

Is there a role for treating inflammation in moyamoya disease?: a review of histo...

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Introduction

Moyamoya disease is a slowly progressing steno-occlusive condition affecting the cerebrovasculature. Affecting the terminal internal carotid arteries (ICA) and their branches, bilaterally, resulting in a fine vascular network in the base of the brain to allow for compensation of the stenosed vessels ([1](#)). The most distinguishing feature of this condition is the progressive stenosis of the ICA which induced further dilation of the perforating arteries that function as collateral pathways. Moyamoya disease is rare, with the incidence highest in East Asia of 0.35-0.94 and a prevalence of 3.16-10.5 per 100,000 ([2](#), [3](#)). The annual incidence is reported to be approximately 10% that of Japan ([4](#)). A bimodal age distribution has been observed with the predominant peak originally occurring at 5 years of age followed by a lower peak at 40 years ([3](#)).

There is significant debate over the pathogenesis involved in moyamoya disease, while there is obvious evidence of the involvement of inflammatory proteins, this has historically not been acknowledged as a causal factor in the condition ([5](#)). Here we describe the histopathology, genetics, and signaling cascades involved in moyamoya and debate whether these factors can be linked as causal factor for the condition or whether they are simply a result of the ischemia described in the condition. As such we hope to draw some novel insight into potential diagnostic and therapeutic inflammatory targets in the condition.

Histopathology

The histopathology of moyamoya disease has stirred interest for a number of years. Stenotic changes occur in the intracranial ICAs immediately distal to the bifurcation. Progression later involves the proximal anterior and middle cerebral arteries and on rare occasions the posterior circulation may become involved. In adults, Weinberg et al. ([6](#)) describe a typical pattern of fibrocellular thickening of the tunica intima with excessive proliferation of the vascular smooth-muscle cells, marked tortuousness of the internal elastic lamina and attenuation of the tunica media ([7](#)). Moyamoya vessels have fibrin deposits in their walls, fragmented elastic laminae, attenuated media, and microaneurysms ([8](#)). Thrombosis as a result of collapse of the lumen is frequently observed in the vessels of patients ([9](#)).

This particular pathology is considered to be responsible for the onset of both ischemic and hemorrhagic stroke in these patients. Interestingly, the consensus view is that at a histological level these vessels lack inflammatory change, which has been considered to rule out an inflammatory component to the condition ([5](#)). However, Masuda et al. ([10](#)) noted the infiltration of macrophages and T cells in non-stenosed areas of the vessels, suggesting that the microthrombi may be a result of the chronic inflammation rather than a cause. In either case, the observation of microthrombi is not specific to moyamoya disease and therefore is unlikely to provide a complete explanation for its pathogenesis.

Although limited by the number of cases involved, there appears to be consensual evidence of inflammation in moyamoya. The lack of animal

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models makes it difficult to ascertain whether these findings contribute to the induction of the condition. Nevertheless, with growing evidence that inflammation is present in the vessel walls the debate of whether this process induces or results from moyamoya may be somewhat academic. The important factor is that there is reversible process in the vessels contributing to stenosis and as such significant questions have to be raised about whether anti-inflammatory agents could play a role in the treatment of the condition.

Genetics

Moyamoya disease has a high familial occurrence accounting for up to 15% of affected patients ([11](#)). The female to male ratio in familial moyamoya disease is 5.0, which is much higher than that in sporadic cases (1.6). The mean age at onset of familial moyamoya disease (11.8 years) is significantly lower than that in sporadic cases (30.0 years). Interestingly, among parent-offspring pairs, the age at onset of offspring is on average 23 years lower than of parents, suggesting strong association with anticipation in familial moyamoya disease ([12](#)).

A number of linkage analysis have demonstrate the involvement of inflammatory genes in moyamoya disease. In particular, Ikeda et al. ([13](#)) demonstrated an association with chromosome 3 and specifically the locus responsible for the maintenance of vascular wall homeostasis. Chromosome 3p a principle site of proteins involved in multiple signaling cascades most notably the *IL5RA* (interleukin-5 receptor alpha), *TGFBR2* (transforming growth factor beta receptor II), *THRB* (thyroid hormone receptor beta), *RARB* (retinoic acid receptor beta), and *PPARG* (peroxisome proliferator-activated

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receptor gamma) are all involved in intricate signaling pathways which control and regulate angiogenic and inflammatory pathways.

