology sexing are the main sperm separating technologies presently used in livestock production at present.

Flow Cytometry in Sex Manipulation

Johson et al. was the first to use flow cytometry to separate sperm according to the difference of DNA content in the head of X- and Y-bearing sperm (the DNA content of X-bearing sperm is 3-4% higher than that of Y-bearing sperm) in 1986 ([63](#), [64](#)). Since then, flow cytometry-based sperm sexing has produced millions of pre-sexed offspring in different species including, rabbits, pigs, cattle, sheep, horses, dogs, cats, etc.

Generally, contemporary sperm-sorting procedures yield a skewed offspring sex ratio of between 85 and 95% accuracy ([65](#)). Flow cytometry has since reached commercial application in cattle ([66](#)), and the quality of sperm for use in a flow sorter is dependent on the species. Therefore, the sorting index of different species is calculated by multiplying the percentage of the

difference in DNA content of the X- or Y-bearing sperm by the area of the flat profile of the sperm head. The sorting index indicates that bull and boar sperm are well-suited for separation in a flow sorter (133 and 115, respectively) ([67](#)). However, use of flow cytometry to sort sperm is minimal in pig production. This could be a result of boar sperm being more susceptible to the influence of flow cytometry since the sorting process damages the boar sperm membrane, which leads to the decline in its vitality, fertilization ability, storage ability, and subsequent processing ability ([68](#) , [69](#)). In addition, the sperm used for artificial insemination per pregnancy in pig production is often significantly higher than in cattle production ([70](#) , [71](#)).

Initial studies focused on identifying the steps in flow cytometry that caused sperm damage. It was earlier thought that optimization of sperm dilution rate ([72](#) - [74](#)), as well as changing the type of dyes and laser power ([75](#) - [77](#)) could reduce sperm damage. However, these methods could not significantly increase sex-sorted sperm motility. This prompted other researches to focus on improving hardware, optics, electronics, and handling protocols before flow cytometry such as reducing the sheath pressure of the sorter from 50 to 40 psi ([78](#)), implementation of pulsed lasers ([79](#)), or transportation of red deep sperm at high sperm concentration without staining before sorting ([80](#)). Although these improvements increased the motility of sperm after separation, subsequent optimization is limited by current mechanical technology. Besides, improving the production speed and sperm quality remains a significant challenge, thereby shifting attention to related technology.

In a study that aimed to describe the effects of flow cytometry on the structure of the glycocalyx of bovine sperm by using lectin histochemistry, both flow cytometry and capacitation had similar effects on bovine sperm, which may account for the low conception rates observed after flow cytometry ([81](#)). Another study that combined AA-2G supplementation and deep insemination to improve the fertility of sex-sorted sperm from Cashmere goats revealed a substantial protective effect on the quality and functions of sperm ([82](#)). Although some researchers prefer artificial insemination technology, laparoscopic insemination (LI) at the uterine horns can achieve a high fertilization rate with a low sex-sorted sperm dose (five times less than intrauterine insemination) ([83](#)). However, improvements such as the outputs of the flow sorters, aspects related to the synchronization protocols, the time of ovulation, and single LI at the uterine horns deserve to be optimized in future studies ([84](#)).

In conclusion, flow cytometry is the most used and mature technology for sperm sexing in livestock production, with a separation accuracy of above 90%. However, due to the high sperm requirement and susceptibility of other species, this strategy has only reached commercial application in cattle production. Future studies should therefore focus on reducing sperm damage caused by flow cytometry, develop related technologies, and optimize artificial insemination technology.

Immunology Sexing in Sex Manipulation

Immunology sexing is achieved through the identification and separation of sex-related proteins. This is mainly done by detecting the H-Y antigen on the plasma membrane of Y-bearing sperm by using the H-Y antibody. Thus, far, <https://assignbuster.com/sex-manipulation-technologies-progress-in-livestock-a-review/>

sperm from many species has been successfully separated including mice ([85](#)), rabbits ([86](#)), and sheep ([87](#)). In addition, because of the simplicity, rapidity, accuracy, and reasonable price of this method, it can be used to verify the purity of sorted X- and Y-bearing sperm ([88](#)).

A recent study successfully separated sperm from mice by combining the immunology sexing and swim-up methods, thereby providing a strategy for immunology sexing. In 2019, three Japanese scientists found that Toll-like receptor 7/8, which is encoded by X chromosome, could influence sperm motility. The motility of X sperm reduces when incubated with R848 (a synthetic agonist which can bind both TLR8 and TLR7) as a result of Toll-like receptor 7/8 binding R848. However, the motility of the Y sperm is not affected. Thus, the sperm is selectively separated by using the swim-up method, in which Y sperm with high motility swims to the upper layer of the medium, while the X sperm sinks to the lower layer of the medium. Besides, R848 had no effect on the acrosome of both X sperm and Y sperm, indicating that sperm fertility is not affected even after incubating with R848. In addition, the motility of X sperm returns to normal once R848 has been washed off. The sperm separated by this method was later used for *in vitro* fertilization and embryo transfer. In the results, upstream sperm produced 77 blastocyst embryos, among which 68 were XY embryos and 9 were XX embryos, while downstream sperm produced 83 blastocyst embryos, among which 58 were XY embryos and 25 were XX embryos ([89](#)).

