

Editorial: autophagy and related transcription factors in liver and gut diseases

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Editorial on the Research Topic

Autophagy and Related Transcription Factors in Liver and Gut Diseases

The cell biologist Yoshinori Ohsumi received the 2016 Nobel Prize in Medicine for his early identification and characterization of the autophagy machinery, in particular, AuTophagy-related (Atg) genes, in yeast. Macroautophagy (hereafter, autophagy) is a cytoprotective pathway for sequestration of cellular components (such as misfolded proteins, damaged organelles, and excessive lipids) into autophagosomal vesicles, followed by clearance *via* the lysosomal system ([Galluzzi et al., 2017](#)). Autophagy is specifically upregulated upon exposure to various stressors such as oxidative and endoplasmic reticulum stress, thus aiding in the prevention of various pathologies. Therefore, autophagy dysregulation may be involved in inflammatory, metabolic, toxic, and infectious diseases and cancer ([Kroemer et al., 2010](#) ; [Eid et al., 2013](#) ; [Horibe et al., 2017](#)). Most organelles also seem to have selective programs of autophagy, including mitochondria, lipid droplets, endoplasmic reticulum, and even lysosomes. Selective autophagic removal of damaged mitochondria, or mitophagy, is an anti-apoptotic mechanism induced and specifically upregulated in response to various damaging agents such as binge ethanol exposure or drug-induced liver injury in animal models ([Otsuki et al., 1994](#) ; [Youle and Narendra, 2011](#) ; [Lemasters, 2014](#) ; [Eid et al., 2016](#) ; [Eid et al., 2019](#)). Autophagy can be regulated not only at the gene level, but its final performance can be modulated by lysosomal lipid composition. For instance, accumulation of lipids (e. g., cholesterol) in lysosomes has been shown to impair the fusion of autophagosomes (containing disrupted mitochondria) with lysosomes,

contributing to the perpetuation of damaged mitochondria, which sensitizes to acetaminophen hepatotoxicity ([Baulies et al., 2015](#)). On the other hand, autophagic clearance of lipid droplets is referred to as lipophagy ([Singh and Cuervo, 2012](#)). Various transcription factors such as transcription factor EB (TFEB), Nrf2, HIF, and Foxo3a play important roles in the regulation of autophagy and mitophagy-related proteins such as LC3, cathepsins, and Parkin ([Sardiello, 2016](#) ; [Horibe et al., 2017](#) ; [Eid et al., 2019](#)). The focus of this Research Topic is to highlight the involvement of these transcription factors in the regulation of liver and gut diseases through autophagy pathway as these are potential therapeutic targets for the restoration of autophagy and in the management of these diseases.

This Research Topic compiles nine articles, including four reviews and five original research contributions. The interesting review by Su et al. , on *Mitophagy in Hepatic Insulin Resistance: Therapeutic Potential and Concerns* , focuses on advances in the understanding of relationship between mitophagy and hepatic insulin resistance and the potential value of mitophagy in the treatment of hepatic insulin resistance and metabolic syndrome (*via* clearance of damaged mitochondria and subsequent reduction of lipid accumulation). This observation is supported by an elegant study demonstrating that loss of Parkin-mediated mitophagy promoted further β -cell failure under pathological stress conditions including STZ exposure and leptin receptor defects ([Hoshino et al., 2014](#)).

Recent advances with incretin-associated drugs have opened new avenues in the management of diabetes. In another interesting review article,

Kanasaki et al. analyzes distinct molecular mechanisms of autophagy regulation by glucagon, GLP-1, and DPP-4 inhibitor. In addition, they also discuss the potential contribution of these regulatory pathways in the induction of beneficial autophagy-upon bariatric surgery, which have implications in the treatment of diabetic diseases ([Adeghate et al., 2019](#)).