Similarly, an association with human leukocyte antigens (HLA) ([14](#)) located on chromosome 6, the 6q25 marker was shared by 84% of families in a recent study. HLA has a strong connection with immune disorders ([15](#)). In particular, alterations in gene transcription and protein folding have been linked to aberrant expression of endothelial-leukocyte adhesion molecule-1 (E-selectin or ELAM-1), vascular cell adhesion molecule-1 (VCAM-1), and intercellular adhesion molecule-1 (ICAM-1) is induced by the inflammatory cytokines interleukin-1 beta (IL-1 β) and tumor necrosis factor-alpha (TNF- α) via the activation of the pro-inflammatory transcription factor nuclear factor kappa b (NF- κ B) ([16](#)).

It is clear that there is a panoply of genes activated in moyamoya disease which have an inflammatory association. Whether, these are responsible for induction of the condition or a result upstream change remains unclear. Very little in moyamoya disease has been translated into animal models. Most recently, the discovery of RNF213 as a susceptibility gene has stirred some interest ([17](#), [18](#)). RNF213 encodes a gene finger protein with an AAA ATPase domain and is abundantly expressed in spleen and leukocytes ([17](#)). An RNA *in situ* hybridization analysis of mouse tissues indicated that mature lymphocytes express higher levels of Rnf213 mRNA than their immature counterparts ([17](#)). Recent studies have suggested that the postnatal vasculature can form through vasculogenesis, a process by which endothelial progenitor cell are recruited from the splenic pool and differentiate into

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mature endothelial cells ([19](#)). Levels of endothelial progenitor cells in the peripheral blood are increased in moyamoya disease patients ([20](#)). It is postulated that RNF213 may be expressed in splenic endothelial progenitor cells and mutant RNF213 might dysregulate the function of the endothelial progenitor cells ([17](#)). However, further research is necessary to elucidate the role of RNF213 in the etiology of the condition.

Signaling Cascades

Cellular signaling cascades provide the interface of genetic and environmental interaction. An understanding of the cellular signaling cascades which are involved in a condition provides a platform for the identifying both diagnostic and therapeutic targets. This stems from the understanding that infection may play a key role in the pathogenesis of moyamoya disease ([21](#)).

The aberrant expression of mitogens, adhesion molecules, and angiogenic factors ([22](#) - [25](#)) and/or alterations in cellular responses to growth factors and cytokines demonstrate the involvement of inflammatory and hematopoietic cascades in vascular cells ([26](#)). This has been postulated to play a crucial role in the development of moyamoya pathogenesis ([10](#)).

Vascular Endothelial Growth Factor

Vascular endothelial growth factor is a 45-kD homodimeric, basic glycoprotein that requires association with heparin in order to function ([27](#)). It plays a central role in pathological vasculogenesis and vascular permeability in intracranial lesions. Similarly, VEGF has been shown to promote angiogenesis in the setting of cerebral ischemia ([28](#)). Interestingly, <https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

VEGF expression has been observed to being NF-kappa B dependent in a number of tissues, including endothelial cells ([29](#)). Furthermore, studies have shown the upregulation of Prox1 ([30](#)) and downregulation of Notch-1 ([31](#)) to have correlating effects on angiogenic processes. This regulation of angiogenesis by a pro-inflammatory transcription factor has drawn insight into the potential signaling cascades available for manipulation in moyamoya disease.

Aberrant expression of VEGF is evident around the affect vasculature in moyamoya disease. In a small study, Sakamoto et al. ([28](#)) observed a fourfold increase in VEGF expression in patients with moyamoya disease. More specifically, the VEGF –634G allele has been identified has having a particularly strong influence on moyamoya disease and poor collateral vessel formation ([32](#)). The expression of VEGF is not limited to the cerebral vasculature in moyamoya disease, Takekawa et al. ([33](#)), describes the growth factor in glial cells and Sakamoto et al. ([28](#)) in the dura matter. The authors suggest that this displays evidence of the pathological mechanisms extending beyond the cerebral vasculature. Nevertheless, this is most likely associated with the induction of pro-inflammatory cascades as a result of ischemia and a secondary marker of disease rather than a primary modality for the pathogenesis.