However, most immunology sexing strategies affect the motility of sperm and reduce the conception rate due to the long processing time. Although

this strategy is presently at the laboratory stage, advances in molecular biology may provide more breakthroughs. Therefore, more efforts should be concerted on the discovery of new sex-related proteins and the blending of other strategies to reduce sperm damage.

Sex Manipulation Within the Female Reproductive Tract

Millions or billions of sperm are deposited into the reproductive tract after mating, but only a few sperm reach the ampulla or the site of fertilization and only one fertilizes an oocyte. The journey that sperm take to reach an oocyte is long and perilous. The female environment contributes toward this successful transit by providing a vehicle for sperm transport, aiding the removal of dead sperm and other pathogens and applying strict selection pressures to ensure only those cells with the highest quality reach the site of fertilization. Besides, in many locations of the female reproductive tract, sperm interact with the epithelium and the luminal fluid, which can affect sperm motility and function ([90](#), [91](#)). Although we know little about this complex interaction between sperm and female environment, many researches sought to reveal the impact of the mammalian female on the sex of her posterity.

By introducing X- or Y-bearing sperm into two separate oviducts of single female pigs using bilateral laparoscopic insemination, researchers demonstrated that the oviduct can screen sperm by modifying the oviduct environment ([92](#)). Besides, research has shown that both time of ovulation and maturational state of oocyte can influence the sex ratio of animal offspring ([93](#), [94](#)). This led to the investigation of the effect of follicular

fluid composition on the sex of an embryo, which was confirmed in subsequent studies ([95](#), [96](#)). In addition, various studies have reported the impact of pH on sperm motility, viability, and capacitation ([97](#)). However, pH did not influence the X- or Y-bearing sperm of bovine *in vitro* ([98](#)). Therefore, extensive studies are needed to examine the effect of external pH on X- or Y-bearing sperm. We speculate that pH might affect X- or Y-bearing sperm by altering other physiochemical factors in the bovine oviduct. Furthermore, there have been several papers suggesting the effect of several glycoproteins and factors from the female reproductive tract on sperm survival including L-selectin ([99](#)), PMCA4 ([100](#)), fibronectin ([101](#)), SBG ([102](#)), etc. Hence, manipulating the sex of the offspring by regulating the expression level of these glycoproteins and factors is possible and could be a new direction for future research.

Above all, although sex manipulation in the female reproductive tract is the most parsimonious strategy proposed thus far, it has the potential of resolving the formerly puzzling and seemingly contradictory findings. With continued research, a better understanding of female influence in sex determination can be attained and could have a huge impact on existing sex manipulation strategies.

Embryo Sexing in Sex Manipulation

Since artificial insemination and embryo transfer technology were implemented in commercial field, research on finding embryo sexing method has become a point of interest for modern researchers ([103](#)). Obstetric ultrasonography is the most common imaging technique in veterinary

reproduction including: confirming pregnancy, identifying congenital fetal defects, defining fetal sex, etc., because it is simple, reliable, and non-invasive ([104](#), [105](#)). However, the sex of mammalian fetus cannot be performed by ultrasonographic examination until several weeks after pregnancy [e. g., 55–85 days in cattle ([106](#)), 59–68 days in horse ([107](#)), 46–55 days in goat ([108](#))], which does not meet the requirements of embryo transfer as the main stage at which embryo transfer is performed is the cleavage stage (day 2–4 after fertilization) or the blastocyst stage (day 5 or 6 fertilization) ([109](#)). Thus, a large number of techniques for genetic sex identification of early embryo in mammals have since been developed. Before Barr body formation ([110](#)) or detection of H-Y antigen ([111](#), [112](#)), methods used were based on cytogenetic analysis ([113](#), [114](#)) and measurement of level of X-linked enzymes (glucose-6-phosphatase dehydrogenase, G6PD). However, due to the high embryo requirement, low accuracy, and the potential of embryo injuries, these methods were replaced by molecular detection.

The blastomere of mammalian embryos is omnipotent and can develop into a complete animal. Thus, each blastomere contains all the genes required for the development of an animal, including genes involved in sex determination such as *Amg* ([115](#)), *Zfy* ([116](#)), and *Sry* ([117](#)). Molecular biologic detection techniques, which include using nucleic acid probe, fluorescence *in situ* hybridization (FISH), polymerase chain reaction (PCR), and loop-mediated isothermal amplification (LAMP), can therefore be used to identify the male-specific gene.