Lipophagy, a process controlled by the autophagy master regulator, TFEB, is key to maintaining a healthy liver. The third review by Yang et al. discusses the different lipophagic responses in rodent hepatocytes after exposure to acute and chronic ethanol. They showed that these responses are controlled by subcellular TFEB localization. They suggest that natural products and drugs such as caffeine/coffee, resveratrol, corosolic acid, zinc, carbamazepine, and rapamycin may activate autophagy/lipophagy for preventing or even aiding in the treatment of alcohol-induced fatty liver. In addition, they stress that the specific upregulation of TFEB by certain small molecules (related to digoxin, ikarugamycin, and alexidine dihydrochloride) may be of therapeutic value in the treatment of human fatty liver disease ([Wang et al., 2017](#)).

In another review article, Zhang L. et al. elegantly summarize the current understanding on the use of herbal medicine extracts and natural products for activation of hepatic autophagy, thus helping in the prevention and treatment of non-alcohol fatty liver diseases (NAFLD). A specific focus is set on mechanisms by which autophagy can target the main events in the pathogenesis of NAFLD, including hepatic steatosis, inflammation, oxidative stress, and apoptosis.

The research article by Fan et al. provides novel data supporting a protective role for methylprednisolone (MP) in an experimental autoimmune hepatitis (AIH) model, possibly mediated by the Akt/mTOR signaling pathway. MP seems to ameliorate apoptosis and promote autophagy in hepatocytes in *in vitro* and *in vivo* mouse model. They suggest a potential use of MP to treat AIH. Their study provides interesting insights into the mechanisms underlying the effect of MP on hepatocytes.

The interesting study by Guo et al. explores the effects of 6-bromo-indirubin-3'-oxime (6BIO), a potent inhibitor of glycogen synthase kinase-3 (GSK-3), on the aging rodent liver. They found that 6BIO mitigates oxidative stress, improves lipid metabolism, enhances autophagy, and significantly retards liver aging *via* modulating the GSK-3 β and mTOR pathways. They suggest that 6BIO could be a potential agent to protect the liver in the field of anti-aging pharmacology.

Hepatitis C virus (HCV) dysregulates lipid metabolism to accomplish several steps of its life cycle ([Paul et al., 2014](#) ; [Strating and van Kuppeveld, 2017](#)). Vescovo et al. investigates the impact of mevastatin (a cholesterol-lowering agent isolated from *Penicillium citinum*) on HCV replication and autophagy in MMHD3 non-transformed hepatocytes harboring sub-genomic HCV replicons, specifically in relation to the extracellular lipid uptake. In contrast to the previous studies in transformed human cell lines, they observed drastic upregulation of intracellular cholesterol in MMHD3 cells upon mevastatin treatment, which is associated with enhanced lipophagy and HCV replication. However, these effects are reversed when cells are cultured in

delipidated serum, which establishes the fact that suppression of extracellular lipid uptake is as important as inhibiting cholesterol biosynthesis in suppressing HCV replication. This study may have implications in the development of treatment modalities targeting cholesterol levels to limit HCV replication.

Baulies, A., Ribas, V., Núñez, S., Torres, S., Alarcón-Vila, C., Martínez, L., et al. (2015). Lysosomal cholesterol accumulation sensitizes to acetaminophen hepatotoxicity by impairing mitophagy. *Sci. Rep.* 5, 18017. doi: 10.1038/srep18017

Eid, N., Ito, Y., Otsuki, Y. (2013). The autophagic response to alcohol toxicity: the missing layer. *J. Hepatol.* 59, 398. doi: 10.1016/j.jhep.2013.03.038

Eid, N., Ito, Y., Otsuki, Y. (2016). Triggering of Parkin mitochondrial translocation in mitophagy: implications for liver diseases. *Front. Pharmacol.* 7, 100. doi: 10.3389/fphar.2016.00100

Eid, N., Ito, Y., Horibe, A., Otsuki, Y., Kondo, Y. (2019). Ethanol-induced mitochondrial damage in sertoli cells is associated with parkin overexpression and activation of mitophagy. *Cells* 8, 283. doi: 10.3390/cells8030283

Galluzzi, L., Baehrecke, E. H., Ballabio, A., Boya, P., Bravo-San Pedro, J. M., et al. (2017). Molecular definitions of autophagy and related processes. *EMBO J.* 36, 1811–1183. doi: 10.15252/embj.201796697

Horibe, A., Eid, N., Ito, Y., Hamaoka, H., Tanaka, Y., Kondo, Y. (2017).