Basic Fibroblast Growth Factor

Basic fibroblast growth factor (bFGF) is an 18-kD protein consisting of 146 amino acids ([34](#)). The primary role of bFGF involves the stimulation of mesodermal and neuroectodermal proliferation ([35](#)), additionally it has

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been shown to induce growth of vascular smooth muscle and, when combined with VEGF, can play a leading role in angiogenesis ([36](#)). A hypothesized pathway for this is via the upregulation of circulating chemokines ([37](#)). In particular, the chemokine-mediated regulation of angiogenesis is highly sophisticated and fine tuned, and involves pro-angiogenic chemokines, for instance, CXCL8/IL8 interacting with the CXCR2 receptor, and anti-angiogenic (i. e., angiostatic) chemokines, for instance, CXCL10/IP10 interacting with the CXCR3 receptor ([37](#)).

Basic fibroblast growth factor has been observed to be aberrantly expressed in the colony-stimulating factor (CSF) of moyamoya disease patients ([24](#), [38](#)), with both groups agreeing that the expression levels were 10-fold higher in moyamoya disease. Additionally, bFGF was also observed in the thickened tunica media, assisting suggestions that the upregulation of the molecule is associated with both stenotic and angiogenic processes ([38](#)). Yan et al. ([39](#)) describes separate *in vitro* and *in vivo* models of bFGF promoting neovascularization. In relation to corneal wound healing, a comparison with recombinant human epidermal growth factor (rhEGF) led the others to believe that the effects of bFGF were too strong to promote controlled healing ([39](#)). Interestingly, the inhibition of bFGF has been demonstrated to inhibit the proliferation and migration of endothelial cells ([40](#)). As such it could be viewed that bFGF plays an intricate role in the development of vessel proliferation and endothelial cell recruitment. With this in mind there may be future scope to incorporate the molecule into new therapeutic strategies.

Hepatocyte Growth Factor

Hepatocyte growth factor (HGF) is one of the largest disulfide-linked cytokines, and in humans the protein is synthesized as a single-stranded 728 amino acid protein ([6](#)). The proteolytic activation of HGF involves the release of a 31 amino acid N-terminal signal peptide which has been observed to potentiate the growth of various epithelial, endothelial, and mesenchymal cells ([41](#)).

In various injury and disease models, the HGF-Met pathway plays a critical role in acute tissue protection and regeneration, and in providing less susceptibility to chronic inflammation and fibrosis ([42](#)).

Nanba et al. ([43](#)) demonstrated a twofold increase in both HGF and its receptor c-Met expression in the tunica media and intima of patients with moyamoya disease compared to control groups with cervical spondylosis and unilateral internal carotid artery occlusion. From this it was postulated that the upregulation of HGF plays a role in the pathogenesis of intimal thickening and vascular smooth-muscle cell migration. Additionally, hypoxia inducible factor-1 α , which promotes smooth-muscle cell proliferation in the presence of bFGF and HGF, is present in elevated levels in moyamoya disease ([44](#)). In addition to HGF being densely found in the carotid fork, its CSF level is markedly elevated in moyamoya disease, suggesting that HGF may be a key protein for pathogenesis of moyamoya disease ([43](#)). From this information it is apparent that the inhibition of HGF in the carotid vasculature would be advantageous. Whether it plays a role in the initiation of other cascades

remains uncertain however, it is clear that inhibiting HGF expression in the carotids could prove beneficial in the treatment of moyamoya disease.

Transforming Growth Factor- β 1

Transforming growth factor- β 1 (TGF β 1) in its original form is a 390 amino acid peptide that is proteolytically activated to form the active 112 amino acid monomeric form of the active TGF β 1 homodimer ([6](#)). Implicated in a variety of cellular processes including cell growth, proliferation, and differentiation ([45](#)), TGF β 1 is involved in the expression cascade of various connective-tissue genes at normal physiological concentrations.

Nevertheless, when it is aberrantly upregulated it has been postulated to contribute to pathological angiogenesis ([46](#)).