Among nucleic acid probes, isotopic labeled probes have high accuracy but require a lot of embryo cells and time, while biotin-labeled probes require little time to detect but can only separate few embryos ([118](#)). FISH, on the other hand, has high accuracy and rapidity and can display a variety of colors due to the emergence of multi-color fluorescence *in situ* hybridization. FISH is however expensive and complex to operate ([119](#)), thereby leaving PCR and LAMP as the mainstream methods for embryo sexing in sex manipulation presently.

PCR is widely used, and besides its ability to analyze the phenotypes based on amplification products of sex-specific genes, it can verify the purity of sorted X- and Y-bearing sperm statistically. To identify the sex of embryos ([120](#)) through PCR, a pair of previously developed sex-specific primers was used. In 1991, this method successfully identified the sex of cattle and sheep embryos for the first time ([121](#)). However, since this method only amplified a pair of sex-specific primers, lack of amplification would correspond to a negative result. This method was therefore replaced by multiplex PCR in which a pair of sex-specific primers with at least one pair of internal primers that amplify house-keeping genes ([122](#)) was designed and utilized.

However, due to the large number of cells required in the conventional PCR method and the limited sensitivity of one amplification, the method was prone to errors. Therefore, nested PCR and multiplex PCR were later combined to improve the accuracy of detection results. Besides increasing the amount of amplification product and the test's accuracy, the combination reduced the number of required cells to 3–8. In addition, conventional PCR has some disadvantages such as a complicated identification process and

long identification time. Until 1992, Cha et al. adopted two-temperature gradient PCR to obtain the ideal amplification results. This proved that DNA polymerase could rapidly catalyze a complete amplification process from annealing to degeneration without extension steps when the fragment amplified is small (<200 bp). Not only did this retain the advantages (high sensitivity, high accuracy, low embryonic damage) of conventional PCR, but it also simplified the conventional PCR and shortened the entire process from 2-4 h to 1 h ([123](#)).

Presently, PCR can be used for further analyses, such as genotyping and molecular diagnosis in preimplantation embryos besides embryo sexing ([124](#)). Moreover, it is feasible for automation and high-throughput sex-typing ([125](#)). Though not suitable for the sex manipulation of twin pregnancies ([126](#)), research has demonstrated that sex manipulation using the DNA of free fetal cells in maternal plasma is efficient for both *SRY* and *AMEL* gene sequences.

Loop-mediated isothermal amplification (LAMP) is a method of DNA amplification that was developed by Eiken Chemical Co. Ltd. in 2000 and works under isothermal conditions. This method of DNA amplification is highly specific, efficient, and rapid ([127](#)). Besides, gene amplification and detection can be completed in one step, and amplification could be up to 10^9 - 10^{10} times in 15-60 min. Since the detection of all target gene sequences can only be determined by either the presence or absence of amplification products, LAMP is considered to be highly specific. Recently, LAMP primers with polyethylenimine (PEI) for precipitation were optimized to improve

sensitivity, and multiplex LAMP was used to improve accuracy ([128](#)). The first LAMP assay for the detection of the *SRY* gene was performed using DNA extracted from 15 blood samples from pregnant women at 8 weeks ([129](#)).

Although PCR and LAMP have made significant progress in embryo sexing owing to their high sensitivity, the impact of the pollution in the external environment on the accuracy of the results remains inevitable. In addition, these two methods are difficult to use in livestock production due to the large sample size and the poor transfer pregnancy rate of recent embryo transfer technology. Therefore, future research should focus on improving tolerance to external environment and the efficiency of these technologies. Besides, these technologies should be applied to identify the accuracy of other sex manipulation strategies, rather than as a sex manipulation strategy.

Conclusion

Sex manipulation technologies are valuable biotechnologies that have the potential to revolutionize cattle production. With the development of molecular biology and cell biology, these emerging sex manipulation strategies significantly expand the ability to bias the sex ratio of animal offspring and would be instrumental in correcting clinical male-to-female sex reversal syndrome in humans. However, the knowledge of sex-determining mechanisms, whether sex-determining gene or female reproductive tract, is still very limited. Since different sex manipulation strategies have their individual advantages and disadvantages, the selection of a specific system is dependent on the expertise of the individual researcher rather than on the

weaknesses of one of these strategies. Collectively, sex determination is a complex process that is influenced by a variety of internal and external factors. Further exploration of sex-determining mechanisms; continuous improvement of the accuracy, sensitivity, and economic benefits of sex manipulation technologies; and the establishment of a new a sex manipulation system that integrates knowledge from various fields can provide great impetus in livestock production and solve the problem of food shortage.

Author Contributions

YX and ZX: conceived the idea, conducted the literature review, and drafted the manuscript. ZW and LH: project investigator, decided a review topic, and revised the manuscript. All the authors have read and approved the final version of the manuscript.

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Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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