Upregulated autophagy in Sertoli cells of ethanol-treated rats is associated with induction of inducible nitric oxide synthase (iNOS), androgen receptor suppression and germ cell apoptosis. *Int. J. Mol. Sci.* 18. doi: 10.3390/ijms18051061

Hoshino, A., Ariyoshi, M., Okawa, Y., Kaimoto, S., Uchihashi, M., Fukai, K., et al. (2014). Inhibition of p53 preserves Parkin-mediated mitophagy and pancreatic β -cell function in diabetes. *Proc. Natl. Acad. Sci. U. S. A.* 111, 3116–3121. doi: 10.1073/pnas

Kroemer, G., Marino, G., Levine, B. (2010). Autophagy and the integrated stress response. *Mol. Cell.* 40, 280–293. doi: 10.1016/j.molcel.2010.09.023

Lemasters, J. J. (2014). Variants of mitochondrial autophagy: types 1 and 2 mitophagy and micromitophagy (type 3). *Redox Biol.* 2, 749–754. doi: 10.1016/j.redox.2014.06.004

Lv, H., Yang, H., Wang, Z., Feng, H., Deng, X., Cheng, G., et al. (2019). Nrf2 signaling and autophagy are complementary in protecting lipopolysaccharide/d-galactosamine-induced acute liver injury by licochalcone A. *Cell Death Dis.* 10, 313. doi: 10.1038/s41419-019-1543-z

Muhammad, I., Wang, X., Li, S., Li, R., Zhang, X. (2018). Curcumin confers hepatoprotection against AFB1-induced toxicity *via* activating autophagy and ameliorating inflammation involving Nrf2/HO-1 signaling pathway. *Mol. Biol. Rep.* 45, 1775–1785. doi: 10.1007/s11033-018-4323-4

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Otsuki, Y., Misaki, O., Sugimoto, O., Ito, Y., Tsujimoto, Y., Akao, Y. (1994). Cyclic bcl-2 gene expression in human uterine endometrium during menstrual cycle. *Lancet* 2 (344), 28–29. doi: 10. 1016/s0140-6736(94)91051-0

Paul, D., Madan, V., Bartenschlager, R. (2014). Hepatitis C virus RNA replication and assembly: living on the fat of the land. *Cell. Host Microbe* 16, 569–579. doi: 10. 1016/j. chom. 2014. 10. 008

Sardiello, M. (2016). Transcription factor EB: from master coordinator of lysosomal pathways to candidate therapeutic target in degenerative storage diseases. *Ann. N. Y. Acad. Sci.* 1371, 3–14. doi: 10. 1111/nyas. 13131

Singh, R., Cuervo, A. M. (2012). Lipophagy: connecting autophagy and lipid metabolism. *Int. J. Cell Biol.* 282041, 1–49. doi: 10. 1155/2012/282041

Strating, J. R., van Kuppeveld, F. J. (2017). Viral rewiring of cellular lipid metabolism to create membranous replication compartments. *Curr. Opin. Cell Biol.* 47, 24–33. doi: 10. 1016/j. ceb. 2017. 02. 005

Wang, C., Niederstrasser, H., Douglas, P. M., Lin, R., Jaramillo, J., Li, Y., et al. (2017). Small-molecule TFEB pathway agonists that ameliorate metabolic syndrome in mice and extend *C. elegans* lifespan. *Nat. Commun.* 9, 2050. doi: 10. 1038/s41467-017-02332-3.

Youle, R. J., Narendra, D. P. (2011). Mechanisms of mitophagy. *Nat. Rev. Mol. Cell Biol.* 12, 9–14. doi: 10. 1038/nrm3028