The upregulation of TGF β 1 has been implicated in the pathogenesis of moyamoya disease. Specifically, Hojo et al. ([46](#)) demonstrated a threefold increase in serum TGF β 1 levels of moyamoya disease patients compared to controls. Additionally, similar studies on atherosclerosis failed to demonstrate significant deviation from control results ([47](#)), speculating that TGF β 1 may play a significant role in the neovascularization process ([46](#)). Furthermore, TGF β 1 has been associated with increased production of elastin synthase, which is involved in intimal cell proliferation, a hallmark of moyamoya disease ([25](#)). Interestingly, a recent study by Liu et al. ([48](#)) failed to observe any aberrant gene expression when sequencing the first exon of TGF β 1 in both European and Japanese cohorts. In particular, they failed to demonstrate the previous association of rs1800471 and tendency toward significance of rs1800470 suggesting that although TGF β 1 may be

aberrantly expressed at the protein level this may be a result of stimulation from upstream mediators rather than mutations within the gene itself ([48](#)).

Granulocyte Colony-Stimulating Factor

Granulocyte colony-stimulating factor (G-CSF), key mediator of the acute inflammatory reaction exists as a 174 amino acid mature protein weighing 19.6 kD ([49](#)). It is a glycoprotein, growth factor, and cytokine, the main function of which is the stimulation of proliferation, survival, and maturation of neutrophil precursors and mature neutrophils. The regulation of such properties by G-CSF is via the activation of Janus kinase (JAK)/signal transducer and activator of transcription (STAT) and Ras/mitogen-activated protein kinase (MAPK) and phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt) signal transduction pathways ([50](#)). The activation of NF-kappa B is also heavily linked to the regulation of the cytokine ([51](#)).

In moyamoya disease elevated concentrations of G-CSF have been demonstrated to be as high as 1.5-fold more than controls ([52](#)).

Interestingly, expression of CSF was also observed within atherosclerotic plaques ([53](#)). As such it is unlikely to be specific to moyamoya disease however, would appear to contribute to vessel narrowing because of its action on cell recruitment ([53](#)). However, the extent to which G-CSF is involved in the pathogenesis of moyamoya disease has yet to be determined.

Extracellular Markers

Individually, prostaglandin E2 ([25](#)), IL-1 β ([25](#)), and cellular retinoic acid binding protein 1 ([54](#)) have been shown to be increased in concentration.

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Additionally, several soluble endothelial adhesion molecules have been observed in the CSF of patients with moyamoya disease. Specifically, the observation of VCAM-1, ICAM-1, and E-selectin ([23](#)).

Additionally, levels of MMP-9 were found to be significantly raised in moyamoya disease comparison to healthy controls ([55](#)). MMP-9 (gelatinase B) attenuates the impenetrability of the blood-brain barrier by interrupting the endothelial basal lamina and as a consequence plays a role in cerebral ischemia, and the formation and rupture of cerebral aneurysms, as well as other CNS pathologies.

Conclusion

There is significant debate over the pathogenesis involved in moyamoya disease. While there is obvious evidence of the involvement of inflammatory proteins, this has historically not been acknowledged as a causal factor in the condition ([5](#)). Here we have reviewed the histopathology, genetics, and signaling cascades involved in moyamoya disease identifying a number of key targets which may assist in the treatment of the condition. Although it remains uncertain to whether these factors play a role in the initiation of signaling cascades or if they are downstream mediators they clearly play significant roles in the pathogenesis. With this in mind it is important to consider these as important targets in the treatment of moyamoya disease.

In particular, significant research will have to be undertaken to fully understand the effects of each signaling molecule and at which part of the pathway they act. The distinct creation of moyamoya vessels is almost

certainly secondary to the initial stenosis observed in the ICA. By preventing this it is possible that the subsequent creation of fragile vessels could be avoided. From this review it is apparent that bFGF and G-CSF play a role in this process and could both demonstrate potential diagnostic and therapeutic relevance in the future.

Conflict of Interest Statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Kuroda S, Houkin K. Moyamoya disease: current concepts and future perspectives. *Lancet Neurol* (2008)7 (11): 1056–66. doi: 10. 1016/S1474-4422(08)70240-0

[CrossRef Full Text](#)

2. Baba T, Houkin K, Kuroda S. Novel epidemiological features of moyamoya disease. *J Neurol Neurosurg Psychiatry* (2008)79 : 900–4. doi: 10. 1136/jnnp.2007. 130666

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

3. Wakai K, Tamakoshi A, Ikezaki K, Fukui M, Kawamura T, Aoki R, et al. Epidemiological features of moyamoya disease in Japan: findings from a nationwide survey. *Clin Neurol Neurosurg* (1997)99 (Suppl 2): S1–5. doi: 10. 1016/S0303-8467(97)00031-0

<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

4. Yonekawa Y, Ogata N, Kaku Y, Taub E, Imhof HG. Moyamoya disease in Europe, past and present status. *Clin Neurol Neurosurg* (1997)99 (Suppl 2): S58-60. doi: 10. 1016/S0303-8467(97)00042-5

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

5. Smith ER, Scott RM. Moyamoya: epidemiology, presentation, and diagnosis. *Neurosurg Clin N Am* (2010)21 (3): 543-51. doi: 10. 1016/j. nec. 2010. 03. 007

[CrossRef Full Text](#)

6. Weinberg DG, Arnaout OM, Rahme RJ, Aoun SG, Batjer HH, Bendok BR. Moyamoya disease: a review of histopathology, biochemistry, and genetics. *Neurosurg Focus* (2011)30 : E20. doi: 10. 3171/2011. 3. FOCUS1151

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

7. Fukui M, Kono S, Sueishi K, Ikezaki K. Moyamoya disease. *Neuropathology* (2000)20 (Suppl): S61-4. doi: 10. 1046/j. 1440-1789. 2000. 00300. x

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

8. Takanashi J Moyamoya disease in children. *Brain Dev* (2011)33 : 229-34. doi: 10. 1016/j. braindev. 2010. 09. 003

[CrossRef Full Text](#)

<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

9. Yamashita M, Oka K, Tanaka K. Histopathology of the brain vascular network in moyamoya disease. *Stroke* (1983)14 : 50-8. doi: 10. 1161/01. STR. 14. 1. 50

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

10. Masuda J, Ogata J, Yutani C. Smooth muscle cell proliferation and localization of macrophages and T cells in the occlusive intracranial major arteries in moyamoya disease. *Stroke* (1993)24 : 1960-7. doi: 10. 1161/01. STR. 24. 12. 1960

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

11. Yamauchi T, Houkin K, Tada M, Abe H. Familial occurrence of moyamoya disease. *Clin Neurol Neurosurg* (1997)99 (Suppl 2): S162-7. doi: 10. 1016/S0303-8467(97)00054-1

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

12. Nanba R, Kuroda S, Tada M, Ishikawa T, Houkin K, Iwasaki Y. Clinical features of familial moyamoya disease. *Childs Nerv Syst* (2006)22 : 258-62. doi: 10. 1007/s00381-005-1230-5

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

13. Ikeda H, Sasaki T, Yoshimoto T, Fukui M, Arinami T. Mapping of a familial moyamoya disease gene to chromosome 3p24. 2-p26. *Am J Hum Genet* (1999)64 : 533-7. doi: 10. 1086/302243

<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

14. Inoue TK, Ikezaki K, Sasazuki T, Matsushima T, Fukui M. Linkage analysis of moyamoya disease on chromosome 6. *J Child Neurol* (2000)15 : 179-82. doi: 10. 1177/088307380001500307

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

15. Hugot JP, Laurent-Puig P, Gower-Rousseau C, Caillat-Zucman S, Beaugerie L, Dupas JL, et al. Linkage analyses of chromosome 6 loci, including HLA, in familial aggregations of Crohn disease. *G. E. T. A. I. D. Am J Med Genet* (1994)52 : 207-13. doi: 10. 1002/ajmg. 1320520216

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

16. Collins T, Read MA, Neish AS, Whitley MZ, Thanos D, Maniatis T. Transcriptional regulation of endothelial cell adhesion molecules: NF-kappa B and cytokine-inducible enhancers. *FASEB J* (1995)9 : 899-909.

[Pubmed Abstract](#) | [Pubmed Full Text](#)

17. Kamada F, Aoki Y, Narisawa A, Abe Y, Komatsuzaki S, Kikuchi A, et al. A genome-wide association study identifies RNF213 as the first Moyamoya disease gene. *J Hum Genet* (2011)56 (1): 34-40. doi: 10. 1038/jhg. 2010. 132

[CrossRef Full Text](#)

18. Liu W, Morito D, Takashima S, Mineharu Y, Kobayashi H, Hitomi T, et al. Identification of RNF213 as a susceptibility gene for moyamoya disease and

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its possible role in vascular development. *PLoS ONE* (2011)6 (7): e22542.

doi: 10. 1371/journal. pone. 0022542

[CrossRef Full Text](#)

19. Zampetaki A, Kirton JP, Xu Q. Vascular repair by endothelial progenitor cells. *Cardiovasc Res* (2008)78 (3): 413–21. doi: 10. 1093/cvr/cvn081

[CrossRef Full Text](#)

20. Rafat N, Beck GCh, Peña-Tapia PG, Schmiedek P, Vajkoczy P. Increased levels of circulating endothelial progenitor cells in patients with moyamoya disease. *Stroke* (2009)40 (2): 432–8. doi: 10. 1161/STROKEAHA. 108. 529420

[CrossRef Full Text](#)

21. Gordon N, Isler W. Childhood moyamoya disease. *Dev Med Child Neurol* (1989)31 (1): 103–7. doi: 10. 1111/j. 1469-8749. 1989. tb08418. x

[CrossRef Full Text](#)

22. Malek AM, Connors S, Robertson RL, Folkman J, Scott RM. Elevation of cerebrospinal fluid levels of basic fibroblast growth factor in moyamoya and central nervous system disorders. *Pediatr Neurosurg* (1997)27 : 182–9. doi: 10. 1159/000121249

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

23. Soriano SG, Cowan DB, Proctor MR, Scott RM. Levels of soluble adhesion molecules are elevated in the cerebrospinal fluid of children with moyamoya
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syndrome. *Neurosurgery* (2002)50 : 544–9. doi: 10. 1097/00006123-200203000-00022

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

24. Takahashi A, Sawamura Y, Houkin K, Kamiyama H, Abe H. The cerebrospinal fluid in patients with moyamoya disease (spontaneous occlusion of the circle of Willis) contains high level of basic fibroblast growth factor. *Neurosci Lett* (1993)160 (2): 214–6. doi: 10. 1016/0304-3940(93)90416-I

[CrossRef Full Text](#)

25. Yamamoto M, Aoyagi M, Tajima S, Wachi H, Fukai N, Matsushima Y, et al. Increase in elastin gene expression and protein synthesis in arterial smooth muscle cells derived from patients with moyamoya disease. *Stroke* (1997)28 (9): 1733–8. doi: 10. 1161/01. STR. 28. 9. 1733

[CrossRef Full Text](#)

26. Yamamoto M, Aoyagi M, Fukai N, Matsushima Y, Yamamoto K. Differences in cellular responses to mitogens in arterial smooth muscle cells derived from patients with moyamoya disease. *Stroke* (1998)29 (6): 1188–93. doi: 10. 1161/01. STR. 29. 6. 1188

[CrossRef Full Text](#)

27. Shinkaruk S, Bayle M, Lain G, Deleris G. Vascular endothelial cell growth factor (VEGF), an emerging target for cancer chemotherapy. *Curr Med Chem Anticancer Agents* (2003)3 : 95–117. doi: 10. 2174/1568011033353452

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

28. Sakamoto S, Kiura Y, Yamasaki F, Shibukawa M, Ohba S, Shrestha P, et al. Expression of vascular endothelial growth factor in dura mater of patients with moyamoya disease. *Neurosurg Rev* (2008)31 (1): 77–81. doi: 10. 1007/s10143-007-0102-8

[CrossRef Full Text](#)

29. Furuno A, Watari K, Nakamura M, Fukunaga Y, Jung JH, Ono M. A natural anti-inflammatory enone fatty acid inhibits angiogenesis by attenuating nuclear factor-kappaB signaling in vascular endothelial cells. *Int J Oncol* (2011)38 : 493–501. doi: 10. 3892/ijo. 2010. 856

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

30. Flister MJ, Wilber A, Hall KL, Iwata C, Miyazono K, Nisato RE, et al. Inflammation induces lymphangiogenesis through up-regulation of VEGFR-3 mediated by NF-kappaB and Prox1. *Blood* (2010)115 : 418–29. doi: 10. 1182/blood-2008-12-196840

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

31. Wang Z, Kong D, Banerjee S, Li Y, Adsay NV, Abbruzzese J, et al. Down-regulation of platelet-derived growth factor-D inhibits cell growth and
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angiogenesis through inactivation of Notch-1 and nuclear factor- κ B signaling. *Cancer Res* (2007)67 (23): 11377–85. doi: 10. 1158/0008-5472. CAN-07-2803

[CrossRef Full Text](#)

32. Park SI, Sunwoo YY, Jung YJ, Chang WC, Park MS, Chung YA, et al. Therapeutic effects of acupuncture through enhancement of functional angiogenesis and granulogenesis in rat wound healing. *Evid Based Complement Alternat Med* (2012)2012 : 464586. doi: 10. 1155/2012/464586

[CrossRef Full Text](#)

33. Takekawa Y, Umezawa T, Ueno Y, Sawada T, Kobayashi M. Pathological and immunohistochemical findings of an autopsy case of adult moyamoya disease. *Neuropathology* (2004)24 : 236–42. doi: 10. 1111/j. 1440-1789. 2004. 00550. x

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

34. Presta M, Rusnati M, Urbinati C, Tanghetti E, Statuto M, Pozzi A, et al. Basic fibroblast growth factor bound to cell substrate promotes cell adhesion, proliferation, and protease production in cultured endothelial cells. *EXS* (1992)61 : 205–9.

35. van Setten GB. Basic fibroblast growth factor in human saliva: detection and physiological implications. *Laryngoscope* (1995)105 (6): 610–2. doi: 10. 1288/00005537-199506000-00009

[CrossRef Full Text](#)

<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

36. Yamada K, Tabata Y, Yamamoto K, Miyamoto S, Nagata I, Kikuchi H, et al. Potential efficacy of basic fibroblast growth factor incorporated in biodegradable hydrogels for skull bone regeneration. *J Neurosurg* (1997)86 : 871–5. doi: 10. 3171/jns. 1997. 86. 5. 0871

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

37. Rosenkilde MM, Schwartz TW. The chemokine system – a major regulator of angiogenesis in health and disease. *APMIS* (2004)112 (7–8): 481–95. doi: 10. 1111/j. 1600-0463. 2004. apm11207-0808. x

[CrossRef Full Text](#)

38. Yoshimoto T, Houkin K, Takahashi A, Abe H. Angiogenic factors in moyamoya disease. *Stroke* (1996)27 : 2160–5. doi: 10. 1161/01. STR. 27. 12. 2160

[CrossRef Full Text](#)

39. Yan L, Wu W, Wang Z, Li C, Lu X, Duan H, et al. Comparative study of the effects of recombinant human epidermal growth factor and basic fibroblast growth factor on corneal epithelial wound healing and neovascularization in vivo and in vitro. *Ophthalmic Res* (2013)49 (3): 150–60. doi: 10. 1159/000343775

[CrossRef Full Text](#)

40. Zhang HY, Zhang X, Wang ZG, Shi HX, Wu FZ, Lin BB, et al. Exogenous basic fibroblast growth factor inhibits ER stress-induced apoptosis and
<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

improves recovery from spinal cord injury. *CNS Neurosci Ther* (2013)19 (1): 20–9. doi: 10.1111/cns.12013

[CrossRef Full Text](#)

41. Gohda E. Function and regulation of production of hepatocyte growth factor (HGF). *Nihon Yakurigaku Zasshi* (2002)119 (5): 287–94, 309. doi: 10.1254/fpj.119.287

[CrossRef Full Text](#)

42. Nakamura Y, Toyota N, Shuji D, Oda S, Namimoto T, Yamashita Y, et al. Clinical significance of the transitional phase at gadoxetate disodium-enhanced hepatic MRI for the diagnosis of hepatocellular carcinoma: preliminary results. *J Comput Assist Tomogr* (2011)35 : 723–7. doi: 10.1097/RCT.0b013e3182372c40

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

43. Nanba R, Kuroda S, Ishikawa T, Houkin K, Iwasaki Y. Increased expression of hepatocyte growth factor in cerebrospinal fluid and intracranial artery in moyamoya disease. *Stroke* (2004)35 (12): 2837–42. doi: 10.1161/01.STR.0000148237.13659.e6

[CrossRef Full Text](#)

44. Takagi Y, Kikuta K, Nozaki K, Fujimoto M, Hayashi J, Imamura H, et al. Expression of hypoxia-inducing factor-1 alpha and endoglin in intimal hyperplasia of the middle cerebral artery of patients with moyamoya
<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

disease. *Neurosurgery* (2007)60 (2): 338–45. [Discussion 345]. doi: 10.1227/01.NEU.0000249275.87310.FF

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

45. Derynck R, Jarrett JA, Chen EY, Eaton DH, Bell JR, Assoian RK, et al. Human transforming growth factor-beta complementary DNA sequence and expression in normal and transformed cells. *Nature* (1985)316 : 701–5. doi: 10.1038/316701a0

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

46. Hojo M, Hoshimaru M, Miyamoto S, Taki W, Nagata I, Asahi M, et al. Role of transforming growth factor-beta1 in the pathogenesis of moyamoya disease. *J Neurosurg* (1998)89 : 623–9. doi: 10.3171/jns.1998.89.4.0623

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

47. Grainger DJ, Metcalfe JC. A pivotal role for TGF- β in atherogenesis? *Biol Rev Camb Philos Soc* (1995)70 (4): 571–96. doi: 10.1111/j.1469-185X.1995.tb01652.x

[CrossRef Full Text](#)

48. Liu F, Yuan W, Liao D, Zhang T, Wang Z. Association of chronic hydrocephalus after aneurysmal subarachnoid hemorrhage with transforming growth factor- β 1 levels and other risk factors. *Nan Fang Yi Ke Da Xue Xue Bao* (2013)33 (3): 382–5. [Abstract].

<https://assignbuster.com/is-there-a-role-for-treating-inflammation-in-moyamoya-disease-a-review-of-histopathology-genetics-and-signaling-cascades/>

49. Zsebo KM, Cohen AM, Murdock DC, Boone TC, Inoue H, Chazin VR, et al. Recombinant human granulocyte colony stimulating factor: molecular and biological characterization. *Immunobiology* (1986)172 : 175-84. doi: 10.1016/S0171-2985(86)80097-3

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

50. Novak U, Ward AC, Hertzog PJ, Hamilton JA, Paradiso L. Aberrant activation of JAK/STAT pathway components in response to G-CSF, interferon-alpha/beta and interferon-gamma in NFS-60 cells. *Growth Factors* (1996)13 : 251-60. doi: 10.3109/08977199609003226

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

51. Okugawa S, Ota Y, Kitazawa T, Nakayama K, Yanagimoto S, Tsukada K, et al. Janus kinase 2 is involved in lipopolysaccharide-induced activation of macrophages. *Am J Physiol Cell Physiol* (2003)285 : C399-408. doi: 10.1152/ajpcell.00026.2003

[Pubmed Abstract](#) | [Pubmed Full Text](#) | [CrossRef Full Text](#)

52. Ma C, Wu F, Kong F, Zhou Y. Culture of dendritic cells in vitro and its anti-tumor immunotherapy. *Zhongguo Fei Ai Za Zhi* (2010)13 (5): 483-7. doi: 10.3779/j.issn.1009-3419.2010.05.19

[CrossRef Full Text](#)

53. Di Gregoli K, Johnson JL. Role of colony-stimulating factors in atherosclerosis. *Curr Opin Lipidol* (2012)23 (5): 412–21. doi: 10.1097/MOL.0b013e328357ca6e

[CrossRef Full Text](#)

54. Kim SK, Yoo JI, Cho BK, Hong SJ, Kim YK, Moon JA, et al. Elevation of CRABP-I in the cerebrospinal fluid of patients with Moyamoya disease. *Stroke* (2003)34 : 2835–41. doi: 10.1161/01.STR.0000100159.43123.D7

[PubMed Abstract](#) | [PubMed Full Text](#) | [CrossRef Full Text](#)

55. Fujimura M, Watanabe M, Narisawa A, Shimizu H, Tominaga T. Increased expression of serum Matrix Metalloproteinase-9 in patients with moyamoya disease. *Surg Neurol* (2009)72 : 476–80. [Discussion 480]. doi: 10.1016/j.surneu.2008.10.009

[CrossRef Full Text](#)